Strategies for Developing Oral Vaccines for Human Papillomavirus (HPV) Induced Cancer using Nanoparticle mediated Delivery System

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ABSTRACT - Human Papillomaviruses (HPV) are a diverse group of small non-enveloped DNA viruses. Some HPVs are classified as low-risk as they are very rarely associated with neoplasia or cancer in the general population, and cause lenient warts. Other HPVs are considered as high-risk types because they are responsible for several important human cancers, including cervical cancer, a large proportion of other anogenital cancers, and a growing number of head and neck cancers. Transmission of HPV occurs primarily by skin-to-skin contact. The risk of contracting genital HPV infection and cervical cancer is influenced by sexual activity. Currently two prophylactic HPV vaccines, Gardasil® (Merck, USA) and Cervarix® (GlaxoSmithKline, UK), are available and recommended for mass immunization of adolescents. However, these vaccines have limitations as they are expensive and require cold chain storage and trained personnel to administer them by injection. The use of nano or micro particulate vaccines could address most of these limitations as they are stable at room temperature, inexpensive to produce and distribute to resource poor regions, and can be administered orally without the need for adjuvants in the formulation. Also it is possible to increase the efficiency of these particulate vaccines by decorating the surface of the nano or micro particulates with suitable ligands for targeted delivery. Oral vaccines, which can be delivered using particulate formulations, have the added potential to stimulate mucosa-associated lymphoid tissue located in the digestive tract and the gut-associated lymphoid tissue, both of which are important for the induction of effective mucosal response against many viruses. In addition, oral vaccines provide the opportunity to reduce production and administration costs and are very patient compliant. This review elaborately discusses different strategies that can be pursued to develop a nano or micro particulate oral vaccine for HPV induced cancers and other diseases.

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INTRODUCTION

All papillomaviruses contain a double-stranded, circular DNA genome approximately 8 kb in size and normally contain eight sets of genes [1-4]. Papillomavirus types found in humans (Human papillomaviruses, HPV) are divided into five genera based on DNA sequence, and they have different life-cycle characteristics and disease associations. Among these five types, two of them, Alpha and Beta, contain about 90% of currently characterized **HPVs** The Alpha [5-7]. papillomavirus type is the largest group of HPVs and includes the genital/mucosal HPV types and cutaneous viruses such as HPV2. This group causes common warts and is rarely associated with cancers [8]. Whereas, Beta papillomaviruses are typically associated with non-apparent cutaneous infections in humans and also associated with the development of non-melanoma skin cancer [9, 10]. It is now clear that most HPV types, including the majority of those within the Alpha and Beta genera, cause only asymptomatic infections in immune-competent individuals and can be detected in skin swabs. Another type, Gamma also causes asymptomatic infections [11-14]. These Gamma type viruses are able to easily adapt themselves in the host, complete their life-cycle and maintain population growth, causing diseases [15, 16]. It has been found that mostly the Beta class of HPVs are involved in cancer.

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Papilloma viruses replicate and assemble exclusively in the nucleus of keratinocytes. The viruses infect keratinocytes in the basal layers of stratified epithelia and replicate in infected keratinocytes in a differentiation-dependent manner. The viral gene expression and replication proceed in a tightly controlled fashion regulated by the keratinocyte differentiation [17]. To date, more than one hundred HPV genotypes have been completely sequenced. Of which, certain types of HPVs, such HPV16, HPV18, and HPV31, are considered as high-risk or oncogenic and are frequently detected in cervical and other genital cancers. A characteristic of the infection caused by these HPVs is that viral genomes are commonly found integrated into the cancer cell genome. Other types of HPVs, such as low-risk or non-oncogenic HPV6 and HPV11, which induce benign genital warts, are very rarely found in genital malignancies [18]. The high risk HPV 16 and 18 are known to be responsible for 70% of cervical cancers worldwide, whereas HPV 6 and 11 are the predominant lowrisk types that cause genital warts and recurrent respiratory papillomatosis (RRP) [19]. HPVcaused human cervical cancer results in the second largest of cancer related deaths of women in the world. In the United States, it became the center of attention when a study showed that 25% of persons between the ages of 14 and 19 and 45% of persons between the ages of 20 and 24 were HPV positive. It is estimated that more than 80% of both men and women in the United States will be infected with HPV at some point in their lives [20, 21]. Estimated yearly cervical cancer cases and deaths associated with HPV are 490,000 and 270,000, respectively [20, 21]. Sexually transmitted HPV is a necessary factor for the development of cervical cancer and its precursor lesions. Cervical HPV infection is found in 5-40 percent of asymptomatic women of reproductive age [22]. Risk of infection increases with increased number of sexual partners, starting sexual intercourse at a younger age, and recent acquisition of new partners [23]. Although the vast majority of these infections are transient, a substantial increase in risk of cervical neoplasia exists for women who develop persistent, longterm infections with oncogenic HPV types [23-26]. Currently, two prophylactic HPV vaccines, Gardasil (Merck, USA) and Cervarix (GlaxoSmithKline, UK) are available and recommended for mass immunization of preteen girls and boys at age 11 to 12 years. Recently the CDC also recommended the HPV vaccines for teen boys and girls who did not get the vaccine when they were younger; including teen girls and young women through age 26, and teen boys and young men through age 21.

Both Gardasil and Cervarix vaccines consist of the immunogenic proteins L1, which are the major proteins of the capsid of papillomavirus. L1 proteins self-assemble into 'virus-like particles' (VLPs) when expressed at high levels in cultured cells. VLPs are multi-protein structures that mimic the organization and conformation of authentic naïve viruses but do not contain any genetic material. When administered, VLPs are able to generate immunity as if the immune system has been confronted with a real virus [27]. However, because VLPs do not contain any genetic material, they are unable to replicate and as such are harmless and safe. Cervarix vaccine contains VLPs of HPV types 16 and 18, whereas Gardasil includes additional VLPs of HPV types 6 and 11 [28]. The VLPs used in Gardasil are manufactured from Saccharomyces cerevisiae (bread yeast) which is transfected with the genes expressing L1 whereas Cervarix VLPs are manufactured from Trichoplusiani insect cell line that was infected with L1 encoding recombinant baculovirus. Gardasil and Cervarix have 3 years and 4 years shelf life respectively, and are stored at 2 to 8° C. The approximate cost for three doses is about \$400 to \$500 [29]. The adjuvant in Gardasil is amorphous aluminum hydroxyphosphate sulfate (AAHS) (225 µg), whereas aluminum hydroxide (500 µg) and 3-O-deacyl-4' monophosphoryl lipid A (50 μ g) are the adjuvants in Cervarix [30]. Both vaccines are administered intramuscularly in a volume of 0.5 mL and require multiple doses. After the administration of the prime dose, two more booster doses are administered, one within 1 to 2 months and another within 6 months. While both Gardasil and Cervarix are shining examples of bench to bedside research, these vaccines have significant drawbacks that limit their applications in the settings where they are most needed. They are expensive, and require cold chain storage and trained personnel to administer them by injection. In addition, there is a growing concern regarding their adverse effects. Patients receiving Gardasil and Cervarix may experience pain, fatigue, redness, swelling, fever, GI symptoms (diarrhea, nausea, vomiting), headache, dizziness, myalgia and arthralgia [31-35]. The most common adverse

effect is injection-related local reaction, such as pain, swelling and erythema with a rate of 95% of light to moderate intensity [32, 33]. Severe adverse effect, such as severe headache with hypertension, gastroenteritis and bronchospasm, were also noted [32]. There are more data available on adverse effect associated with Gardasil than Cervarix; however, the major adverse effect for the latter vaccine is also injection-related local pain (78%) [32]. Other disorders that Gardsil may cause include infection and infestation (52%), gastrointestinal disorder (13.4%), nervous system disorder (9.4%), and reproductive and breast disorders (24.8%). Darja Kanduc has shown that HPV 16 antigen can induce autoimmune reaction against human proteins which might lead to pathologies such as spinal muscular atrophy, proximal muscle weakness that cause maddling gait, toe-weakening, lordosis, frequent falls, difficulty in standing up and climbing stairs, cardiovascular and musculskeletal abnormalities, disorder of lipoprotein metabolism leading to hypercholesterolemia, and increased proneness to coronary artery disease [36]. The aluminium adjuvant in both vaccines has also been shown to cause adverse effects. Stephanie Seneff has shown that children may not react acutely to the aluminum adjuvated vaccine, which can lead to neural damage that is partly mediated by exuberant production of nitric oxide [37].

