Microneedles as Enhancer of Drug Absorption Through the Skin and Applications in Medicine and Cosmetology

Pablo Serrano-Castañeda^{1,2}, José Juan Escobar-Chávez², Isabel Marlen Rodríguez-Cruz³, Luz María Melgoza-Contreras¹, Jessica Martínez-Hernández²

¹Doctorado en Ciencias Biológicas y de la Salud de la Universidad Autónoma Metropolitana. México. ²Facultad de Estudios Superiores Cuautitlán-Universidad Nacional Autónoma de México. Unidad de Investigación Multidisciplinaria. Estado de México, México. ³Hospital Regional de Alta Especialidad de Zumpango. Unidad de Enseñanza e Investigación. Zumpango Estado de México, México.

Received January 1, 2018; Revised, February 1, 2018; Accepted, February 2, 2018; Published February 19, 2018.

ABSTRACT - The microneedles technology has found applications in many health-related fields. For example, their application in drugs and vaccines delivery as well, as the determination of biomarkers, has been reported. They also have a place in the dermatology and cosmetic areas such as the treatment of wounds from burns, scars, acne, depigmentation, and alopecia will be shown. Microneedles are used in therapeutic applications and are manufactured using materials such as metal (steel, titanium, nickel), polymer (oly-glycolic acid (PGA), poly-lactide-co-glycolide acid (PLGA), poly-L-lactic acid (PLA), chitosan), glass, silicon, ceramic, carbohydrates (trehalose, sucrose, mannitol). Examples of application of microneedles and their advantages and disadvantages are discussed.

This article is open to **POST-PUBLICATION REVIEW**. Registered readers (see "For Readers") may **comment** by clicking on ABSTRACT on the issue's contents page.

INTRODUCTION

At present, the human skin is a route for delivering drugs with either local effect or with systemic therapeutic effect, and it represents a highly attractive alternative compared to conventional dosage forms including injections, capsules, and tablets (1). This route of drug administration has advantages, such as avoiding first-pass metabolism, controlled drug release, and reduction of dosages. It is painless, easy to administer, and noninvasive (1-4). Nevertheless, the principal limitation for a transdermal drug delivery system is the skin itself (1, 5-6). The external layer of the skin is the stratum corneum (SC), which is the principal barrier of protection from microorganisms and dangerous agents (1-7). For this reason, it is necessary to use chemical or physical penetration enhancers with the intention of increasing permeability of different substances through the skin

In the case of physical enhancers, microneedles have been shown to increase the drug permeability through the skin by up to 3 orders of magnitude by passing the SC. Microneedles can allow transdermal delivery of many drugs and macromolecules, such as

insulin, peptides, and other biomolecules that normally

cannot diffuse through the skin (6, 18-20). In addition, the microneedles enter the upper layer of the skin without reaching the nerves, making the drug delivery painless (6, 18-20).

The aim of this review is to offer an overview of the use of microneedles as a drug delivery system in medicine, pharmacy, and cosmetology and to show the principles, limitations, and pharmacological profiles for each field.

SKIN STRUCTURE

Human skin has several functions such as photoprotection, thermoregulation, hormonal synthesis, sensory perception, and protective function as a barrier for chemical, physical, and microbial agents (5, 21-22). Anatomically, the skin

Corresponding Author: José Juan Escobar-Chávez, Facultad de Estudios Superiores Cuautitlán-Universidad Nacional Autónoma de México. Unidad de Investigación Multidisciplinaria. Laboratorio 12: Sistemas Transdérmicos Carretera Cuautitlán Teoloyucan, km 2.5 San Sebastián Xhala C.P 54714 Cuautitlán Izcalli, Estado de México, México. josejuanescobar@comunidad.unam.mx

has three main layers, the epidermis, dermis, and hypodermis (subcutaneous tissue). Figure 1 shows the component layers of the skin (23).

The epidermis thickness is around 0.12 mm and comprises five layers: the basal or germinative layer, the stratum spinosum, the granular layer, the lucidum layer, and the SC (24-28). The SC is the main barrier that shields the skin from the entry of foreign substances. The SC has an average thickness of 20 μm; this layer is composed of corneocytes (dead anucleated epidermal cells), which are filled with keratin filaments and embedded in a continuous multilamellar lipid matrix. The lipid composition is complex, it includes components like long-chain ceramides, fatty acids, and cholesterol compared to most other biological membranes that basically have phospholipids (24). Moreover, the SC is a selective membrane that controls the penetration of substances into the skin and prevents water loss. The lower three sublayers (stratum granulosum, stratum spinosum, and stratum basale) constitute the viable part of the

epidermis that has cells like melanocytes, Langerhans cells, and Merkel cells (5, 21-24).

The dermis has a thickness of 3-5 mm. This matrix is formed of connective tissue, and it is made up of collagen fibers and elastin (25). This tissue is vascularized, presenting blood vessels and nerves and includes apocrine glands, sweat glands, and pilosebaceous follicles (5, 26-27). The dermis is constituted by the subcutaneous fascia and chorion layers.

The hypodermis is the innermost layer of the skin, where the function of transporting nutrients and migrating cells is observed. It acts as an isolator, which helps the body to retain heat. This layer is constituted of adipose tissue. This skin layer is where more irrigation exists because there are many blood vessels in addition to numerous nerve endings (5, 24-27).

It is important to mention that the skin thickness is different according to gender, race, age, and anatomical region (24-27). Moreover, the thickness of the skin is a critical factor in transdermal drug delivery.

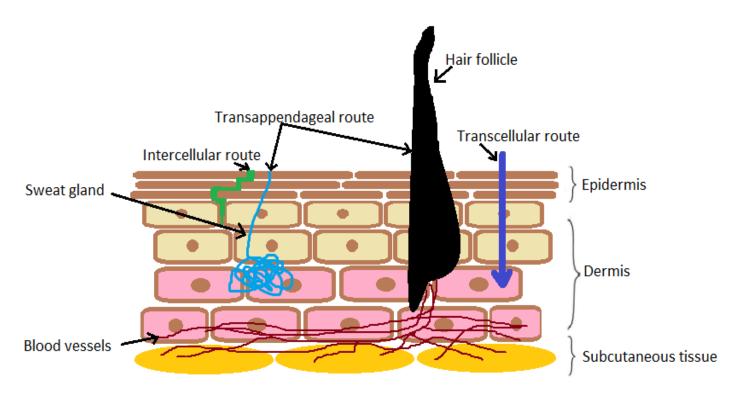


Figure 1. Layers of the skin and penetration routes.

PENETRATION DRUG ROUTES THROUGH THE SKIN

Human skin is an interesting route for drug therapy since it offers an accessible route, avoiding the first-pass hepatic metabolism. The main routes of penetration through the SC barrier are transappendageal, intercellular, and intracellular routes.

The *trans-appendageal route* is the transport via of pores, embracing sweat glands and hair follicles with their associated sebaceous glands; nonetheless, the classical concept considers that the transappendageal route is not a significant transdermal route because hair follicles and sweat glands occupy 0.1% of the surface of the human skin (29-30). However, it is an important mode of entrance for large polar molecules and ions, which can barely pass through the SC.

The *intercellular route* is the principal route of entry for lipophilic drugs due to the dense packing of proteins within the corneccytes, which make them almost impermeable (31-32).

In the *intracellular route*, the drug is mainly driven by its partition coefficient (log P). Hydrophilic drugs can diffuse via the intracellular route. In contrast, lipophilic drugs preferably cross the SC via the intercellular domain (5, 29, 31-32).

Transdermal penetration through the SC is illustrated in Figure 1. As seen, transdermal penetration may occur between the cells (intercellular) or through cells (transcellular route). The relative contribution of these pathways depends on the solubility, partition coefficients, and diffusivity of the drug within proteins or lipids. These skin diffusion processes are mediated by a limiting process of passive diffusion. The rate and extent of transport follow Fick's law (2, 5, 33).

Only molecules with appropriate physicochemical properties can passively diffuse through the skin membrane. In recent years, many different chemicals and physical permeation enhancers have been developed. Among the most recent and promising techniques is the use of a microneedle based on improving the skin transport of molecules.

MICRONEEDLES

Microneedle technology for drug delivery was first developed in 1971 for an invention by Gerstel and Place (34-36), but until the 1990s, microneedle drug delivery advanced with microelectronics technology (6, 35). Microneedles are micron-sized needles that are used to open holes into the skin to create micro-

channels for the subsequent delivery of drugs and thus cross the SC (6). The microneedles enter the upper layer of the skin without touching the nerves, thus delivering drugs transdermally in a painless manner. Microneedles usually have a diameter of a few hundred microns, which recedes toward the sharp tip and has a length of 50 to 900 μ m (37).

Classification of Microneedles

Microneedles can be classified based on applications (medicine, pharmacy, and cosmetology), material (metal, polymer, glass, silicon, ceramic, hydrogel, and sugar), manufacturing technique (etching, injection molding, micromachining, micro-molding, and lithography electroforming replication), or design (hollow or solid).

Microneedles should have the appropriate combination of mechanical strength, toughness, and hardness to disrupt the SC without fracture and buckling failure. In addition, microneedle size must be small enough to ensure painlessness and minimal invasiveness. Moreover, the drug delivery efficiency should also be fully considered during microneedle design (38).

Microneedles by Material

Microneedles can be fabricated from a wide diversity of materials, for example, metal, polymer, glass, silicon, ceramic, hydrogel, and sugar.

Silicon. This material has been developed for several decades because this material has relatively high hardness. However, the manufacturing methods of silicon are expensive and need clean room processing (39). Moreover, silicon is a fragile material; thus, silicon microneedles are prone to fracture in transportation and application (38-39). Therefore, microneedles made from brittle materials like silicon, ceramic, and glass could present problems at the time of application. In addition, silica glass causes granulomas in the skin. Currently, there insufficient evidence regarding biocompatibility of silica glass and silicon for microneedle manufacture (35-39).