Thus it is very important to develop an affordable, safe and highly effective HPV vaccine to fight virus induced diseases globally. To develop a highly efficient and cost effective vaccine, several factors must be taken into consideration. The vaccine formulation must be safe and easy to administer. The vaccine should address the issue of adverse effect. The vaccine formulation should also be capable of eliciting the desired immune responses, humoral and/or cellular mediated. An ideal vaccine should require least number of doses without the need for a booster dose(s). It is also important that all components of the vaccine are commercially available, safe, affordable and nontoxic. The process of vaccine manufacturing should be easy, affordable, and amenable to other steps of preparation such as sterilization. lyophilization, spray drying or vacuum drying, packaging and reconstitution of the dried powder. The vaccine formulation should be stable with respect to size, surface morphology and size distribution throughout the process of preparation, storage and administration. Moreover, the antigen has to be chemically and physically stable throughout the process of antigen loading and there should be no premature release/leakage of antigen. To address the issues with the currently available HPV vaccines and to prepare a highly efficient and cost effective alternative vaccine based on the aforementioned criteria, a particulate formulation of nano or micron size is considered to be the most desirable candidate. Nano or micro particulate formulations of vaccines have the potential to overcome the limitations of currently available vaccines as the nano or micro particulate formulations may be stable at room temperature and can be administered orally. In this review, we discuss the different aspects of developing a highly efficient, stable and cost effective HPV vaccine using nano/micro particle for oral administration.

VLP-BASED HPV VACCINES

The first vaccination against HPV was demonstrated by Shope in 1937 where neutralizing antibodies protected rabbits against high-dose viral challenge with cottontail rabbits papilloma virus (CRPV). The study found that generating serum neutralizing antibody to the virus capsid protein is an effective strategy for prophylactic vaccination against the infection [38]. The currently available HPV vaccines, Gardasil and Cervarix, are subunit vaccines consisting of VLPs assembled from the major L1 proteins of HPV type 16, 18, 6 and 11 (Gardasil) and HPV type 16 and 18 (Cervarix). As the VLPs have no genetic material in them and cannot grow or cause any infection, there are huge opportunities for using VLPs as antigens in viruscausing diseases that are hard to cure. There are numerous advantages of using VLPs as antigens in formulation. VLPs vaccine are excellent prophylactics because they are self-assembling bio-nanoparticles (20 to 60 nm in diameter) that expose multiple epitopes on their surface and very accurately mimic the native virions [39]. In addition, VLPs are superior to bacterial vaccines and viral antigens in a number of ways and bacterial antigens can sometimes revert to the virulent form [40, 41]. On the other hand, in the case of viral antigens, the authentic and attenuated virions cannot be used as antigens in a prophylactic vaccine because they would contain oncogenic viral genomes that would be infectious [42]. A VLP has no such side effects and can eliminate these risks. VLPs not only resemble authentic

virions morphologically, but they also mimic virions immunologically, which means that they are able to induce high titers of neutralizing antibodies to conformational epitopes when vaccinated [43, 44]. The surface of VLPs consists of an array of antigenic epitopes that mimic the surface of native virions more reliably than specific isolated subunits or subcomponents of the virus [43]. VLPs can be produced in either a prokaryotic or eukaryotic cells by expressing the protein in a different medium such as mammalian cells, insect cells, yeast, or even bacteria [45].

Both Gardasil and Cervarix vaccines induce the generation of high concentrations of neutralizing antibodies to L1 and have been shown to be highly efficacious in randomized and controlled trials. Instead of flagging the antigen, the antibodies are able to neutralize the biological effect of the antigen. It has also been shown that the neutralizing antibodies cause cell-mediated cytotoxicity to the virus [46]. Mechanistic studies of the HPV infection revealed that the virus first causes microabrasion and removal of the full thickness of the epithelium but keeps the epithelium basement membrane intact since the virion attaches first to the basement membrane before entering basal cells. In the epithelial basement membrane, the virus binds to the heparin sulfate proteoglycans via L1 protein. The virus capsid then undergoes a conformational change and allows the exposure of L2 protein that binds to the surface molecules of keratinocytes. The capsid then undergoes further conformational changes, leading to the exposure of cellular receptor binding sites on L1 protein. Subsequently, the virus binds to cellular receptor via L1 protein and enters the cell [47]. Following HPV L1 VLP immunization, antibodies are produced which prevent both initial binding of HPV virus to the basement membrane and binding of the virus to the keratinocyte cell surface [48]. It has been shown that the antibodies to L1 are effective at very low concentrations, consistent with data from the animal papilloma virus model and from natural infections in humans [49, 50]. The virus-like-particle can be produced by expressing the specific HPV protein in eukaryotic cells. VLPs in Gardasil are produced in yeast cells by cell disruption and purified by a series of chemical and physical methods. Cervarix VLP, on the other hand, is produced in insect cells. Asghar et al have also reported the production of recombinant HPV-16L1 protein in Eukaryotic Sf9

insect cells. The recombinant protein L1 was predominantly ~ 60 kD indentified by western blot analysis. VLP formation was confirmed by SDS-PAGE with distinct immunoreactivity in western blot analysis and electron microscopy [51]. The HPV infection causes several

several manifestations. including common warts. epidermodysplasia verruciformis, anogenital warts, cervical and vulvar cancer of the penis, vagina and anus, and recurrent respiratory papillomatosis [52]. Although HPV is asymptomatic and auto-limited, it is a public health concern because of its association with genital tract malignant disease among men and women [53]. HPV genital infection is mainly transmitted by genital-to-genital contact often during sexual intercourse. Both of the currently available HPV vaccines are VLP-based vaccines. They are classified as the quadrivalent HPV vaccine (Gardasil) and the oncogenic HPV bivalent vaccine (Cervarix) [54]. Clinical studies have shown that the quadrivalent vaccine offers protection against persistent HPV infection; cervical, vaginal and vulvar lesions that are precursors for cancer; and genital warts caused by HPV types 6, 11, 16 or 18 in women aged 16 to 26 years old who were not previously infected by these HPV types [54]. On the other hand, the bivalent vaccine contains only VLPs of oncogenic HPV types 16 and 18. Cancer vaccines are medicines that belong to a class of substances known as biological response modifiers. Biological response modifiers work by stimulating or restoring the immune system's ability to fight infections and disease. There are two broad types of cancer vaccines: preventive (or prophylactic) vaccines, which are intended to prevent cancer from developing in healthy people; and treatment (or therapeutic) vaccines, which are intended to treat an existing cancer bv strengthening the body's natural defenses against the cancer. Cancer preventive vaccines target infectious agents that cause or contribute to the development of cancer. They are similar to traditional vaccines, which help prevent infectious diseases, such as measles or polio, by protecting the body against infection. Both cancer preventive vaccines and therapeutic vaccines are based on antigens that are carried by infectious agents and that are relatively easy for the immune system to recognize as foreign. The current HPV vaccines, Cervarix and Gardasil, are prophylactic vaccines designed to reduce the occurrence of cervical

cancer. Although both vaccines have been proven to be highly effective, the limitation of these vaccines is their cost; they are expensive in terms of preparation and preservation. In addition, the administration of these vaccine requires trained personnel. Due to these limitations, mass application of these vaccines around the world is severely hindered. The ultimate goal of vaccination is to ensure the production of strong and lasting immune responses after a single dose of antigen without the need for booster doses [55, 56]. In order to ensure the quality and quantity of the immune response, it is highly important that the immune system is presented with antigens at the right location of the targeted pathogen in sufficient amount [57, 58]. Nano or micro particulate formulations of vaccines have the potential to address most, if not all, of these limitations as they may be stable at room temperature, inexpensive to produce, more effective as a particulate carrier, and can be administered orally. A biodegradable polymer based particulate vaccine can act as an adjuvant itself, therefore there is no need for using salt based adjuvants, which will eliminate the adverse effects caused by adjuvants.

In addition, it is also possible to increase the efficiency of the particulate vaccine by adding appropriate ligands, charged particle or any other biocompatible chemical to increase the specificity of the nano or micro particles for targeted delivery [59]. Human papilloma virus (HPV)-16 is the prevalent genotype associated with cervical tumours. Virus-like-particle-based vaccines have proven to be effective in limiting new infections of high-risk HPVs, but the high cost has hampered their use, especially in poor developing countries. Avipox-based recombinants are replicationrestricted to avian species and represent efficient and safe vectors for immunocompromised hosts. These recombinants can elicit a complete immune response. A new fowlpox virus recombinant encoding HPV-L1 (FPL1) was engineered and evaluated side-by-side with a FP recombinant coexpressing L1 and green fluorescent protein (FPL1GFP). This fowlpox virus recombinant correctly express the L1 in vitro in different cell lines which was confirmed by western blot, immunofluorescence, real-time PCR, and electron microscopy. Mice were also immunized to determine its immunogenicity. It was also demonstrated that the FPL1 recombinant better expresses L1 in the absence of GFP, correctly assembles structured capsomers into virus-like particles (VLPs), and elicits an immune response in a preclinical animal model. Thus far this is the first report of HPV VLPs assembled in eukaryotic cells using an avipox recombinant [59].

OTHER HPV VACCINES

Particulate vaccine can deliver a wide variety of antigens such as attenuated, killed or inactivated pathogens, recombinant protein, peptides from oncogenic protein. synthetic peptide. carbohydrates, lipids and DNA. Biodegradable polymer based nanoparticle can be used as a suitable carrier for the development of effective and affordable DNA and protein subunit vaccines. development Rational of such vaccine formulations requires a detailed understanding of physico-chemical properties, cell-free their environment and in vitro behavior. Also it is necessary to understand the process of particle uptake and processing mechanisms of antigen presenting cells (APC), which are capable of stimulating safe and effective immune responses.

One effective vaccine is peptide vaccine, which offers several advantages over classical vaccines. However, peptides alone are not immunogenic and need a delivery system that can boost their recognition by the immune system. In recent years, nanotechnology-based approaches have become one of the most promising strategies in peptide vaccine delivery [60]. In case of HPV vaccine, peptides can be obtained from the Human Papillomavirus (HPV) E6 and E7 oncogenes and can be an effective antigen to develop a therapeutic vaccine for HPV induced cancers. These peptide sequences derived from the oncogenic E6 and E7 viral proteins have been shown to represent suitable tumor associated antigens (TAAs) for cervical cancer and are considered as ideal candidates for developing therapeutic vaccines [61-63]. These peptides are easily recognized by CD8 T lymphocytes, which are the most effective components of the adaptive immune system capable of recognizing and destroying viralinfected and transformed malignant cells [64-66]. In addition to peptides obtained from the E6 and E7 viral proteins, synthetic peptides representing these TAAs have also been tested in numerous ways in human patients and mouse cancer models for their ability to generate anti-tumor T cell responses capable of exhibiting anti-tumor effects [67-69]. However, a significant limitation observed of these

antigens is that they show only a modest T cell responses capable of dealing with very early disease stages. Therefore it is necessary to develop improved peptide-based immunization strategies which will have significant impact against advanced disease stages. Kelly Barrios et al has developed a synthetic peptide vaccination strategy, called TriVax, that is effective in generating vast numbers of antigen-specific T cells in mice capable of persisting for long period of time [70]. They have described an improved peptide vaccination strategy in mice that shows a significant immune response involving a large number of CD8 T cells [71, 72]. The vaccine TriVax, using HPV16-E749-57, induced large and persistent T cell responses that were therapeutically effective against established HPV16-E7 expressing tumors. In most cases, TriVax was successful in acting against 6-11 day old tumors. In addition, TriVax induced long-term immunological memory, which prevented tumor recurrences. The TriVax vaccine consists of a synthetic peptide corresponding to the minimal T cell epitope, poly-IC adjuvant and costimulatory monoclonal anti-CD40 antibodies (aCD40 mAb), which are mixed together and administered intravenously.

More recently, Rahimian has attempted to develop a HPV cancer vaccine formulation [73]. Synthetic long peptides (SLPs) derived from HPV16 E6 and E7 oncoproteins have been used for therapeutic vaccination. In preclinical and clinical studies, adjuvants based on mineral oils (such as incomplete Freund's adiuvant (IFA) and Montanide) are used to create a sustained release depot at the injection site. While the depot effect of mineral oils is important for induction of robust immune responses, their administration is associated with severe and long lasting adverse effects. In order to develop an alternative to mineral oil based vaccine, polymeric nanoparticles (NPs) based on hydrophilic polyester (poly(d,l lactic-co-hydroxymethyl glycolic acid) (pLHMGA)) were prepared. These NPs were loaded with a synthetic long peptide (SLP) derived from HPV16 E7 oncoprotein and a Toll like receptor 3 (TLR3) ligand (poly IC) by double emulsion solvent evaporation technique. The therapeutic efficacy of the nanoparticulate formulations, was compared to that of HPV SLP+poly IC formulated in incomplete Freund's adjuvant (IFA). The results showed that the encapsulation of HPV SLP antigen in NPs substantially enhanced the population of HPVspecific CD8+ T cells when combined with poly IC either co-encapsulated with the antigen or in its soluble form. Although the therapeutic efficacy of NPs containing poly IC in tumor eradication was equivalent to that of the IFA formulation, the administration of pLHMGA nanoparticles was not associated with adverse effects and therefore these biodegradable nanoparticles are excellent substitutes for IFA in cancer vaccines.

A DNA vaccine can be another alternative to fight the HPV infection. DNA vaccines have emerged as an attractive approach for antigenspecific T cell-mediated immunotherapy to combat cancers. In HPV infection, two oncogenic proteins, E6 and E7, are consistently co-expressed in HPVexpressing cervical cancers and are important in the induction and maintenance of cellular transformation. Therefore, immunotherapy targeting E6 and/or E7 proteins may provide an opportunity to prevent and treat HPV-associated cervical malignancies. Chien et al has shown that a DNA vaccine can be effectively used against HPV infection [74]. In the case of HPV, T cell-mediated immunity is one of the most crucial components in our defense against HPV infections and HPVassociated lesions. Therefore, the goal of DNAbased vaccination is to generate strong E6/E7specific T cell-mediated immune responses. Intradermal administration of DNA vaccines via a gene gun represents an efficient way to deliver DNA vaccines into professional antigen-presenting cells in vivo. Professional antigen-presenting cells. such as dendritic cells, are the most effective cells for priming antigen-specific T cells. Using the gene gun delivery system, several DNA vaccines that employ intracellular targeting strategies for enhancing MHC class I and class II presentation of encoded model antigen HPV-16 E7 were tested. Furthermore, a strategy to prolong the life of DCs to enhance DNA vaccine potency was developed. More recently, a strategy to generate antigenspecific CD4+ T cell immune responses to further enhance DNA vaccine potency was also developed. The impressive preclinical data generated from ourthese studies have led to several HPV DNA vaccine clinical trials

A live bacterial-based HPV vaccine can be another choice to fight HPV infections. Bacterialbased vaccines are inexpensive and feasible to prepare in a regular laboratory set up. Yan et al evaluated the potential value of live attenuated Shigella. flexneri 2a sc602 strain-based HPV16L1 as a high-efficiency, low-cost HPV16L1 mucosal vaccine [75]. The study revealed that the recombinant sc602/L1 vaccine induced high L1specific systemic and mucosal immune responses as well as cell-mediated Th1 and Th2 immune responses in guinea pig model. Sc602/L1 vaccine induced higher L1-specific IgG and IgA antibodies as well as HPV16-neutralizing antibodies in genital region in sc602/L1 mucosal immunized animals than in L1 intramuscular immunized animals. Though both are via mucosal delivery, immunized sc602/L1 vaccine by rectum route induced higher L1-specific IgA and IgG titers in genital region than by conjunctiva route. In addition, sc602/L1 also strongly increased L1-specific IFN-y and IL-4 expression, implying its effect on cell-mediated immune response. The study proves that sc602/L1 bacterial-based vaccine may have a significant protective effect against HPV infection.

NANO-PARTICULATE VACCINES

Emerging nanotechnology in medical science has provided an unparalleled opportunity to advance the treatment of various severe diseases. Nanoparticles exhibits several unique properties, such as higher surface-to-volume ratio, small size, ability to encapsulate various drugs, and tunable surface chemistry, which give them many advantages over their bulk counterpart. These advantages include multivalent surface modification with targeting ligands, efficient navigation of the complex in an in vivo environment, increased intracellular trafficking, addition of any charged particles to increase target selectivity and sustained release of drug. These advantages make nanoparticles an ideal candidate for formulating drugs for most prevalent and challenging diseases including cancer [76]. Nanoparticulate drug carriers are passively targeted to cancer tumors using the enhanced permeability and retention effect (EPR) in tumor area, thus they are the most suitable contestant for the delivery of chemotherapeutics in cancer treatment. In fact, advances in nanomedicine have rapidly made possible some of these drugs to be used in clinical practice. To date, there are five nanoparticle chemotherapeutics that have been approved for cancer treatment and many more are under clinical investigation [77].