Metal. The metals used in the manufacture of microneedles are stainless steel, titanium, nickel, palladium, and palladium-cobalt arrays. Metal microneedles usually have quite well-integrated mechanical properties, including high toughness, strength, and hardness, which can protect microneedles against mechanical failure (38). Metal

microneedles can be manufactured at relatively low cost using a variety of methods (electroplating, photochemical etching, micro-milling, and laser cutting). Titanium is a viable alternative to stainless steel because this material is adequately strong for biomedical applications. Titanium has been used mainly for biosensors and as transdermal delivery systems. Titanium alloys have good biocompatibility with excellent corrosion resistance (39). Metals are likely a more proper material to substitute silicon in microneedle production. Nevertheless, metal microneedles produce sharp bio-hazardous tip waste (38-39).

Polymer. Microneedles made of polymers generally have high toughness to support the polymer microneedles to avoid brittle fracture during their insertion into the skin. Some polymers are biodegradable, such as poly-glycolic acid (PGA), poly-lactide-co-glycolide acid (PLGA), poly-Llactic acid (PLA), and chitosan, or water-soluble, so that drugs can be encapsulated in these dissolvable microneedles. After insertion into the skin, drugs will release with the degradation or dissolution of these dissolvable microneedles. Biodegradable polymeric microneedles induce almost no harsh side effects: thus, these microneedles are considered the materials most promising due to biocompatibility, biodegradability, low toxicity, strength against breaking, and low cost. The main materials used for this kind of microneedle are poly (methyl methacrylate), poly (carbonate), poly (vinylpyrrolidone (PVP)), poly (vinyl alcohol (PVA)), polystyrene, poly (methyl vinyl ether-comaleic anhydride) and poly (methyl vinyl ether-comaleic acid) (35-39).

Ceramic. The main type of ceramic is alumina. The main advantage of this material is the resistance and good biocompatibility; nevertheless, under tensile stress, ceramic is brittle. Other types of ceramic used to prepare microneedles include calcium sulfate dihydrate and calcium phosphate dihydrate. These materials have good mechanical and drug-loading properties (36-39).

Glass. Silica glass is physiologically inert, allowing the visualization of fluid flow. Moreover, microneedles can be produced with different geometries and dimensions. However, glass is a brittle material. Borosilicate glass is more elastic because it presents a lower value of elastic moduli.

In general, glass microneedles required more time for production because they are created by hand; thus, these microneedles are not recommendable for the industry (38-39).

Sugar. Maltose is the most common sugar used to prepare microneedles. Carbohydrates (trehalose, sucrose, mannitol, xylitol, and galactose) are good alternatives because they are affordable and safe for human health (38). Nevertheless, sugar microneedles present problems in the processing, storage, and application on the skin. Moreover, the main disadvantage of this microneedle is that it needs thermal treatment in manufacturing (35-39).

Microneedle Design Solid Microneedles

These microneedles penetrate the upper layer of the skin and thus allow passage of the drug through the lower layers of the skin (35-40). The drug can be coated on the surface of solid microneedles to be inserted into the skin and then the drug dissolves into the skin. Subsequently, the needles are removed from the skin. Regarding manufacture, solid microneedles are easier to make than hollow microneedles, and they have better mechanical strength and sharper tips. Solid microneedles can be fabricated with metals or polymers, such as PLGA, PLA, PGA, hyaluronic acid, PVP, PVA, sodium alginate, chitosan, zein, carboxymethyl cellulose (CMC), and hydroxypropyl cellulose (HPC), as well as silk, chondroitin sulfate, ceramics, and sugars, such as maltose, galactose and dextrin (35-40).

Hollow Microneedles

Hollow microneedles allow the flux of the drug through the holes in the needles. Injection of drug solutions using hollow microneedles can provide control over the time and the amount of drug delivered (35-39). Once the injection is applied on the skin, the drug diffuses through the epidermis to be absorbed by the blood vessels in the dermis. One of the main benefits of the hollow microneedles is that they allow continuous diffusion through the skin (35-39). Solid and hollow microneedles have been manufactured from silicon, metal, and glass. Biodegradable microneedles can be made of biopolymers like chitosan, sugar glass, PLGA, hyaluronic acid, PVA, and PVP (19, 40-55).

ADVANTAGES AND DISADVANTAGES OF MICRONEEDLES

The advantages and disadvantages of using microneedles as a physical penetration enhancer are summarized in Table 1.

TECHNIQUES TO INSERT MICRONEEDLES INTO THE SKIN

There are diverse ways of releasing drugs from microneedles. The first is a novel technique called "poke with patch" (18-19, 59, 65-69) where solid silicon or metal microneedles are used to create micro-channels and then applying a transdermal patch to the surface of the skin. The transport occurs by drug diffusion. The second is called "coat and poke" (70-72), where the needles are first coated with the drug and then inserted into the skin. After that, the drug is released. A variation of this second method is "dip and scrape" where the microneedles are first immersed in a solution containing the drug and then the entire surface of the skin is scraped to introduce the drug into the micro-abrasions created by the needles (18, 32). The third is "Poke and flow"

for hollow microneedles delivering a drug like a micro-injection (73-74). Finally, "poke and release" is for dissolving microneedles fabricated from polymers or polysaccharides, releasing the drug during the dissolution of microneedles (See Figure 2) (67, 75-79).

APPLICATIONS OF MICRONEEDLES

Microneedles are used in different areas related to health, based on the numerous advantages they offer (48, 66) (Table 2). Microneedles are an attractive candidate to administer several drugs (anti-cancer drugs, oligonucleotides, vaccines, proteins, DNA, and even nanoparticles) throughout the skin (6, 80-81). Moreover, microneedles have many applications in the pharmacy, medicine, and cosmetology fields.

The use of microneedles in medicine has grown and allowed to administer drugs through different medical procedures such as the case of treatment for glaucoma, other important applications have been in the use of diagnostics such as monitoring of various biomarkers.

Table1. Advantages and disadvantages of microneedles.

Advantages	Disadvantages
Microneedles are a minimally invasive technique for transdermal	Microneedles can only be inserted into the skin if
drug delivery (37).	they have the correct shape and adequate physical
	properties (37).
Microneedles are very small (length of 50 to 900 μ m) (37).	Microneedles need to be applied with the required
	force to avoid breaking or bending before insertion
251 11 11 (2.1.)	(64).
Microneedles avoid first-pass effect (34).	Microneedles made of metal, stainless steel, or
	silicon have fracture risks. They can leave
16 11 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	fragments in the skin (54).
Microneedles can penetrate the SC without stimulation of nerves	Microneedles can cause skin irritation and in some
(18, 54-55).	cases allergy (55).
With a constant rate and a prolonged period, the drug can be	Microneedles need micro-tools and
administered, allowing the correct dose of drugs (56).	microelectronics to be produced in bulk (55).
There is a reduction of adverse reactions (56).	
Microneedles are easy and safe to use.	
Microneedles can be produced with high precision, accuracy, and	
low cost (18, 55). Hollow needles can be used with patches and timed pumps to	
deliver drugs at precise times (57-58).	
Hollow microneedles offer continuous infusion through the skin	
(35-39).	
Small microneedles could target drugs to each cell (6).	
They are biologically nontoxic (55, 57).	
Microneedles have less microbial penetration than conventional	
needles (55).	
Microneedles can be removed immediately if adverse reactions	
occur (58-63).	

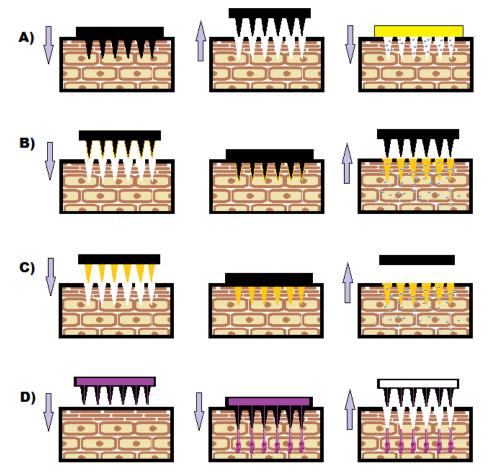


Figure 2. Schematic representation of different microneedles and mechanisms of application for transdermal drug delivery: A) solid microneedles using poke with patch, B) solid microneedles coat and poke, C) biodegradable microneedles with the mechanism poke and release, D) hollow microneedles for injections (poke and flow).

Therapeutic Applications Antiglaucoma

Kim et al. used hollow microneedles to introduce drugs into the supraciliary space. They show dramatic dose sparing of antiglaucoma agents compared with eye drops. Targeted delivery in this way increases safety, diminishes side effects, and permits a single injection with enough drug for prolonged-term sustained delivery (82).Microneedles have a safe, simple, and efficacious ocular drug delivery. However, the limited drug carrying capacity of devices demonstrated to date may limit the potential for clinical translation. Omid et al. developed microneedles which serve as reservoirs for passive delivery, the capacity of the microneedles can up to five-fold relative to solid microneedles (83).

Diagnostics

It is impossible to make a good therapy without a proper diagnosis. This is basic and essential for the success of therapy; thus, the use of microneedles has helped in this field. Sun et al. developed microneedles that can be used to release protein antigens and therapeutic proteins in the skin for allergen skin testing or immunotherapy. The microneedles with PVP deliver intact proteins or peptides to the skin for diagnostic or therapeutic applications (84).

On the other hand, Skoog et al. developed *in vivo* biosensors that enable continuous real-time detection of biomolecules for monitoring patient health. They fabricated nitrogen ultra-nanocrystalline diamond-coated titanium alloy microneedle arrays that are capable of electrochemical detection of dopamine and uric acid (85). Moreover, Li et al. fabricated

hollow microneedles with the drawing lithography technique, and a sharp tip was generated with a lasercutting system. Their results demonstrated that hollow microneedles can extract mouse blood in vivo (20 µl) at a good rate. Those microneedles with correct geometry can penetrate the skin without problems (86). Hollow microneedles can be combined with other technologies, such as biosensors and microfluidic chips to create blood analysis systems for diagnostics that would be minimally invasive, or they can be inclusively used for electrochemical detection of drugs in vivo (86-87). Dae et al. Obtain biochemical information, they develop a micro-scale needle for minimally invasive and painless blood sampling. The challenge was to combine the features of a sharp tip shape, appropriate length, and a hollow structure simultaneously (1.8 mm, an inner diameter of 60 µm, an outer tip of 100 μm, and a 60 ° bevel angle), these microneedles can be used to collect blood volumes up to 840 µL at a pressure 0.4 kPa, microneedles facilitate efficient and minimally invasive blood extraction or drug injection. Microneedles have potential applications in various blood analysis or sensor systems in pointof-care diagnostics (88).

The advantage that the microneedles offer in the diagnosis is that they can penetrate deeper layers of the skin, allowing the monitoring of bio-signals with greater reliability and thereby reducing the interferences with other substances.

Glucose monitoring

The commercial continuous glucose monitoring sensors are implanted subcutaneously for a period of 7-14 days. The subsequent biofouling effects have implications on the performance of the sensors over time, especially at low glucose concentrations. In adding, the sensors are sensitive to the presence of interfering substances like acetaminophen. Sanjiv et al. develop microneedles to eliminates this interference the microneedles operate at a lower potential (400 mV) in contrast with the commercial glucose monitoring (700 mV), reducing the interferences (89). Chua et al. made hollow silicon microneedle arrays for minimally continuous glucose monitoring (90). Vacuum pumpfluid sampling assisted interstitial microneedles in humans was studied. Microneedles show a good determination of glucose levels following insulin injection with a time lag of fewer than 20 minutes (91). The microneedles penetrating deeper layers of the skin compared to other devices

for monitoring insulin, besides they allow glucose monitoring without interference and they give us greater sensitivity.

Cancer diagnosis

Keum et al. made a dual-diagnostic system with an endo-microscope and microneedle sensor that has high-resolution imaging combined with electrical real-time detection of nitric oxide released from cancer tissues. This system can be used for simple, fast, and accurate detection of cancer in biomedical applications (92). In comparison with the traditional endomicroscopy that can only identify microscopic pathological features and often requires biopsy sampling of suspicious lesions for additional histopathological examination of cancers.

Biomarkers

Li et al. have shown that surface-modified microneedle arrays could reliably and quickly quantify biomarkers in the upper dermis after laser treatment. It could be safely performed by brief laser irradiation of the microneedle array application site. The assay was independent of the length of the microneedles or molecular mass, as IgG can be measured by this noninvasive procedure (93-94).

Other applications are being monitored, such as in electrocardiography (ECG), electromyography (EMG), and electroencephalography (EEG), which are important for understanding pathological and physiological conditions in humans. The electrodes are currently used, but they have disadvantages that can result in wrong results if not properly applied and need the use of the gel. Therefore, Forvi et al. developed microneedles based on dry electrodes for an alternative to the conventional wet electrodes in recording bio-signals in clinical examinations. Their microneedles were tested on ECG, EEG, and EMG in the short term, and they were better than conventional wet electrodes under static conditions in signal acquisition and under dynamic conditions where the wet electrode is more susceptible to artifacts. These microneedles motion comfortable, easy to apply, and do not require preparation of the skin (95). Renxin et al. improved the microneedles for monitoring EEG microneedle electrode arrays (MNEAs parylene-based) have been used as dry electrodes that could be capable of EEG monitoring without skin abrasion and gel. In the study, they develop a flexible MNEAs that can be adapted to the skin which could provide not only conformal but also robust contact, in comparison with the conventional devices (96). Lee et al. prepared microneedles of silicon for infusion of drugs in the brain used to identify neuronal connections and activities. They conclude that the proposed thin microneedles can deliver drugs in specific regions of the brain. They want to incorporate multiple inlets and outlets for delivering various drugs simultaneously and adding neural probes with drug delivery capability to detect neural spike signals (97).

Perivascular delivery

In order to inhibit intimal hyperplasia (IH) caused by abnormal growth of smooth muscle cells (SMCs) in tunica media, various perivascular drug delivery devices are reported for delivery of anti-proliferation drugs into vascular tissue. However, there still remain conflicting requirements such as local and unidirectional delivery vs device porosity, and conformal tight device installation vs pulsatile expansion and constriction of blood vessels. For these reasons, Lee et al. developed wrappable microneedles of PLGA for treat intimal hyperplasia (IH). Microneedles showed significantly reduced neointimal formation (11.1 %) compared with structure coated with 1 µg of sirolimus and microneedles with 1 µg of sirolimus (23.7 and 22.2 %) after 4-week in vivo animal study. Additionally, wrappable MN meshes effectively suppressed side effects such as IH due to mechanical constriction, loss of toxic drug to the surroundings, and cell death that were frequently observed with other previous perivascular drug delivery devices (98).

Vaccines

New vaccine technologies using microneedles offer an efficient and painless method for introducing antigens into skin that, in the future, could solve some problems in the traditional vaccination. Today vaccination programs have cold chain storage, generate vaccine waste and hazardous waste, and require trained personnel. These factors add significant costs to immunization programs. Vaccine development programs aim to reduce the cost of each dose of the vaccine, and self-administered administration does not require trained staff. Microneedle dermal vaccines avoid these problems. Moreover, stabilization of a vaccine and the problems associated with reconstitution in a liquid solution for administration are solved (60, 79).

Matsuo et al. used microneedles for permeation of peptides with different molecular weights and observed that microneedles can remarkably enhance the transdermal delivery of all hydrophilic peptides. The skin permeation of peptides depends on their molecular weight and decreases as the molecular weight increases. In addition, the enhanced skin permeation of peptides produced by microneedle pretreatment may be caused by the generation of convection. They demonstrated that microneedles provide an attractive route to deliver low molecular weight peptides to the skin (79).

Hirobe et al. created a microneedle patch MicroHyala (MH) with hyaluronic acid, and the vaccination with MH induced a strong immune response against various antigens in mice. They studied the clinical safety and effectiveness of the transcutaneous influenza vaccine (flu-MH), which contains trivalent influenza hemagglutinins (15 mg each), and showed that the administration of influenza vaccines in humans using the MH system induced high levels of immunity (54).

Edens et al. examined the formulation of microneedles with a patch to vaccinate rhesus macaques against measles. The microneedles were inserted in the skin with thumb pressure. They were dissolved in the skin in 10 min, and they caused only mild to transient skin erythema. The groups of macaques generated antibody responses to measles. In addition, the microneedles have an adequate level of potency after storage at elevated temperatures, indicating thermostability compared with standard vaccine. They concluded that microneedle vaccine for measles was immunogenic in primates and may be used as a vaccine in humans (99).

Hiraishi et al. developed a vaccine with the bacillus Calmette-Guérin (BCG). The prevention and control of tuberculosis would benefit from a novel method of BCG vaccination that eliminates dangerous residues that conventional vaccines have. In this work, they reported that the design and engineering of a BCG-coated microneedle vaccine patch improved intradermal delivery of the vaccine. The microneedles induced a robust cell-mediated immune response in the lungs and spleen of guinea pigs (100). Levin evaluated the device (MicronJetTM) that can inject antigens close to the skin's dendritic cells. A dose-sparing study was in 280 healthy adults using trivalent virosomal adjuvant influenza vaccine. The MicronJetTM provides better response in comparison with the conventional vaccine (101).

Damme et al. studied the same device (MicronJetTM) and a low-dose influenza vaccine

delivered intradermally with microneedles. They showed that this gadget has a similar dose of intramuscular vaccination. The device shows that microneedles are effective, secure, and reliable (102). Microneedle delivery of nucleic acids as plasmid DNA (pDNA) into the skin is a potential method for the clinical management of genetic skin cutaneous cancers and diseases and intracutaneous genetic immunization. Zhu W et al. developed a microneedle to increase the immunogenicity of conventional influenza vaccines. A new 4M2e-tFliC fusion protein construct containing M2e sequences from different subtypes was generated and loaded on microneedles. The results demonstrated that mice receiving a conventional inactivated vaccine followed by the treatment with microneedles boost could better maintain the humoral antibody response than the only the use of conventional vaccine alone. Compared with an intramuscular injection, the mice with receiving microneedles showed significantly enhanced cellular immune responses, titers, hemagglutination-inhibition (HAI) and neutralization titers. The results of this study demonstrate that receiving 4M2e-tFliC microneedles of carboxymethylcellulose boosting immunization after the conventional influenza vaccine prime is an efficient and speedy approach to acquire extra protection against homologous influenza virus infection and cross protection against heterologous viral strains (103).

Mikszta et al. studied the delivery of naked pDNA into skin using microneedles. They used the technique "dip and scrape" *in vivo* to create microabrasions. They reported that, in a mouse model, the topical gene transfer was increased 2800-fold by microneedles, in contrast with topical application alone. Using DNA plasmid encoding hepatitis B surface antigen, microneedles induced stronger immune responses compared with hypodermic injection and required fewer immunizations for full seroconversion. The importance of this study was that the DNA vaccine delivery generated an immune response using the microneedles. It also established the feasibility of using blunt-tipped microneedles to scrape the skin for increased delivery (104).

Stimulation of CD4+ and CD8+ T cell immunity it's an inefficient process. The delivery of prophylactic vaccines is mainly mired by low transfection efficacy, poor immunogenicity, and safety issues from the materials employed. For this reason, to improve the immunogenicity Huu et al.

developed a polyelectrolyte multilayer assembly on microneedles loaded with DNA vaccine. The charge reversal pH-responsive copolymer, composed of oligo (sulfamethazine) -b-poly (ethylene glycol) -bpoly (amino urethane) (OSM-b-PEG-b-PAEU), was used as a triggering layer in the polyelectrolyte multilayer assembly on microneedles. Charge reversal characteristics of this copolymer exhibit is a positive charge at low pH (pH 4.03) and negative charge (pH 7.4), allowing the facile assembly and disassembly of polyelectrolyte multilayers. The microneedles can delivery of a DNA vaccine to antigen-presenting cells and their subsequent stimulation of CD4+ and CD8+ T cell immunity. In vivo immunization of BALB / c mice, the results demonstrated that targeted delivery of a DNA vaccine encoding AB fusion protein to antigen present cells induced a robust antigen-specific immune response (105).