In addition, to their therapeutic use, nanoparticles can also be useful as a new strategy

for vaccine development. Current successful vaccines are live, attenuated, killed or fragmented pathogens. However, due to their complex nature, such vaccines can vary in quality from batch to batch and can induce adverse effects such as those associated with the whole pertussis, Sabin polio, measles, respiratory syncytial virus (RSV) or rotavirus vaccines [78-81]. A particulate formulation has huge potential in vaccine development as the particle can be used as antigen career and/or adjuvant and can address the issue of adverse effects that are caused by conventional vaccines. New vaccine strategies can take advantage of particulate compounds - especially nanoparticles - to target antigen presenting cells more efficiently [82]. Particulate formulations offer a number of advantages in vaccine development. Particulate carriers can serve as an effective antigen delivery system that is able to enhance and/or facilitate the uptake of antigens by antigen-presenting cells (APCs) such as dendritic cells (DCs) or macrophages [83, 84]. Particlebased antigen carriers can also serve as a depot for controlled release of antigen, thereby increasing the availability of antigen to the immune cells. It has been found that antigen release may enhance not only the level of the immune response but also its quality [85, 86]. In addition, particle-based adjuvants possess the ability to modulate the type of induced immune responses when used alone or in combination with other immune-stimulatory compounds [87]. Particulates have the ability to protect the integrity of antigens against degradation until delivered to the immune cells [88]. This is particularly important in oral vaccine formulations where antigens must be protected from the harsh acidic conditions of the stomach and enzymatic degradation in the GI tract [89]. More importantly, particulate vaccines can potentially cross-present the antigen; antigen cross-presentation is especially important to generate CD8+ T-cell responses against viral infections [90, 91]. Another advantage of using particulate formulation of a vaccine is that it can eliminate the use of adjuvants which do not have much immunogenic effect. For example, the vaccine Cervarix contains both aluminium hvdroxide ASO4 (3-O-desacyl-4and monophosphoryl lipid A) as adjuvants. Theses adjuvants only improve humoral immunity but do not contribute to cell mediated immune response, the main immune function of the VLP [92, 93]. The immunologic effect of particulate vaccines is

related to the size, stability, antigen loading and antigen-release kinetic properties of the particle [94]. The immune response is also influenced by particle interaction with APCs and antigen presentation and processing by APCs [95]. Several synthetic polymers are used in the preparation of the particles such as poly (lactic acid) (PLA), poly(ortho esters), poly(lactic-co-glycolic acid) polyethylene (PLGA), glycol. and polyphosphazene. Natural polymers such as albumin, gelatin, collagen, chitosan and alginate have also been used in vaccine candidate formulations [96-99]. When compared to micro particles, nano sized particles offer more options as the surface ratio is higher when the size is downgraded to the nano scale [100]. Advantages of nanoparticle-based delivery of vaccines/drugs, include improved biological stability of antigen/drug and efficacy in targeting APCs for induction of innate and adaptive immunity due to Class I and Class II presentations [101]. Nanoparticles may also provide enhanced intracellular concentrations, controlled release of vaccine antigen/drug, and reduced number of administrations due to enhanced immune response.

One of the most significant advantages of nano size particle is that the particle can act as immunestimulating adjuvant. Gamvrellis, et al. have shown that a nano-particulate antigen delivery system was able to induce a substantial immune response without inducing any inflammation. The antigen appears to induce substantial immune responses in mice and sheep without adding stimulators such as toll-like receptors or other pathogen recognition receptors [102]. Nano particulate formulations can also be used to develop a safe and effective cancer vaccine formulations. Poly (d, l-lactic-co-glycolic acid) nanoparticles (PLGA-NPs) can be used to formulate a cancer vaccine delivery system which has potential in the development of future therapeutic cancer vaccines [103]. This nano particle can target dendritic cells (DCs) which can effectively initiate antitumor activity. The PLGA nano particle containing antigens along with immune-stimulatory molecules (adjuvants) can not only target antigen actively to DCs, but also provide immune activation and rescue impaired DCs from tumor-induced immuo-supression [104]. The authors further assessed the extent of maturation of DCs after treatment with the antigen, monophosphoryl lipid А (MPLA), and encapsulated PLGA nanoparticles. The generation of primary T-cell immune responses elicited by DCs was monitored. Results showed that the high amounts of pro-inflammatory and TH1 (T helper 1) polarizing cytokines and chemokines released by the nanoparticles are greater than that achieved by MPLA in solution [105]. These results confirmed that the nanoparticle formulation of a vaccine is more immunogenic when compared to the solution form of the antigen. Dendritic cells in peripheral tissues are important as they act as sentinels of the immune system, detect and capture pathogens entering the body, and present their antigens to T cells to trigger responses directed towards the elimination of the pathogen. Diwan et.al. investigated the formulation of a pharmaceutically biodegradable, acceptable, and strategic nanoparticulate delivery system and its application for efficient antigen loading of DCs to achieve antigen specific T cell activation. The results of the investigation indicated that PLGA nanoparticles are able to mimic certain features of pathogens and can efficiently act as delivery systems for targeting vaccine antigens to DCs and activating potent T cell responses [105].

In the intestine, particles are readily phagocytized by the antigen presenting cells, mostly microfold cells (M cells) which are present in the underlying region of the Peyer's patches of the small intestine [106-108]. The first step of mucosal immune response is the trans-epithelial transport of antigens and pathogens. The antigen then reaches the site of immune response. The delivery of antigens across the epithelial barrier to the underlying lymphoid tissue is mediated by M cells, a specialized epithelial cell type that occurs only in lymphoid follicle associated epithelium. Particulate formulation of vaccines where antigens are coupled to or encapsulated are found to be transported through the epithelial layer by M cells more efficiently than live, attenuated, killed pathogens or antigens. In case of particulate vaccine, it is also possible to enhance the binding capacity, target ability and uptake of the particle by adding ligand, charges particle at the surface. Such modification leads the particulate vaccine to the receptors on the M cell surface [109]. These M cells then transport the particles to the macrophages or other cells underlying the gutassociated lymphoid tissues [110, 111]. Thus, Mcell targeting lectins, such as Ulex Europaeus Agglutinin (UEA-1), Aleuria Aurantia Lectin (AAL), and Wheat Germ Agglutinin (WGA), can

increase cellular uptake and efficiency of particulate vaccine and can be added to the formulation to enhance targeting of the Peyer's patches. Further immune-stimulatory cytokines such as IL-1 and IL-12 can be added for enhanced immunity. Since the antigen is presented in a particulate formulation, there is no need for added adjuvants due to the sustained nature of antigen release from the particles. Several technologies for the oral administration of drugs/vaccines using nanoparticles, microparticles (microspheres), and a biodegradable polymer-based number of microparticle formulations have been studied as [108. effective deliverv systems 112]. Biodegradable and biocompatible polymers, copolymers and lipids have been used to prepare nano/micro-particle as vaccine-delivery systems [113-115]. The material is selected based on factors. including biocompatibility, several degradation rate, hydrophilicity or lipophilicity, surface charge, and polarity. Examples of nanoparticle-based vaccines include oral biodegradable microspheres with recombinant anthrax vaccine for immunization against anthrax infection, poly (DL-lactide-co-glycolide (DL-PLG) microspheres encapsulating phosphorylcholine Salmonella against typhimurium, and albumin-chitosan mixed matrix microsphere-filled coated capsule formulation of the typhoid vaccine [114-116].

POTENTIAL BENEFITS OF AN ORAL NANO/MICRO-PARTICULATE HPV VACCINE

Oral administration is the most preferred route for drug delivery as it is most patient compliant. Therefore developing an oral HPV vaccine with high efficiency and low cost will eliminate the limitations that the current vaccines have. The oral route will also eliminate the need for a trained personnel to administer the vaccine, which will give the vaccine a more global character as it will be easily available and applicable in resource poor countries. In addition, oral vaccines have the potential to stimulate mucosa-associated lymphoid tissue (MALT) located in the digestive tract and the gut-associated lymphoid tissue (GALT). Both of these tissues are important for the induction of an effective mucosal response against many viruses [117]. For Human papillomavirus, an elevated mucosal immune response could serve as a first line of protection against the infection. Alternative immunization routes for HPV other than the intramuscular route of administration have been investigated. The oral delivery of vaccines as an alternative immunization route and the efficiency of mucosal immunization for different antigens have been described [118]. In addition the intranasal route of administration for vaccine delivery has been investigated. Results from studies of both oral and intranasal routes of administration show the potential of mucosal immunization with HPV VLPs for inducing a neutralizing antibody response and L1-specific cytotoxic T-lymphocytes [118, 119].