Chen et al. Evaluated the potential of a chitosan microneedle patch loaded with antigen ovalbumin (OVA at a dosage of 200 µg) for low-dose immunization. This system comprises antigenloaded chitosan microneedles made of polyvinyl alcohol/polyvinylpyrrolidone. The microneedles allow a sustained release OVA for up to 28 days. We found that rats immunized with microneedles had persistently high antibody levels for 18 weeks, which were significantly higher than intramuscular injection of OVA at a dosage of 500 µg, demonstrating at least 2.5-fold dose sparing. Moreover, OVA-encapsulated chitosan MNs had superior immunogenicity to OVA plus chitosan solution, indicating that MN-based delivery and prolonged skin exposure can further enhance chitosan's adjuvanticity (106).

Insulin and macromolecules

Microneedles have been proposed to be a kind of delivery system that permits the entry of drugs, therapeutic proteins, and insulin with minimal skin invasiveness (48). Zhang et al. investigated the utility of solid microneedle (150 mm in length) in enhancing transdermal peptide delivery. Four model peptides were used: (Gly-Gln-Pro-Arg [tetrapeptide-3,456.6Da], Val-Gly-Val-Ala-Pro-Gly [hexapeptide, 498.6Da], AC-Glu-Glu-Met-Gln-Arg-Arg-NH₂ [acetyl hexapeptide-3,889Da], and Cys-Tyr-Ile-Gln-Asn-Cys-Pro-Leu-Gly-NH₂ [oxytocin, 1007.2Da]). The penetration was evaluated using porcine ear skin with Franz diffusion cells. Peptide permeation across the skin was significantly enhanced by microneedle

pretreatment, and permeation rates were dependent on peptide molecular weights. They concluded that solid microneedles are effective to enhance skin delivery of peptides (107).

Ling et al. made dissolving microneedles with starch and gelatin for rapid and efficient transdermal delivery of insulin in diabetic rats. The dissolution of the microneedles was 5 min after application. They quickly released their encapsulated payload into the skin. Pharmacodynamic and pharmacokinetic results showed a comparable hypoglycemic effect in rats receiving insulin-loaded microneedles and a subcutaneous injection of insulin. The bioavailability of insulin was around 92%, demonstrating that insulin has pharmacological activity encapsulation and release from microneedles. Storage stability was more than 90% of the insulin even after storage at 25°C or 37°C for 1 month, generally, the insulin should be stored at 4°C-8°C. These results supported that encapsulation of biomolecules in microneedles has great potential for transdermal delivery of diverse biomolecules (108). Other studies using microneedles with insulin lispro showed that they can deliver insulin effectively to treat diabetes (108-110). Yu et al. To reduce the painful subcutaneous injection of insulin, developed biodegradable microneedle patches that fabricated from 3-aminophenyl boronic acid-modified and crosslinked alginate was prepared for transdermal drug delivery of insulin. Microneedles exhibited a strong mechanical strength to penetrate the skin and good degradability to release the insulin. In vivo had been applied the microneedles using diabetic mice. The insulin in microneedles maintain the high pharmacological activity, having a sustained hypoglycemic effect in diabetic mice with relative pharmacologic availability and bioavailability of insulin from microneedle were $90.5\pm6.8\%$ and $92.9\pm7\%$ in mice compared with that of subcutaneous injection route with same insulin dose (111).

Drug Delivery

Statin or HMG-CoA reductase inhibitor. Serrano et al. used a dermaroller to increase the permeation of sodium pravastatin patch. They evaluated pravastatin penetration using microneedles of two different lengths (250 μ m and 2250 μ m). The result gives a therapeutic dosage equivalent of a 10 mg tablet with the constant drug release. The authors state the advantage of the route to be avoidance of the

presystemic hepatic loss of the drug that is expected after oral administration (112).

The μ-Opioid receptor antagonist. Wermeling et al. conducted a clinical study using microneedles for delivering naltrexone formulated in a transdermal patch. Application of the naltrexone patch in the skin over 72 h produced undetectable drug plasma levels. However, pretreatment of skin with microneedles achieved steady-state plasma concentrations within 2 h of patch application, and they were maintained for at least 48 h. The microneedles arrays were painless upon administration and did not damage the skin during insertion, and microneedles were not broken off in the skin (6, 113).

Anti-cancer agents. Cancer is the principal cause of death worldwide. Localized intratumoral anti-cancer injections can be an attractive treatment, but conventional hypodermic injections result in poor distribution of the drug in the tumor and leakage of the drug into the systemic circulation. For these reasons, Ma et al. developed coated microneedles for delivery of intratumoral anti-cancer Moreover, PLGA nanoparticles encapsulating doxorubicin were prepared and coated on microneedles. The hypodermic injection of different volumes into porcine buccal tissue confirmed significant leakage (about 25% of the injected 80 µl). This study confirmed that microneedles can deliver anti-cancer drugs on localized oral cancers reducing the percentage of leakage. (114). Vemulapalli et al. investigated transdermal iontophoretic delivery (0.4 mA/cm² applied for 60 minutes) of methotrexate, alone or in combination with maltose microneedle array, in vivo and in vitro using rats. Delivery was enhanced with iontophoresis and microneedles (in vitro and in vivo). A synergistic 25-fold enhancement of in vivo drug delivery using microneedles and iontophoresis was obtained compared with each physical enhancer alone. This result is due to the fact that both methods are penetration enhancers and each of them has different mechanisms one based is in microabrasions (microneedles) and the other by the use of electric current. (6, 115).

Calcium channel blockers. Kaur et al. studied the effect of microneedle rollers and stainless-steel microneedles on the percutaneous penetration of verapamil hydrochloride and amlodipine. Verapamil is a calcium channel blocker. It regulates hypertension by decreasing myocardial contractility,

heart rate, and impulse conduction. Amlodipine is a calcium channel blocker too, used to treat hypertension and ischemic heart disease. In vitro passive diffusion studies of verapamil and amlodipine across the skin using vertical static Franz diffusion cells with porcine ear skin as a membrane was low. They conclude that it is possible to develop transdermal microneedle patches for these drugs because the use of microneedles increased the diffusion of verapamil and amlodipine (116). In another study, Kolli and Banga characterized maltose microneedles and evaluated the capacity to enhance transdermal drug delivery of nicardipine hydrochloride in vitro and in vivo in rats. Microneedles penetrated the skin creating microchannels, and nicardipine hydrochloride in vitro was increased after pretreatment (flux 7.05 mg/cm²·h) compared with untreated the skin (flux 1.72 mg/cm² h). The same case was observed in vivo (117). The advantages of using microneedles are that by avoiding the first hepatic metabolism, more drug enter to the bloodstream, allowing it to do its therapeutic effect, and besides avoiding side effects due to the dose presented by the tablets.

Heparin. This drug is used to prevent clots in patients with certain medical conditions or in medical procedures that increase the chance of their formation. Heparin has another use to prevent the growth of clots formed in blood vessels, but cannot be used to decrease the size of these clots. Gomma et developed a laser-engineered dissolving microneedle array made with aqueous blends of 15% w/w poly (methylvinylether-co-maleic anhydride) for transdermal delivery of nadroparin calcium. The microneedles were loaded with 630 IU of nadroparin. The technique "poke and release" was used. Microneedles allowed permeation of 10.6% of the nadroparin over a 48-h study period. The cumulative amounts of nadroparin permeated at 24 h and 48 h were 12.28 \pm 4.23 IU/cm² and 164.84 \pm 8.47 IU/cm², respectively. Skin permeation of nadroparin can be modulated by the length and array density of microneedles (118). The microneedles have a good potential for transdermal delivery of nadroparin at a low cost.

Ito et al. generated dissolving needles loaded with a polysaccharide with heparin. They used three polymers, chondroitin, dextran, and dextrin, as the base of the microneedles. The insertion was in rat skin with doses of 25, 50, and 100 IU/kg. The values of C_{max} were 0.40 ± 0.03 , 0.46 ± 0.03 , and

 0.47 ± 0.06 IU/ml. The physiological availabilities were 102.3%, 81.5%, and 97.7%, respectively (119). These results confirm the use of biodegradable microneedles for the percutaneous administration of polysaccharide drugs like heparin. These studies demonstrated that it is possible to load the microneedles with high molecular weight biomolecules and allow the passage of these through the skin, maintaining the advantages of transdermal administration.

Local anesthesia. Lidocaine is indicated as a local anesthetic. It is used on the intact skin for minor surgery (superficially) and preparation for infiltration anesthesia. The poor patient acceptance of injections, principally in the case of the pediatric population, makes microneedles an interesting choice to eliminate pain associated with conventional needles. They can give a drug administration, eliminating the pain associated with conventional needles.

Caffarel et al. developed biodegradable microneedles loaded with lidocaine. Their results suggested that microneedles can dissolve quickly in the skin around 15 min upon application. They concluded that microneedles are a viable alternative to administer lidocaine in pediatric patients, avoiding needle phobia for children (120).

Central nervous system stimulant (CNSS). Caffeine is a CNSS drug, but another use of caffeine is to treat the apnea of prematurity in infants. Nevertheless, the only pharmaceutical preparation of caffeine for treatment of apnea in infants is an intravenous infusion and an oral dosage form. For these reasons, Caffarel developed biodegradable et al. poly(methylvinylether/maleic anhvdride) microneedles using caffeine. They obtained a gradual and sustained increase in plasma caffeine concentration during a period of 24 h, but at 2h, they have therapeutic concentrations. Their results suggested that a single application of biodegradable microneedles has a continuous delivery of caffeine. maintaining therapeutic concentration for more than 24 h. The advantage is in simplifying treatment (2–3 days dosing) instead of the current dosage (once daily) oral or intravenous route. Microneedles can benefit neonates who commonly cannot tolerate the enteral administration of caffeine, particularly in the early postnatal period. Another advantage is that microneedles could be easy to remove in cases of suspected toxicity and it is not necessary to apply

intravenously that it is sometimes complicated in neonates to apply an intravenous and sometimes causing more damage if it is not done correctly (120).

Anti-obesity. Manita et al. developed 500 μm long microneedles loaded with caffeine made of Carboxymethylcellulose CMC, polyvinyl pyrrolidone, polyvinyl alcohol and applied to obese mice for weight loss (121).

Cosmetology Applications

The application of microneedles into human skin emerged as a popular tool in the cosmetic area because microneedles are useful in bettering conditions like seborrheic keratosis (122), scars (125), striae, anti-aging, wrinkles, or depigmentation (123). The original instrument used is the "dermaroller," which consists of a handle with a cylinder with stainless steel needles (0.5-2 mm length). Treatment with a dermaroller requires four to eight weeks to have the desired effect on the skin. Microneedles or dermaroller treatment is popular in the world, not only in the treatment of post-acne but also in anti-aging therapy with no sequelae (123).

Burn scars. The treatment is a simple method for treating burn scars using microneedles in comparison with laser treatments. The procedure is safe and applicable in areas where a laser cannot be used (124). Microneedle procedures effectively manage hypertrophic scars. Microneedles provoke the collagen fiber rearrangement in scar tissue (125). An important advantage of using microneedles compared to a laser is that the treatment is cheaper.

Acne scars and stretch marks. The use of microneedles is a simple procedure. The area to be treated is previously anesthetized (45 min to 1 h). After that, the dermaroller is passed 15 to 20 times in horizontal, vertical, and oblique directions. The pretreated zone is cleaned with wetted saline pads. The complete procedure continues for 15 to 20 min. A minimum of six weeks is recommended between two treatments for new natural collagen to form. Approximately 3 to 4 treatments are needed for moderate acne scars (126). Skin needling is a simple and cost-effective technique used for the treatment of acne scarring (125, 127). The dermabrasion used to improve the quality of the skin is based on the "ablation" (destruction or injury of the superficial layers of the skin), the dermabrasion induced by microneedles generate an equilibrate

proliferation facilitates the repair of the skin without leaving scars that other techniques cannot do efficiently.

Anti-aging, wrinkles, depigmentation or pigmentation. Microneedle fractional radiofrequency (RF) is a noninvasive method. Growth factors can be used as a novel anti-aging treatment. Seo et al. evaluated the effectiveness and security of microneedle RF for rejuvenation using stem cells that have many growth factors and cytokines. They concluded that RF is safe and effective in skin rejuvenation and has better results combined with stem cells (128). The therapies with percutaneous collagen offer to rejuvenate and repair the skin appearance without risk of depigmentation (129).

In the case of melasma that is a common acquired symmetrical hypermelanosis characterized by irregular light to-dark-brown macules and patches on sun-exposed areas of the skin. The tranexamic acid (TXA) is administered orally, and locally or via localized intradermal injections, results in lower melasma severity. TXA seems to inhibit the synthesis of melanin by interfering with the interaction of melanocytes and keratinocytes. Also, TXA can reverse the abnormal dermal changes induced by melasma, such as increased vasculature. Machekposhti et al. generated microneedles (1200 μm height, 280 μm base width, 36 microneedles in the array) of PVP and methacrylic acid loaded with tranexamic acid. These microneedles had adequate properties to be applied to the skin and have a release in the TXA with the possibility of being an alternative for the treatment of melasma (130). Another case of pigmentation is the seborrheic keratosis or senile lentigo are commonly seen on people >50 years of age, the treatment is with alltrans retinoic acid (ATRA), ATRA has some drawbacks such as its poor water solubility and photostability, and skin irritation reactions limit its topical use. Because the skin permeability is relatively low Sachiko et al. developed an ATRAloaded microneedle patch (ATRAN-MN) to increase the permeability of ATRA and microneedle patch loaded with retinoic acid. ATRA-MN was applied to the lesion site of each subject for 6 h once per week for 4 weeks. The use of microneedles did not induce severe local or systemic adverse effects. The treatment with microneedles is promising as a safe and effective therapy for seborrheic keratosis and senile lentigo (131).

Alopecia areata. Alopecia areata (AA) is a chronic autoimmune disease that may be mediated by T cells, affecting hair follicles and sometimes nails, but the mechanism is not clear. The disease can present as a single demarcated patch (hair loss zones), as the total hair loss (called alopecia), and as the loss hair of the head and body (called alopecia universalis) (132). The AA is usually difficult to treat. Topically applied corticosteroids are useful but painful in large

patches (hair loss zones) because they are injected. Microneedles are a good option to apply corticosteroids in large hair loss zones without pain. Moreover, they increase blood supply to the hair follicles providing them with more nutrients. Another hypothesis is that the microinjury generated by microneedles helps in recruiting and inducing growth factors (133). All the applications of microneedles are summarized in Table 2.

Table 2. Applications of microneedles

	Table 2. Applications of microneedles Modical and Pharmacoutical Applications			
Medical and Pharmaceutical Applications Study Outcome Reference				
Transfer-molded Wrappable Microneedle Meshes for Perivascular Drug Delivery	Developed a wrappable MN mesh of poly(lactic-co-glycolic acid) (height of 640 µm) to deliver the anti-proliferative drug into an injured blood vessel for IH reduction with minimal mechanical stress (98).	Lee et al. (2017)		
A microneedle electrode array on flexible substrate for long-term EEG monitoring	The use of microneedles (silicone height of $190 \mu m$) has been used that could be capable of EEG monitoring. Microneedles can be adapted to the skin providing robust contact with the skin (96).	Renxin et al. 2017		
Fenestrated microneedles for ocular drug delivery	They develop fenestrated microneedles (lengths $500 - 1500$ µm) which serve as reservoirs for passive delivery, the capacity of the microneedles can up to five-fold relative to solid microneedles (83).	Omid et al. 2016		
Microneedle for Minimally Invasive and Painless Blood Sampling	Develop microneedles to obtain biochemical information (1.8 mm, an inner diameter of 60 μ m, an outer tip of 100 μ m, and a 60 ° bevel angle), that can extract blood volumes up to 840 μ L (88).	Dae et al. (2018)		
Microneedle array electrodes for continuous glucose monitoring sensors	The use of microneedles eliminates this interference to detect glucose in comparison with other devices glucose (89).	Sanjiv et al. (2017)		
Silicon microneedles for deep brain drug infusion.	Microarray prepared on silicon for infusion of drugs in the brain to identify connections and neuronal activities (93).	Lee et al. (2015)		
Targeted delivery of antiglaucoma drugs to the supraciliary space using microneedles.	Use of hollow microneedles loaded with a drug for the treatment of glaucoma applied in the eye (space intraciliary) as a new alternative for the treatment (82).	Kim et al. (2014)		
Nitrogen-incorporated ultra- nanocrystalline diamond microneedle arrays for electrochemical biosensing.	Nitrogen-incorporated ultra-crystalline diamond-coated titanium alloy microneedle arrays can detect electrochemical signals (dopamine and uric acid) (84).	Skoog et al. (2015)		
Hollow microneedles for minimally invasive blood extraction. Microscopic gel-liquid interfaces supported by a hollow microneedle array for voltammetric drug detection.	Hollow microneedles can be incorporated with other technologies, such as biosensors and fluidic chips, to create blood analysis system (85-86).	Li et al. (2013) Vázquez et al. (2014)		
Influence of microneedle shapes on skin penetration for continuous in vivo glucose monitoring. Microneedle-based automated therapy for diabetes mellitus.	Hollow microneedles of silicon that can determine the amount of blood glucose through a sample of interstitial fluid (87-88).	Chua et al. (2013) Khanna et al. (2008)		
Microneedle biosensor for real-time electrical detection for in situ cancer diagnosis.	Microneedle sensor has high-resolution imaging combined with electrical real-time detection of cancer. The system can be a new platform for detection of cancer (89).	Keum et al. (2015)		

Table 2. Continued...