Several animal studies were conducted using HPV L1 VLPs or different assembly forms (capsomeres) in the form of a solution, or in edible products (HPV L1 VLP expressed in potato) [120]. Recombinant clones of attenuated Salmonella enterica (Serovar Typhi and Typhimurium) strains expressing HPV-16 and HPV-18 L1 antigens were also shown to induce a strong immune response and are currently in the pre-clinical testing phase for oral or mucosal administration [121]. However, oral vaccine formulations without adjuvants have thus far required large amounts of antigens compared to the intramuscular route when delivered in solution form. Nano or micron sized particles may provide enhanced intracellular concentrations, controlled release of antigens, stability, and a reduced number of administrations. It has been shown that microparticles prepared from biodegradable polymer can be easily prepared, well characterized, administered orally and be a reliable career of variety of drugs and vaccine antigen such as oligonucleotide antisense to NF-kB, plasmid DNA encoding hepatitis-B surface antigen [122, 123]. Studies have shown that microparticulated formulation increases the bioavailability of orally administered antisense. Antisense drug is considered as next generation drug due to their specific targeting ability to mRNA and minimum toxic effect. However, the drug has poor biological stability, short half-life and limited cellular uptake [124]. The bioavailability of antisense solution via oral administration was only 9%, whereas the bioavailability of the antisense encapsulated in bovine serum albumin increased up to 70% [125]. The adjuvant-like properties of the nanoparticles also enhance the immune response due to their ability to target Peyer's Patches and M cells (microfold cells). Peyer's Patches are aggregations of lymphoid tissue normally found in

the lowest portion of the small intestine and are full of M cells. To evoke the mucosal immune response, antigens on the mucosal surface must first cross the impermeable epithelial barrier into lymphoid structures such as Peyer's Patches where the M cells take up the antigen and then process it and present it to the antigen cells such as macrophages and dendritic cells. This process, called antigen transcytosis, is mediated mainly by M cells [126]. A potential problem with a vaccine's efficacy is the lack of the vaccine's targeting ability. Thus, addition of any targeting ligand to the particle or modification of the particles in a way that they can target M cells in Peyer's Patches enhances the vaccine's efficiency. One such modification can be done with Chitosan. Chitosan is a positively charged polysaccharide that can be used to provide a positive charge to the surface of the vaccine particle which will then enhances its ability to target M cells, as the surface of M cells is negatively charged. Also a ligand such as AAL (aurantia aleuria lectin), which is very specific to some receptors at the surface of M cell layers, can increase the targeting ability of the vaccine. In addition, cytokines such as IL-2 and IL-12 are able to enhance the immune response. The combination of all of these strategies can produce a more efficient vaccine as well as circumvent the issues with the current intramuscularly administered VLP solution vaccines.

CONCLUSION

HPV-caused cancers and diseases remain an important health concern in the United States and throughout the world. Thus far there are only two vaccines available for HPV related cancers and other diseases. These vaccines are proved to be highly efficient in terms of preventing diseases. however, they have several disadvantages which limit their use, particularly in resource-poor countries. Furthermore, the side effects of these intramuscular vaccines are raising important health questions. Therefore, there is a great need for developing a new alternative HPV vaccine which will be cost effective and can significantly contribute to global public health. There are a number of choices as alternatives to the current VLP- based HPV vaccines, such as a DNA based vaccine, peptides from HPV oncogenic protein, synthetic peptides, and live bacteria. The problem with these vaccines alternatives is that their delivery system is inefficient. A biodegradable polymer based nanoparticle is the most suitable formulation to address this delivery issue. Evidence suggests that it is possible to develop an alternative oral HPV vaccine using a nano or micro particulate formulation that promises to be highly efficient, more cost effective, and more patient compliant than the existing formulations. Studies have shown that VLPs, which are multi-protein structures that mimic the exact organization and conformation of native viruses but lack the viral genome, are perfect for preparing safer and cheaper vaccine candidates. In addition, studies have also revealed that smaller size VLPs such as nano or micron size offer numerous advantages in vaccine development when compared to solution form. VLP technology Thus. combining with biodegradable polymer based nano particulate formulations appears to be a very promising new approach for the future development of desirable oral HPV vaccines.

REFERENCES

- 1. Monie A, Tsen S-W, Hung C-F and Wu T-C. Therapeutic HPV DNA vaccines. Exper Rev Vaccines, 2009; 8(9): 1221-1235.
- Nomura K, Ogawa T, Sugawara M, Honkura Y, Oshima H, Arakawa K, Oshima T and Katori Y. Association between Septal Deviation and Sinonasal Papilloma, Tohoku J. Exp. Med., 2013; 231: 315-319.
- Bernard HU, Burk RD, Chen Z, van Doorslaer K, zur Hausen H, and de Villiers E-M. Classification of Papillomaviruses (PVs) Based on 189 PV Types and Proposal of Taxonomic Amendments. Virology 2010; 401 (1): 70–79.
- 4. Bravo IG, de Sanjosé S, Gottschling M. The clinical importance of understanding the evolution of papillomaviruses. Trends Microbiol. 2010; 18(10):432-438.
- 5. Doorbar J. Molecular biology of human papillomavirus infection and cervical cancer. Clin Sci (Lond) 2006; 110(5):525–41.
- Bosch FX, Burchell AN, Schiffman M, Giuliano AR, de Sanjose S, Bruni L, Epidemiology and natural history of human papillomavirus infections and type-specific implications in cervical neoplasia. Vaccine 2008; 26 (Suppl10): K1–16.
- Ekström J, Bzhalava D, Svenback D, Forslund O, Dillner J. High throughput sequencing reveals diversity of Human Papillomaviruses in cutaneous lesions. Int J Cancer 2011; 129(11):2643–50.

- 8. De Villiers, E. M., Fauquet, C., Broker, T. R., Bernard, H. U. and zur Hausen, H. Classification of papillomaviruses. Virology 2004; 324: 17–27.
- 9. Harwood CA. and Proby CM. Human papillomaviruses and non-melanoma skin cancer. Curr Opin Infect Dis, 2002; 15:101–114.
- Pfister, H. Human papillomavirus and skin cancer. J. Natl. Cancer. Inst. Monogr. 2003; 31, 52–56.
- 11. Nindl I, Gottschling M, Stockfleth E. Human papillomaviruses and nonmelanoma skin cancer: basic virology and clinical manifestations. Dis Markers 2007; 23(4):247–59.
- Gottschling M, Göker M, Köhler A, Lehmann MD, Stockfleth E, Nindl I. Cutaneotropic human beta-/gamma-papillomaviruses are rarely shared between family members. J Invest Dermatol 2009; 129(10):2427–34.
- Bottalico D, Chen Z, Dunne A, Ostoloza J, McKinney S, Sun C. The oral cavity contains abundant known and novel human papillomaviruses from the Betapapillomavirus and Gammapapillomavirus genera. J Infect Dis. 204(5):787-92.
- 14. Ekstrom J, Forslund O, Dillner J. Three novel papillomaviruses (HPV109, HPV112 and HPV114) and their presence in cutaneous and mucosal samples. Virology 2010; 397(2): 331–336.
- 15. Ekström J, Bzhalava D, Svenback D, Forslund O, Dillner J. High throughput sequencing reveals diversity of Human Papillomaviruses in cutaneous lesions. Int J Cancer 2011; 129(11):2643–2650.
- Forslund O. Genetic diversity of cutaneous human papillomaviruses. J Gen Virol 2007; 88(Pt 10):2662–2669.
- Tao M, Kruhlak M, Xia S, Androphy E and Zheng Z-M. Signals That Dictate Nuclear Localization of Human Papillomavirus Type 16 Oncoprotein E6 in Living Cells. Journal of Virology, 2003; 77(24):13232–13247.
- Lowy DR, and Howley PM. Papillomaviruses, in Knipe DM, Howley PM., Griffin DE, Lamb RA, Martin MA, Roizman B and Straus SE (ed.), Fields virology, 4th ed., vol. 2. Lippincott Williams & Wilkins, Philadelphia, PA, pp 2231–2264, 2001.
- 19. Donne AJ, Hampson L, Homer JJ, I.N. Hampson. The role of HPV type in Recurrent Respiratory Papillomatosis. International Journal of Pediatric Otorhinolaryngology (2010). 74: 7–14.
- Kahn JA. HPV vaccination for the prevention of cervical intraepithelial neoplasia. N. Engl. J. Med. 2009; 361 (3): 271–8.
- 21. National Cervical Cancer Coalition (NCCC). www.nccc-online.org.
- 22. Human papillomaviruses. IARC monographs on the evaluation of carcinogenic risks to humans.