Preliminary technological assessment of microneedle-based dry electrodes for biopotential monitoring in clinical examinations.	Microneedle-based dry electrodes tested in ECG, EEG, and EMG are comparable to wet electrodes in static conditions and better in ECG dynamic conditions (92).	Forvi et al. (2012)
Microneedle with charge reversal pH- sensitive copolymers improve antigen presenting cells-homing DNA vaccine delivery and immune responses	Microneedles with charge reversal copolymer that can stimulate the CD4+ and CD8+ T cell immunity. <i>In vivo</i> demonstrated the delivery of a DNA vaccine encoding Aβ fusion protein to antigen present cells induced a robust antigen-specific immune response (105).	Huu et al. (2018)
A boosting skin vaccination with dissolving microneedle patch encapsulating M2e vaccine broadens the protective efficacy of conventional influenza vaccines	The results of this study demonstrate that receiving 4M2e-tFliC microneedles of carboxymethylcellulose boosting immunization after the conventional influenza vaccine, the array has 100 microneedles (250 and 650 µm, diameter and length, respectively) (103).	Zhu W et al. (2017)
Chitosan microneedles patch to enhancing immunogenicity of antigens	A strong and persistent antibody responses for at least 18 weeks by microneedles loaded with OVA and resulted in at least a 2.5-fold antigen dose reduction (106).	Chen et al. (2017)
Microneedles of alginate and hyaluronate for transdermal delivery of insulin	Microneedles (650 μ m) patches made with alginate and hyaluronate loaded with insulin. The relative pharmacologic availability and relative bioavailability of insulin from microneedle were 90.5±6.8% and 92.9±7% in mice (111).	Yu et al. (2017)
Anti-obesity effect of caffeine-loaded in microneedle patch	Microneedles of PVP, PVP, and PVA (500 μ m long) loaded with caffein, has a weight loss of 12.8 \pm 0.75% in obese C57BL/6J mice (121).	Manita et al. (2017)
A transcutaneous immunization system by microneedle array for soluble and particulate antigens.	Use of microneedles for permeation of peptides with different molecular weights. Microneedles can remarkably enhance the transdermal delivery of all hydrophilic peptides and be used for safe vaccination. (79).	Matsuo et al. (2002)
Clinical study of transcutaneous influenza vaccination using a dissolving microneedle patch.	Microneedles of hyaluronic acid with hemagglutinins of influenza for vaccination, inducing a strong immune response (54).	Hirobe et al. (2015)
Microneedle patch containing the measles vaccine is immunogenic in non-human primates.	Vaccination with a microneedle loaded with an antibody of measles that was immunogenic in non-human primates (94).	Edens C. et al. (2015)
Bacillus Calmette-Guérin (BCG) vaccination using a microneedle patch.	Vaccination patch coupled with microneedles, preloaded with BCG for the treatment and prevention of tuberculosis (95).	Hiraishi et al. (2011)
Clinical evaluation of a microneedle device for the intradermal delivery of	Silicon microneedles using 0.45 mm long (MicronJet TM) for injecting the influenza antigens very close to the skin's	Levin et al. (2014)
an influenza vaccine. Safety and efficacy of a microneedle device for influenza vaccination in healthy adults.	dendritic cells. MicronJet TM gives a superior response to influenza vaccination and warrants further clinical evaluation. The microneedle injection device is effective, safe, and reliable (96-97).	Damme et al. (2009)
Droplet-born air blowing: made dissolving microneedle.	Developed biodegradable microneedles by the blowing method that are loaded with insulin (57).	Dong et al. (2013)
Coated microneedles for transcutaneous delivery of live virus vaccines.	Silicon microneedles coated by a method of spray, containing adenovirus antigen and virus Ankara that stimulated CD8 cells of the immune system (60).	Vrdoljak et al. (2012)
Enhanced delivery of hydrophilic peptides in vitro by transdermal	An array of solid microneedles of 150 µm in length was used to make the skin more permeable (pig's ear) and facilitate	Zhang et al. (2014)
microneedle pretreatment. Improved genetic immunization via micromechanical disruption of skin.	transport of peptides (61, 98).	Mikszta et al. (2002)

Table 2. Continued...

Development and a characterization of a pravastatin transdermal patch coupled with solid microneedles.	Used a dermaroller [®] to increase the permeation of sodium pravastatin formulated in a transdermal patch. The pravastatin penetration using microneedles (250 μm and 2250 μm of lengths), and obtained viable results to achieve a therapeutic equivalent dose of a 10 mg tablet (103).	Serrano et al. (2014)
Transdermal delivery of microneedles to the skin in medication to humans.	Clinical study of microneedles for enhanced delivery of naltrexone in a transdermal patch. This human proof-of-concept study demonstrated the systemic administration of hydrophilic medication using a microneedle for enhanced transdermal delivery (104).	Wermeling et al. (2008)
Drug-coated microneedles for treatment of oral carcinomas.	Development of coated microneedles for direct and minimally invasive intratumoral delivery of anti-cancer drugs (105).	Ma et al. (2015)
The effect of iontophoresis and microneedles for transdermal delivery of methotrexate.	Synergistic 25-fold enhancement of delivery in vivo in combination (microneedle-iontophoresis), compared to each one alone (106).	Vemulapalli et al. (2008)
Microneedle delivery of verapamil hydrochloride and amlodipine besylate.	Microneedles increased penetration of verapamil hydrochloride and amlodipine. It is possible to generate transdermal microneedle patches for these drugs (107).	Kaur et al. (2014)
Characterization of solid maltose microneedles and their use for transdermal delivery.	Increase transdermal drug delivery of nicardipine hydrochloride <i>in vitro</i> and <i>in vivo</i> across hairless rat skin (108).	Kolli et al. (2008)
Laser-engineered dissolving microneedles for active transdermal delivery of nadroparin calcium.	Development of laser-engineered dissolving microneedle arrays fabricated by 15% w/w poly (methylvinylether-comaleic anhydride) with nadroparin calcium. The microneedles offer immense potential as a relatively low-cost functional delivery system (109).	Gomma et al. (2012)
Evaluation of dissolving microneedles containing low molecular weight heparin in rats.	The use of biodegradable microneedles for the percutaneous administration of polysaccharide drugs like heparin (110).	Ito et al. (2008)
Potential of hydrogel-forming and dissolving microneedles for use in pediatric populations.	Application of biodegradable microneedles allows continuous delivery of caffeine into systemic circulation, maintaining the therapeutic concentration for more than 24h. Microneedles are a viable alternative for pediatric administration (111).	Caffarel Salvador et al. (2015)
	Cosmetology Applications	
Study Microneedle for transdermal delivery of tranexamic acid	Outcome Microneedles of PVP and methacrylic acid loaded with tranexamic acid. They concluded that polymer microneedle as a highly efficient method for delivering tranexamic acid for treat melasma (130).	Reference Machekposhti et al. (2017)
Clinical study of a retinoic acid- loaded microneedle patch for seborrheic keratosis or senile lentigo	Microneedles of sodium hyaluronate (800 µm length) loaded with ATRA to treat seborrheic keratosis and senile lentigo. The treatment with microneedles is promising as a safe and effective therapy for seborrheic keratosis and senile lentigo (131).	Sachiko et al. (2017)
Management of hypertrophic scar after burn wounds using microneedles. Skin needling as a treatment for acne scarring.	Use of microneedles for treating acne scars in which the skin is prepared using local anesthesia and a dermaroller passed several times in different directions. Skin needling is a simple technique for treating acne scars (115-117).	Kim et al. (2009) Doddaballapur et al. (2009) Harris (2015)

Table 2. Continued...

Skin rejuvenation by microneedle	Fractional RF was used with microneedles in the skin using	Seo et al. (2013)
fractional RF and a human stem cell	stem cells for skin rejuvenation. The therapy with	Aust et al. (2008)
conditioned medium. Percutaneous	percutaneous collagen offers a modality with which to	
collagen induction: minimally	rejuvenate (118,119).	
invasive skin rejuvenation without		
risk of hyperpigmentation.		
Alopecia areata (AA): successful	Application of corticosteroids using microneedles,	Chandrashekar et
outcome with microneedles and	promoting blood supply to hair follicles for treatment of AA.	al. (2014)
triamcinolone acetonide AA: a new	Helps in recruiting growth factors and inducing hair growth	Alsantali (2011)
treatment plan.	(132,133).	

CONCLUSIONS

Microneedles allow painless insertion with minimum tissue damage, better control over the dosage of the drug, do not generate infectious waste and are more acceptable and comfortable for patients. Consequently, microneedles have been growing in fields of drug development, therapeutics and cosmetology. It is possible to administer peptides, avoiding multiple daily injections like the case of insulin. In addition, the use of microneedles is growing in medicine. They are being used in the diagnosis, treatment for glaucoma, and in monitoring bio-signals. Finally, the use of microneedles has been shown to be very useful for the treatment of alopecia, anti-aging, and scars. Due to all the advantages and alternatives that the use of microneedles offers, increased research on them concerning their applications in many fields of health and beauty is likely.

ACKNOWLEDGMENTS

Dr. Escobar-Chávez wants to acknowledge Cátedra PIAPIC 1619, PAPIIT IT 200218 y PAPIME PE 201418. M. Sci. Serrano Castañeda wants to acknowledge a CONACyT grant with reference number 272133.

REFERENCES

- 1. Chen Y, Quan P, Liu X, Wang M, Fang L. Novel chemical permeation enhancers for transdermal drug delivery. AJPS, 2014; 9: 51-65.
- Escobar Chávez JJ. Study of the penetration through the skin of naproxen sodium using promoters (Azone and Transcutol), and chlorhexidine

- digluconate by iontophoresis. [Doctoral Thesis]. State of Mexico: UNAM FESC, 2006.
- 3. Bhowmik B, Chiranjib, Chandira M, Jayakar B, Sampath B. Recent advances in transdermal drug delivery system. Int J Pharm Tech Res. 2010, 2(1): 68-77.
- 4. Shingade G, Aamer Q, Sabale P, Grampurohit N, Gadhave M, Jadhav S, Gaikwad D. Review on: Recent trend on transdermal drug delivery system. JDDT. 2012, 2(1): 66-75.
- Soler Ranzani L. Development and Evaluation Biopharmaceutical of a Transdermal System of Alprazolam. [Doctoral Thesis]. Barcelona Spain: University of Barcelona, Faculty of Pharmacy; 2006.
- Escobar Chavez JJ, Bonilla Martinez D, Villegas Gonzales M, Molina Trinidad E, Casas Alancaster N. Microneedles: A Valuable Physical Enhancer to Increase Transdermal Drug Delivery. J. Clin. Pharmacol, 2011; 51: 964-977.
- 7. Hadgraft J. Skin deep. Eur J Pharm Biopharm, 2004; 58: 291-299.
- 8. Barry B. Lipid-protein-partitioning theory of skin penetration enhancement. J Control Release, 1991; 15(3): 237-248.
- 9. Barry B, Williams A. Penetration enhancers. Adv Drug Deliv Rev, 2003; 56: 603-618.
- Escobar-Chávez JJ, Quintanar Guerrero D, Ganem-Quintanar A. In vivo skin permeation of sodium naproxen formulated in Pluronic F-127 gels: Effect of azone[®] and transcutol[®]. Drug Develop Ind Pharm, 2005; 31:447-454.
- 11. Grimnes, S. Pathways of ionic flow through human skin in vivo. Acta DermVenereol, 1984; 64:93-98.
- Escobar-Chávez JJ, Bonilla-Martínez D, Villegas-González MA, Revilla-Vázquez AL. The electroporation as an efficient physical enhancer for transdermal drug delivery. J Clin Pharmacol, 2009; 49(11):1262-83.
- 13. Escobar-Chávez JJ, Merino-Sanjuán V, López-Cervantes M, Rodríguez Cruz IM, Quintanar

- Guerrero D, Ganem-Quintanar A. The use of iontophoresis in the administration of drugs through the skin for smoking cessation. Curr Drug Discov Technol, 2009; 6(3):171-185.
- 14. Preat V, Vanbever R. Skin electroporation for transdermal and topical delivery. Adv Drug Deliv Rev, 2004; 56: 659-674.
- 15. Zhou Q, Chen Z, Wang Y, Yang F, Lin Y, Liao Y. Ultrasound-Mediated Local Drug and Gene Delivery Using Nanocarriers, Biomed Res Int, 2014; 2014:1-13.
- Escobar-Chávez JJ, Bonilla Martínez D, Villegas González M. Sonophoresis: An alternative physical enhancer to increase transdermal drug delivery. Current technologies to increase the transdermal drug delivery. Bentham Science Publishers, 2010; 53-77.
- 17. Escobar Chávez JJ, Bonilla Martínez D, Villegas González A, Rodríguez Cruz IM, Domínguez Delgado CL. The use of sonophoresis in the administration of drugs through the skin. J Pharm Pharm Sci, 2009; 12(1): 88-115.
- 18. Prausnitz Mark R. Microneedles for transdermal drug delivery. Adv Drug Deliver Rev, 2004; 56: 581-587.
- 19. Serrano Castañeda P, Escobar-Chávez JJ, Morales Hipólito E, Domínguez Delgado C, Abrego Reyes V. Microneedles and Transcutol® as transdermal penetration enhancers of sibutramine formulated in a transdermal patch. Revista Cubana de Farmacia. 2013; 3(47):289-299.
- Davidson A, Al-Qallaf B, Bhusan Das D. Transdermal drug delivery by coated microneedles: Geometry effects on effective skin thickness and drug permeability. Chem Eng Res Des, 2008; 86: 1196-1206.
- 21. Shwayder T, Akland T. Neonatal skin barrier: structure, function, and disorders. Dermatol Ther, 2005; 18: 87-103.
- 22. Delgado-Charro B, Guy H. R. Effective use of transdermal drug delivery in children. Adv. Drug Delivery Rev, 2014; 73: 63-82.
- 23. Sinko P. Martin's physical pharmacy and pharmaceutical sciences. 6^a. Ed. Philadelphia: Lippincott Williams & Wilkins; 2010.
- 24. Planz V, Lehr C.M, Windbergs M. In vitro models for evaluating safety and efficacy of novel technologies for skin drug delivery. J Control Release, 2016;242:89-104.
- 25. Wilkes GL, Brown IA, Widnaurer RH. The Biomechanical Properties of Skin. Bioeng, 1973; 6: 452.
- Honeyman J, Maira M, Valdés P, Pérez P. Dermatology. Santiago de Chile: Tecnoprint Ltda; 1997. p. 1-10.
- 27. Merk HF, Jugert FK, Frankenberg S. Biotransformations in the skin in

- Dermatotoxicology. USA: Taylor & Francis; 1996. p. 61-73.
- 28. Escalona Pérez E. Dermatology. México, D.F: Impresiones Modernas; 1975. p. 9-15.
- Majella E. Skin penetration enhancers. Int J Pharm, 2013; 447: 12-21.
- 30. Kneep VM, Hadgraft J, Guy RH. Transdermal drug delivery: Problems and possibilities. Crit Rev Ther *Drug Carrier* Syst, *1987*; *4:13–37*.
- 31. Bolzinger M, Briançon S, Pelletier J, Chevalier Y. Penetration of drugs through skin, a complex rate-controlling membrane. Curr Opin Colloid Interface Sci, 2012; 17: 156-165.
- 32. Blume U, Massoudy L, Patzelt A, Lademann J, Dietz E, Rasulev U, Garcia N. Follicular and percutaneous penetration pathways of topically applied minoxidil foam. Eur J Pharm Biopharm, 2010; 76:450-453.
- Barry B. Drug delivery routes in skin: a novel approach. Adv Drug Deliv Rev, 2002; 54(1): S31-S41.
- 34. Alexander A, Dwivedi A., Ajazuddin, Giri T., Saraf S., Saraf S, Tripathi D. Approaches for breaking the barriers of drug permeation through transdermal drug delivery. J Control Release, 2012; 164;26-40.
- 35. Hiraishi Y, Nakagawa T, Quan Y, Kamiyama F, Hirobe S, Okada N, Nakagawa S. Performance and characteristics evaluation of a sodium hyaluronate-based microneedle patch for a transcutaneous drug delivery system. Int J Pharm, 2013; 441: 570-579.
- 36. Park JH, Allen MG, Prausnitz MR. Biodegradable polymer microneedles: fabrication, mechanics and transdermal drug delivery. J Control Release, 2005; 104: 51-66.
- 37. Donnelly R, Garland M, Morrow D, Migalska K, Sing T, Majithiya R, Woolfson D. Optical coherence tomography is a valuable tool in the study of the effects of microneedle geometry on skin penetration characteristics and in skin dissolution. J Control Release, 2010; 147:333-341.
- 38. Ma G, Wu C. Microneedle, bio-microneedle and bio-inspired microneedle: A review. J Control Release, 2017; 251:11-23.
- 39. Larrañeta E, Lutton R, Woolfson A.D, Donnelly R. Microneedle arrays as transdermal and intradermal drug delivery systems: Material science manufacture and commercial development. Mat Sci Eng R, 2016; 104:1-32.
- 40. Liu S, Jin M, Quan Y, Kamiyama F, Kusamori K, Katsumi H, Sakane T, Yamamoto A. Transdermal delivery of relatively high molecular weight drugs using novel self-dissolving microneedle arrays fabricated from hyaluronic acid and their characteristics and safety after application to the skin. Eur J Pharm Biopharm, 2014; 86: 267-276
- 41. Donnelly R, Morrow D, McCarron P, Woolfson A, Morrissey A, Juzenas P, Juzeniene A, Iani V,

- McCarthy H, Moan J. Microneedle-mediated intradermal delivery of 5-aminolevulinic acid: Potential for enhanced topical photodynamic therapy. J Control Release, 2008; 129: 154-162.
- 42. Yan G, Warner K, Zhang J, Sharma S, Gale B. Evaluation needle length and density of microneedle arrays in the pretreatment of skin for transdermal drug delivery. Int J Pharm, 2010; 391: 7-12.
- 43. BanksS, Pinninti R, Gill H, Crooks P, Prausnitz M, Stinchcomb A. Flux across microneedle-treated skin is increased by increasing charge of naltrexone and naltrexol in vitro. Pharm Res, 2008; 25: 1677-1685.
- Chen H, Zhu H, Zheng J, Mou D, Wan J, Zhang J, Shi T, Zhao Y, Xu H, Yang X. Iontophoresis-driven penetration of nanovesicles through microneedleinduced skin microchannels for enhancing transdermal delivery of insulin. J Control Release, 2009; 139: 63-72.
- 45. Harvey A, Kaestner S, Sutter D, Harvey N, Mikszta J, Pettis R. Microneedle-based intradermal delivery enables rapid lymphatic uptake and distribution of protein drugs. Pharm Res, 2011; 28: 107-116.
- 46. Wang P, Cornwell M, Hill J, Prausnitz M. Precise microinjection into skin using hollow microneedles. J Invest Dermatol, 2006; 126: 1080-1087.
- 47. Xie Y, Xu B, Gao Y. Controlled transdermal delivery of model drug compounds by MEMS microneedle array. Nanomedicine, 2005; 1: 184-190.
- 48. Kim Y, Park J, Prausnitz M. Microneedles for drug and vaccine delivery. Adv Drug Deliv Rev, 2012; 64: 1547-1568.
- 49. Martin C, Allender C, Brain K, Morrissey A, Birchall J. Low temperature fabrication of biodegradable sugar glass microneedles for transdermal drug delivery applications. J Control Release, 2012; 158(1): 93-101.
- 50. Park J, Allen M, Prausnitz M. Polymer microneedles for controlled-release drug delivery. Pharm Res, 2006; 23(5): 1008-1019.
- Liu S, Jin M, Quan Y, Kamiyama F, Katsumi H, Sakane T, Yamamoto A. The development and characteristics of novel microneedle arrays fabricated from hyaluronic acid, and their application in the transdermal delivery of insulin. J Control Release, 2012; 161: 933-941.
- Ke H, Lin Y, Hu Y, Chiang W, Chen K, Yang W, Liu H, Fu C, Sung H. Multidrug release based on microneedle arrays filled with pH-responsive PLGA hollow microspheres. Biomaterials, 2012; 33: 5156-5165.
- 53. DeMuth P, Garcia Beltran D, Ai Ling M, Hammond P, Irvine D. Composite Dissolving Microneedles for Coordinated Control of Antigen and Adjuvant Delivery Kinetics in Transcutaneous Vaccination. Adv. Funct Mater, 2013; 23: 161-172.