International Agency for Research on Cancer, Vol. 64. Lyon, France, 1995.

- 23. Ho GY, Bierman R, Beardsley L, et al. Natural history of cervicovaginal papillomavirus infection in young women. N Engl J Med 1998; 338:423–8.
- 24. Hildesheim A, Schiffman MH, Gravitt PE. Persistence of type-specific human papillomavirus infection among cytologically normal women. J Infect Dis 1994; 169: 235–40.
- 25. Moscicki AB, Shiboski S, Broering J. The natural history of human papillomavirus infection as measured by repeated DNA testing in adolescent and young women. J Pediatr 1998; 132:277–84.
- 26. Franco EL, Villa LL, Sobrinho JP. Epidemiology of acquisition and clearance of cervical human papillomavirus infection in women from a high-risk area for cervical cancer. J Infect Dis 1999; 180:1415–23.
- Roldão A, Mellado MC, Castilho LR, Carrondo MJ, Alves PM. Virus-like particles in vaccine development. Expert Rev Vaccines. 2010; 9(10):1149-76.
- 28. Einstein MH, Baron M, Levin MJ, Chatterjee A, Edwards RP, Zepp F, Carletti I, Dessy FJ, Trofa AF, Schuind A and Dubin G. Comparison of the immunogenicity and safety of Cervarix and Gardasil(R) human papillomavirus (HPV) cervical cancer vaccines in healthy women aged 18-45 years. Hum. Vaccine. 2009; 5:705-719.
- 29. Human Papillomavirus (HPV) Vaccines. National Cancer Institute FactSheet, National Cancer Institute at the National Institute of Health. http://www.cancer.gov/cancertopics/factsheet/pre vention/HPV-vaccine.
- Einstein MH, Baron M, Levin MJ, Chatterjee A, Fox B, Scholar S, Rosen J, Chak N, Meric D, Ressy FJ, Datta SK, Decamps D, Dublin G. Comparative immunogenicity and safety of human papillomavirus. Hum Vaccine, 2011; 7(12):1343-1358.
- Gonçalves AK, Cobucci RN, Rodrigues HM, de Melo AG, Giraldo PC. Safety, tolerability and side effects of human papillomavirus vaccines: a systematic quantitative review. Braz J Infect Dis. 2014; pii: S1413-8670(14)00069-5.
- 32. E.S. Trindade, Safety profile of human papillomavirus vaccines Rev Bras Patol Trato Genit Infer, 2 (2012), pp. 76–80
- 33. S. Kang, K.H. Kim, Y.T. Kim. Safety and immunogenicity of a vaccine targeting human papillomavirus types 6, 11, 16 and 18: a randomized, placebo-controlled trial in 176 Korean subjects. Int. J Gynecol Cancer, 18 (2008), pp. 1013–1019.
- 34. N. Bhatla, V. Suri, P. Basu. Immunogenicity and safety of human papillomavirus-16/18 AS04-adjuvanted cervical cancer vaccine in healthy

Indian women J Obstet Gynaecol Res, 36 (2010), pp. 123–132.

- 35. S. Khatun, S.M. Akram Hussain, S. Chowdhury. Safety and immunogenicity profile of human papillomavirus-16/18 AS04 adjuvant cervical cancer vaccine: a randomized controlled trial in healthy adolescent girls of Bangladesh. Jpn J Clin Oncol, 42, pp. 36–41. 2012.
- 36. Daraj Kanduc. Quantifying the possible crossreactivity risk of an HPV16 vaccine Journal of Experimental therapeutics and oncology, vol 8, 65-76. 2009.
- Stephanie Seneff, Robert M. Davidson, Jingjing Liu. Is Cholesterol Sulfate Deficiency Common Factor in Preeclampsia, Autism, and Pernicious Anemia? Entropy 14, 2265-2290. 2012,
- 38. Shope RE. Immunization of rabbits to infectious papillomatosis. J. Exp. Med. 1937; 65:607–624.
- 39. Qinjian Z, Shaowei L, Hai Y, Ningshao X, and Yorgo M. Virus-like particle-based human vaccines: quality assessment based on structural and functional properties. Trends in Biotechnology, 2013; 31 (11): 654-63.
- 40. Oyewumi MO, Kumar A and Cui Z. Nanomicroparticles as immune adjuvants: correlating particle sizes and the resultant immune responses. Expert Rev Vaccines 2010; 9(9): 1095–1107.
- Schijns V. Immunological concepts for vaccine adjuvant activity. Curr Opin Immunol 2000; 1 2(4): 456–463.
- 42. Schiller JT, Hidesheim A. Developing HPV viruslike particle vaccines to prevent cervical cancer: a progress report. Journal of Clinical Virology, 2000; 19: 67–74.
- 43. Kirnbauer R, Booy F, Cheng N, Lowy DR, Schiller JT. Papillomavirus L1 major capsid protein self-assembles into virus-like particles that are highly immunogenic. Proc Natl Acad Sci., 1992; 89:12180–4.
- 44. Rose RC, Reichman RC, Bonnez W. Human papillomavirus (HPV) type 11 recombinant viruslike particles induce the formation of neutralizing antibodies and detect HPV-specific antibodies in human sera. Journal of General Virology, 1994; 75:2075–9.
- 45. Schiller JT, Roden RBS. Papillomavirus-like particles: basic and applied studies. In: Lacey C, editor. Papillomavirus Reviews: Current Research on Papillomaviruses. Leeds Medical Information, Leeds, UK. 1996; pp 101–12.
- 46. Petit AM, Rak J, Hung MC, Rockwell P, Goldstein N, Fendly B, Kerbel RS. Neutralizing Antibodies against Epidermal Growth Factor and ErbB-2lneu Receptor Tyrosine Kinases Down-Regulate Vascular Endothelial Growth Factor Production by Tumor Cells in Vitro and in Vivo. American Journal of Pathology, 1997; 151(6):1523-30.

- 47. Sapp M, Bienkowska-Haba M. Viral entry mechanisms: human papillomavirus and a long journey from extracellular matrix to the nucleus. FEBS J. 2009; 276:7206–7216.
- 48. Day PM. In vivo mechanisms of vaccine-induced protection against HPV infection. Cell Host Microbe, 2010; 8:260–270.
- 49. Safaeian M. Epidemiological study of anti-HPV16/18 seropositivity and subsequent risk of HPV16 and -18 infections. J. Natl. Cancer Inst. 2010; 102:1653–1662.
- 50. Stanley MA. Intra-epithelial vaccination with COPV L1 DNA by particle-mediated DNA delivery protects against mucosal challenge with infectious COPV in beagle dogs. Vaccine 2001; 19:2783–2792.
- Asghar Abdoli, Hoorieh Soleimanjahi, Fatemeh Fotouhi, Ali Teimoori, Shahram Pour Beiranvand, Zahra Kianmehr. Human Papillomavirus Type16-L1 VLP Production in Insect Cells. Iran J Basic Med Sci.; 16(8): 891–895. 2013
- 52. Bonnez W, Reichman RC. Papilomaviruses. In: Mandell GL, Bennett JE, Dolin R. Principles and practice of infectious diseases. New York: Churchill Livingstone Inc., pp 2035-50, 2010.
- 53. Markowitz LE, Dunne EF, Saraiya M, Lawson HW, Chesson H, Unger ER; Centers for Disease Control and Prevention (CDC); Advisory Committee on Immunization Practices (ACIP). Quadrivalent Human Papillomavirus Vaccine: Recommendations of the Advisory Committee on Immunization Practices (ACIP). MMWR Recomm Rep. 2007; 56(RR-2):1-24.
- 54. Kari P Braaten KP, Laufer MR. Human Papillomavirus (HPV), HPV-Related Disease and the HPV Vaccine. Rev Obstet Gynecol. 2008 Winter; 1(1): 2–10.
- Caputo A, Sparnacci K, Ensoli B, Tondelli L. Functional polymeric nano/microparticles for surface adsorption and delivery of protein and DNA vaccines. Curr Drug Deliv 2008; 5(4):230– 242. [PubMed:18855591]
- 56. Petrovsky N, Aguilar JC. Vaccine adjuvants: current state and future trends. Immunol Cell Biol 2004; 82 (5):488–496.
- 57. Copland MJ, Baird MA, Rades T. Liposomal delivery of antigen to human dendritic cells. Vaccine 2003; 21(9–10):883–890.
- Schijns V. Immunological concepts for vaccine adjuvant activity. Curr Opin Immunol 2000; 12(4): 456–463.
- 59. Bissa M, Zanotto C, Pacchioni S, Volonté L, Venuti A, Lembo D, De Giuli Morghen C, Radaelli A. The L1 protein of human papilloma virus 16 expressed by a fowlpox virus recombinant can assemble into virus-like particles in

mammalian cell lines but elicits a non-neutralising humoral response Antiviral Res. 2015 Feb 5.

- 60. Skwarczynski1 M, Toth I. Recent advances in peptide-based subunit nanovaccines. Nanomedicine, Vol. 9, No. 17, Pages 2657-2669.
- 61. Hung CF, Ma B, Monie A, Tsen SW, Wu TC. Therapeutic human papillomavirus vaccines: current clinical trials and future directions. Expert Opin Biol Ther. 2008; 8:421–439.
- 62. Daayana S, Elkord E, Winters U, Pawlita M, Roden R, Stern PL, Kitchener HC. Phase II trial of imiquimod and HPV therapeutic vaccination in patients with vulval intraepithelial neoplasia. Br J Cancer. 2010; 102:1129–1136.
- Kenter GG, Welters MJ, Valentijn AR, Lowik MJ, Berends-van der Meer DM, Vloon AP, Essahsah F, Fathers LM, Offringa R, Drijfhout JW, Wafelman AR, Oostendorp J, Fleuren GJ, van der Burg SH, Melief CJ. Vaccination against HPV-16 oncoproteins for vulvar intraepithelial neoplasia. N Engl J Med. 2009; 361:1838–1847.
- 64. Kaech SM, Wherry EJ, Ahmed R. Effector and memory T-cell differentiation: implications for vaccine development. Nat Rev Immunol. 2002; 2:251–262.
- 65. Yee C, Thompson JA, Byrd D, Riddell SR, Roche P, Celis E, Greenberg PD. Adoptive T cell therapy using antigen-specific CD8+ T cell clones for the treatment of patients with metastatic melanoma: in vivo persistence, migration, and antitumor effect of transferred T cells. Proc. Natl. Acad. Sci. 2002; 99:16168–16173.
- 66. Dudley ME, Wunderlich JR, Robbins PF, Yang JC, Hwu P, Schwartzentruber DJ, Topalian SL, Sherry R, Restifo NP, Hubicki AM, Robinson MR, Raffeld M, Duray P, Seipp CA, Rogers-Freezer L, Morton KE, Mavroukakis SA, White DE, Rosenberg SA. Cancer regression and autoimmunity in patients after clonal repopulation with antitumor lymphocytes. Science. 2002; 298:850–854.
- 67. Wu CY, Monie A, Pang X, Hung CF, Wu TC. Improving therapeutic HPV peptide-based vaccine potency by enhancing CD4+ T help and dendritic cell activation. J Biomed Sci. 2010; 17:88.
- 68. Melief CJ, van der Burg SH. Immunotherapy of established (pre)malignant disease by synthetic long peptide vaccines. Nat Rev Cancer. 2008; 8:351–360.
- 69. van Driel WJ, Ressing ME, Kenter GG, Brandt RM, Krul EJ, van Rossum AB, Schuuring E, Offringa R, Bauknecht T, Tamm-Hermelink A, van Dam PA, Fleuren GJ, Kast WM, Melief CJ, Trimbos JB. Vaccination with HPV16 peptides of patients with advanced cervical carcinoma: clinical evaluation of a phase I-II trial. Eur J Cancer. 1999; 35:946–952.

- 70. Barrios K1, Celis E. TriVax-HPV: an improved peptide-based therapeutic vaccination strategy against human papillomavirus-induced cancers. Cancer Immunol Immunother. 2012 Aug; 61(8):1307-17.
- Assudani D, Cho HI, DeVito N, Bradley N, Celis E. In vivo expansion, persistence, and function of peptide vaccine-induced CD8 T cells occur independently of CD4 T cells. Cancer Res. 2008; 68:9892–9899.
- 72. Cho HI, Celis E. Optimized peptide vaccines eliciting extensive CD8 T-cell responses with therapeutic antitumor effects. Cancer Res. 2009; 69:9012–9019.
- 73. Rahimian S, Fransen MF, Kleinovink JW2, Christensen JR1, Amidi M1, Hennink WE, Ossendorp F. Polymeric nanoparticles for codelivery of synthetic long peptide antigen and poly IC as therapeutic cancer vaccine formulation. J Control Release. 2015 Feb 5.
- Chien-Fu H, Archana M, Ronald DA, and T.-C. DNA vaccines for cervical cancer: from bench to bedside. Exp Mol Med. 2007 December 31; 39(6): 679–689.
- 75. Yan X, Wang D, Liang F, Fu L, Guo C. HPV16L1attenuated Shigella recombinant vaccine induced strong vaginal and systemic immune responses in guinea pig model. Hum Vaccin Immunother. 2014 Nov 1:0.
- 76. Xu X, Ho W, Zhang X, Bertrand N, Farokhzad O. Cancer nanomedicine: from targeted delivery to combination therapy. Trends Mol Med. 2015 Feb.
- 77. Andrew ZW. Robert L. Omid CF, Nanoparticle Delivery of Cancer Drugs, Annual Review of Medicine, Vol. 63: 185-198. February 2012.
- Arlen PM, Kaufman HL, DiPaola RS. Pox viral vaccine approaches. Semin Oncol 2005; 32(6):549–55.
- 79. Moll H. Antigen delivery by dendritic cells. Int J Med Microbiol 2004;294(5):337–44.
- Liu M, Acres B, Balloul JM, Bizouarne N, Paul S, Slos P, et al. Gene-based vaccines and immunotherapeutics. Proc Natl Acad Sci USA 2004;101 :14567–71.
- 81. Hilleman MR. Vaccines in historic evolution and perspective: a narrative of vaccine discoveries.Vaccine 2000;18(15):1436–47.
- Combadière B1, Mahé B. Particle- based vaccines for transcutaneous vaccination. Comp Immunol Microbiol Infect Dis. 2008 Mar;31(2-3):293-315. Epub 2007 Oct 30.
- Reddy ST, Rehor A, Schmoekel HG, Hubbell JA, Swartz MA. In vivo targeting of dendritic cells in lymph nodes with poly(propylene sulfide) nanoparticles. J Controlled Release 2006; 112(1):26–34.

- 84. Walter E, Dreher D, Kok M, et al. Hydrophilic poly(D,L-lactide-co-glycolide) microspheres for the delivery of DNA to human-derived macrophages and dendritic cells. J Controlled Release 2001; 76 (1–2):149–168.
- 85. Rice-Fichte AC, Arenas-Gambia AM, Kahl-McDonagh MM, Ficht TA. Polymeric particles in vaccine delivery. Reviews particle-based properties, such as surface chemistry and sizes that will affect antigen presentation and processing by APCs. Curr Opin Microbiol 2009; 13(1):106–112.
- 86. Thomasin C, Corradin G, Men Y, Merkle HP, Gander B. Tetanus toxoid and synthetic malaria antigen containing poly(lactide)/poly(lactide-coglycolide) microspheres: importance of polymer degradation and antigen release for immune response. J Controlled Release 1996; 41(1– 2):131–145.
- Mallapragada SK, Narasimhan B. Immunomodulatory biomaterials. Int J Pharm 2008; 364(2):265–271.
- Slütter B, Soema PC, Ding Z, Verheul R, Hennink W, Jiskoot W. Conjugation of ovalbumin to trimethyl chitosan improves immunogenicity of the antigen. J Controlled Release 2010; 143(2):207–214.
- O'Hagan DT. Microparticles and polymers for the mucosal delivery of vaccines. Adv Drug Deliv Rev 1998; 34(2–3):305–320.
- 90. Jain S, Yap WT, Irvine DJ. Synthesis of proteinloaded hydrogel particles in an aqueous two-phase system for coincident antigen and CpG oligonucleotide delivery to antigen-presenting cells. Biomacromolecules 2005; 6(5):2590–2600.
- 91. Shen Z, Reznikoff G, Dranoff G, Rock K. Cloned dendritic cells can present exogenous antigens on both MHC class I and class II molecules. J Immunol 1997; 158(6):2723–2730.
- Singh M, Briones M, Ott G, O'Hagan DT. Cationic microparticles: a potent delivery system for DNA vaccines. Proc Natl Acad Sci USA 2000; 97:811–816.
- 93. Petrovsky N, Aguilar JC. Vaccine adjuvants: current state and future trends. Immunol Cell Biol 2004; 82(5):488–496.
- 94. Panyam J, Labhasetwar V. Biodegradable nanoparticles for drug and gene delivery to cells and tissue. Adv Drug Deliv Rev 2003; 55(3):329– 347.
- Tobío M, Gref R, Sánchez A, Langer R, Alonso MJ. Stealth PLA-PEG nanoparticles as protein carriers for nasal administration. Pharm Res 1998; 15(2):270–275.
- 96. Rieger J, Freichels H, Imberty A. Polyester nanoparticles presenting mannose residues: toward the development of new vaccine delivery systems combining biodegradability and targeting

properties. Biomacromolecules 2009; 10(3):651-657.

- 97. Andrianov AK, Marin A, Roberts BE. Polyphosphazene polyelectrolytes: a link between the formation of noncovalent complexes with antigenic proteins and immunostimulating activity. Biomacromolecules 2005; 6(3):1375– 1379.
- 98. Zwiorek K, Bourquin C, Battiany J. Delivery by cationic gelatin nanoparticles strongly increases the immunostimulatory effects of CpG oligonucleotides. Pharm Res 2008; 25(3):551–562.
- 99. Coppi G, Iannuccelli V, Sala N, Bondi M. Alginate microparticles for polymyxin B Peyer's patches uptake: microparticles for antibiotic oral administration. J Microencap Micro Nano Carriers 2004; 21(8):829–839.
- 100. Massimiliano Tait, Alessandro Pegoretti, Andrea Dorigato, Kyriaki Kalaitzidou. The effect of filler type and content and manufacturing process on the performance of multifunctional carbon polylactide composites. Carbon 2011; 49: 4280-4290
- 101. Parkin J, Cohen B. An overview of the immune system. The Lancet 2001; 357(9270):1777-89.
- 102. Ahsan F, Rivas IP, Khan MA, Torres Suarez AI. Targeting to macrophages: role of physicochemical properties of particulate carriersliposomes and microspheres-on the phagocytosis of microspheres. J Control Rel 2002; 79:29–40.
- 103. Thiele L, Merkle HP, Walter E. Phagocytosis and phagosomal fate of surface-modified microparticles in dendritic cells and macrophages. Pharm Res 2003; 20:221–8.
- Kreeuter J. Nanoparticles and microparticles for drug and vaccine delivery. Journal of Anatomy 1996; 189 (3): 503–505.
- 105. Elamanchili P, Diwan M, Cao M, Samuel J. Characterization of poly(D,L,-lactic-co-glycolic acid) based nanoparticulate system for enhanced delivery of antigens to dendritic cells. Vaccine 2004; 22 (19):2406–2412.
- 106. Cui Z, Mumper RJ. Topical immunization using nanoengineered genetic vaccines. J Controlled Release 2002; 81(1–2):173–184.
- 107. Cui Z, Mumper RJ. Genetic Immunization Using nanoparticles engineered from microemulsion precursors. Pharm Res 2002; 19(7):939–946.
- 108. Sloat BR, Sandoval MA, Hau AM, He Y, Cui Z. Strong antibody responses induced by protein antigens conjugated onto the surface of lecithinbased nanoparticles. J Controlled Release 2010; 141 (1):93–100.
- 109. Frey A, Neutra MR. Targeting of mucosal vaccines to Peyer's patch M cells. Behring Inst Mitt. 1997 Feb; (98):376-89.

- 110. Flick-Smith,HC, Eyles,JE, Hebdon R, Waters EL, Beedham RJ, Stagg TJ, Miller J, Alpar HO, Baillie LW, Williamson ED. Mucosal or parenteral administration of microsphere-associated Bacillus anthracis protective antigen protects against anthrax infection in mice. Infect. Immun. 2002, 70; 2022-2028.
- 111. Laoui-Attarki K, Pecquet S., Fattal E, Trolle S, Chachaty E, Couvreur P, and Andremont A."Protective immunity against Salmonella typhimurium elicited in mice by oral vaccination with phosphorylcholine encapsulated in poly(DLlactide-co-glycolide) microspheres". Infect. Immun. 1997; 65, 853-857.
- 112. Uddin AN, Bejugam NK, Gayakwad S, Akther P, and D'Souza MJ. Oral delivery of gastro-resistant microencapsulated typhoid vaccine. J. Drug Target. 2009; 17, 553-560.
- 113. Gamvrellis A1, Gloster S, Jefferies M, Mottram PL, Smooker P, Plebanski M, Scheerlinck JP. Characterization of local immune responses induced by a novel nano-particle based carrieradjuvant in sheep. Vet Immunol Immunopathol. 2013; 155(1-2):21-9.
- 114. Hamdy S1, Haddadi A, Hung RW, Lavasanifar A. Targeting dendritic cells with nano-particulate PLGA cancer vaccine formulations. Adv Drug Deliv Rev. 2011; 63(10-11):943-55.
- 115. Elamanchili P1, Lutsiak CM, Hamdy S, Diwan M, Samuel J. "Pathogen-mimicking" nanoparticles for vaccine delivery to dendritic cells. J Immunother. 2007; 30(4):378-95.
- 116. Diwan M1, Elamanchili P, Lane H, Gainer A, Samuel J. Biodegradable nanoparticle mediated antigen delivery to human cord blood derived dendritic cells for induction of primary T cell responses. J Drug Target. 2003; 11(8-10):495-507.
- 117. Sasagawa T, Tani M, Basha W, Rose RC, Tohda H, Giga-Hama Y, Azar KK, Yasuda H, Sakai A, and Inoue, M. A human papillomavirus type 16 vaccine by oral delivery of L1 protein, Virus Res. 2005; 110:81-90.
- 118. Thönes N, Müller M. Oral immunization with different assembly forms of the HPV16 major capsid protein L1 induces neutralizing antibodies and cytotoxic T-lymphocytes. Virology 2007; 20; 369(2):375-88.

- 119. Chang S, Warner J, Liang L, Fairman J. A novel vaccine adjuvant for recombinant flu antigens. Biologicals 2009; 37(3):141–147.
- 120. Gerber S, Lane C, Brown DM, Lord E, Dilorenzo M, Clements JD, Rybicki E, Williamson AL, and Rose RC. Human Papillomavirus Virus-Like Particles Are Efficient Oral Immunogens when Coadministered with Escherichia coli Heat-Labile Enterotoxin Mutant R192G or CpG DNA. Journal of Virology 2001; 75:4752-4760.
- 121. Fraillery D, Baud D, Pang SY, Schiller J, Bobst M, Zosso N, Ponci F, Nardelli-Haefliger D. Salmonella enterica serovar Typhi Ty21a expressing human papillomavirus type 16 L1 as a potential live vaccine against cervical cancer and typhoid fever. Clin Vaccine Immunol 2007; 14(10):1285-95.
- 122. Patel NJ, Addo R, Ubale R, Uddin, MN, D'Souza MJ, Jobe L. The Effect of Antisense to NF-κB in an Albumin Microsphere Formulation on the Progression of Left-Ventricular Remodeling Associated with Chronic Volume Overload in Rats. Journal of Drug Targeting. 2014; 22, 9,796-804.
- 123. Bhowmik T, D'Souza B, Uddin MN, D'Souza MJ. Oral delivery of microparticles containing plasmid DNA encoding hepatitis-B surface antigen. Journal of Drug Targeting, 2012; 20(4): 364–371.
- 124. Uddin MN, Do DP, Pai BS, Gayakwad S, Oettinger CW, D'Souza MJ. A methodology for quantitation and characterization of oligonucleotides in albumin microspheres. Analyst, 2009, 134, 1483–1489
- 125. Uddin MN, Patel NJ, Bhowmik T, D'Souza B, Akalkotkar A, Etzlar F, Oettinger CW, D'Souza M. Enhanced bioavailability of orally administered antisense oligonucleotide to nuclear factor kappa B mRNA after microencapsulation with albumin. J Drug Target. 2013; 21(5):450-7.
- 126. Koji H, Kazuya K, Tomonori N, Gemilson SP, Shinji F, Masashi E, Kazunori K, Toru T, Yumiko F, Sayaka K, Atsuko Y, Satoshi W, Gaku N, Shunsuke KL, Takaya M, Mitsutoshi IK, Kimiyo H, Shin-Ichi FU, Anson WL, Kikuji I, Hiroshi K, Hiroshi O. Uptake through glycoprotein 2 of FimH+ bacteria by M cells initiates mucosal immune response. Nature, 2009, 462, 226-230.