- 54. Hirobe S, Azukizawa H, Hanafusa T, Matsuo K, Quan Y, Kamiyama F, Katayama I, Okada N, Nakagawa S. Clinical study and stability assessment of a novel transcutaneous influenza vaccination using a dissolving microneedle patch. Biomaterials, 2015; 57: 50-58.
- 55. Indermun S, Luttge R, Choonara Y, Kumar P, du Toit L, Modi G, Pillay V. Current advances in the fabrication of microneedles for transdermal delivery. J Control Release, 2014; 185: 130-138.
- 56. Jasim Uddin M, Scoutaris N, Klepetsanis P, Chowdhry B, Prausnitz M, Douroumis D. Inkjet printing of transdermal microneedles for the delivery of anticancer agents. Int. J. Pharm. 2015.
- 57. Dong Kim J, Kim M, Yang H, Lee K, Jung H. Droplet-born air blowing: Novel dissolving microneedle fabrication. J Control Release, 2013; 170: 430-436.
- 58. Birchall JC. Microneedle array technology: the time is right but is the science ready? Expert Rev Med Devices, 2006; 3: 1-4.
- 59. Zhou C, Liu Y, Wang H, Zhang P, Zhang J. Transdermal delivery of insulin using microneedle rollers in vivo. Int J Pharm, 2010; 392: 127-133.
- Vrdoljak A, McGrath M, Carey J, Draper S, Hill A, O'Mahony C, Crean A, Moore A. Coated microneedle arrays for transcutaneous delivery of live virus vaccines. J Control Release, 2012; 159: 34-42
- 61. Zhang S, Qiu Y, Gao Y. Enhanced delivery of hydrophilic peptides in vitro by transdermal microneedle pretreatment. Acta Pharm Sin B, 2014; 4(1):100-104.
- 62. McGrath M, Vucen S, Vrdoljak A, Kelly A, O'Mahony C, Crean A, Moore A. Production of dissolvable microneedles using an atomised spray process: Effect of microneedle composition on skin penetration. Eur J Pharm Biopharm, 2014; 86: 200-211.
- 63. Ling Teo A, Shearwood C, Chye Ng K, Lu J, Moochhala S.Transdermal microneedles for drug delivery applications. Mat Sci Eng B-Solid, 2006; 132: 151-154.
- 64. Gill Harvinder S, Prausnitz Mark R. Pocketed microneedles for drug delivery to the skin. J Phys Chem Solids, 2008; 69(5-6): 1537-1541.
- 65. Bariya S, Gohel M, Mehta T, Sharma O. Microneedles: an emerging transdermal drug delivery system. J Pharm Pharmacol, 2012; 64(1): 11-29.
- 66. Ding Z, Verbaan F, Bivas Benita M, Bungener L, b, Huckriede A, van den Berg D, Kersten G, Bouwstra J. Microneedle arrays for the transcutaneous immunization of diphtheria and influenza in BALB/c mice. J Control Release, 2009; 136: 71-78.
- 67. Wang Q, Yaoa G, Donga P, Gonga Z, Lic G, Zhanga K, Wu C. Investigation on fabrication process of

- dissolving microneedle arrays to improve effective needle drug distribution. Eur J Pharm Sci, 2015; 66: 148-156.
- Martanto W, Davis S, Holiday N, Wang J, Gill H, Prausnitz M. Transdermal delivery of insulin using microneedles in vivo. Pharm Res, 2004; 21(6): 947-952.
- Martanto W, Moore J, Couse T, Prausnitz M. Mechanism of fluid infusion during microneedle insertion and retraction. J Control Release, 2006; 112(3): 357-361.
- Martanto W, Moore J, Kashlan O, Kamath R, Wang P, O'Neal J, Prausnitz M. Microinfusion using hollow microneedles. Pharm Res, 2006; 23(1): 104-113.
- Cormier M, Johnson B, Ameri M, Nyam K, Libiran L, Zhang D, Daddona P. Transdermal delivery of desmopressin using a coated microneedle array patch system. J Control Release, 2004; 97(3): 503-511
- Widera G, Johnson J, Kim L, Libiran L, Nyam K, Daddona P, Cormier M. Effect of delivery parameters on immunization to ovalbumin following intracutaneous administration by a coated microneedle array patch system. Vaccine, 2006 24(10); 1653-1664.
- 73. Verbaan F, Bal S, van den Berg D, Groenink W, Verpoorten H, Lüttge R, Bouwstra J. Assembled microneedle arrays enhance the transport of compounds varying over a large range of molecular weight across human dermatomed skin. J Control Release, 2007; 117(2), 238-245.
- 74. Nordquist L, Roxhed N, Griss P, Stemme G. Novel microneedle patches for active insulin delivery are efficient in maintaining glycaemic control: an initial comparison with subcutaneous administration. Pharm Res, 2007; 24(7): 1381-1388.
- 75. Matsuo k, Okamoto H, Kawai Y, Quan Y, Kamiyama F, Hirobe S, Okada N, Nakagawa S. Vaccine efficacy of transcutaneous immunization with amyloid β using a dissolving microneedle array in a mouse model of Alzheimer's disease. J Neuroimmunol, 2014; 266:1-11.
- Ito Y, Hagiwara E, Saeki A, Sugioka N, Takada K. Feasibility of microneedles for percutaneous absorption of insulin. Eur J Pharm Sci, 2006; 29: 82-88.
- 77. Lee J, Park J, Prausnitz M. Dissolving microneedles for transdermal drug delivery. Biomaterials, 2008; 29; 2113-2124.
- 78. Pearton M, Barrow D, Gateley C, Anstey A, Wilke N, Morrissey A, Allender C, Brain K, Birchall J. Hydrogels based on PLGA-PEG-PLGA triblock copolymers as sustained release reservoirs for the delivery of pDNA to microneedle treated human skin. J Pharm Pharmacol, 2005; 57: S12–S13.

- 79. Matsuo K, Yokota Y, Zhai Y, Quan Y, Kamiyama F, Mukai Y, Okada N, Nakagawa S. A low invasive and effective transcutaneous immunization system using a novel dissolving microneedle array for soluble and particulate antigens. J Control Release, 2002; 161: 10-17.
- 80. Arora A, Prausnitz M, Mitragotri S. Micro-scale devices for transdermal drug delivery. Int J Pharm, 2008; 364: 227–236.
- 81. Chabri F, Bouris K, Jones T, Barrow D, Hann A, Allender C, Brain K, Birchall. Microfabricated silicon microneedles for nonviral cutaneous gene delivery. Br. J Dermatol, 2004; 150: 869–877.
- 82. Kim Y, Edelhauser H, Prausnitz M. Targeted delivery of antiglaucoma drugs to the supraciliary space using microneedle, 2014; 55(11): 7387-97.
- 83. Omid K, Malik Y, Masaru P. Fenestrated microneedles for ocular drug delivery. Sensors and Actuators B: Chemical, 2016; 223: 15-23.
- 84. Sun W, Araci Z, Inayathullah M, Manickam S, Zhang X, Bruce M, Marinkovich M, Lane A, Milla C, Rajadas J, Butte M. Polyvinylpyrrolidone microneedles enable delivery of intact proteins for diagnostic and therapeutic applications. Acta Biomater, 2013; 9(8): 7767-7774.
- 85. Skoog S, Miller P, Boehm R, Sumant A, Polsky R, Narayan R. Nitrogen-incorporated ultrananocrystalline diamond microneedle arrays for electrochemical biosensing. Diam Relat Mater, 2015; 54: 39-46.
- 86. Li C, Lee C, Lee K, Jung H. An optimized hollow microneedle for minimally invasive blood extraction. Biomed Microdevices, 2013; 15(1): 17-25.
- 87. Vazquez P, Herzog G, O'Mahony C, O'Brien J, Scully J, Blake A, O'Mathuna C, Galvin P. Microscopic gel–liquid interfaces supported by hollow microneedle array for voltammetric drug detection. Sensors and Actuators B, 2014; 201: 572-578.
- 88. Dae L, Cheng G, Chunhwa I. A Three-Dimensional and Bevel-Angled Ultrahigh Aspect Ratio Microneedle for Minimally Invasive and Painless Blood Sampling. Sensors and Actuators B, 2018; 255: 384-390.
- 89. Sanjiv S., Eri T, Tony C, Wakako T, Koji S. Minimally Invasive Microneedle Array Electrodes Employing Direct Electron Transfer Type Glucose Dehydrogenase for the Development of Continuous Glucose Monitoring Sensors. Procedia Technology, 2017; 27: 208-209.
- 90. Chua B, Desai S, Tierney M, Tamada J, Jina A. Effect of microneedles shape on skin penetration and minimally invasive continuous glucose monitoring in vivo. Sensor Actuat A-Phys, 2013; 203: 373-381.

- 91. Khanna P, Strom J, Malone J, Bhansali S. Microneedle-Based Automated Therapy for Diabetes Mellitus. J Diabetes Sci Technol, 2008; 2(6):1122-1129.
- 92. Keum D, Jung H, Wang T, Shin M, Kim Y, Kim K, Ahn G, Hahn S. Microneedle Biosensor for Real-Time Electrical Detection of Nitric Oxide for In Situ Cancer Diagnosis During Endomicroscopy. Adv Healthc Mater, 2015; 4(8): 1153-8.
- 93. Strimbu K, Tavel J. What are Biomarkers? Curr Opin HIV AIDS. 2010; 5(6):463-466.
- 94. Li B, Wang J, Yang S, Zhou C, Wu M. Sample-free quantification of blood biomarkers via laser-treated skin. Biomaterials, 2015; 59: 30-38.
- 95. Forvi E, Bedoni M, Carabalona R, Soncini M, Mazzoleni P, Rizzo F, O'Mahony C, Morasso C, Cassarà G, Gramatica F. Preliminary technological assessment of microneedles-based dry electrodes for biopotential monitoring in clinical examinations. Sensor Actuat A-Phys, 2012; 180: 177-186.
- 96. Renxin W, Xiaoming J, WeiWang Z. A microneedle electrode array on flexible substrate for long-term EEG monitoring. Sensors and Actuators B. 2017; 244: 750-758.
- 97. Lee H, Son Y, Kim D, Kyung Kim K, Choi N, Yoon E, Cho I. Hyunjoo J. Lee. A new thin silicon microneedle with an embedded microchannel for deep brain drug infusión. Sensor Actuat B-Chem, 2015; 209: 413-422.
- 98. JiYong Lee, Dae Hyun Kim, Kang Ju Lee, Il Ho Seo, Seung Hyun Park, Eui Hwa Jang, Youngjoo Park, Young-Nam Youn, WonHyoung Ryu. Transfermolded wrappable microneedle meshes for perivascular drug delivery. J. Control. Release, 2017; 268: 237-246.
- 99. Edens C, Collins M, Goodson J, Rota P, Prausnitz M. A microneedle patch containing measles vaccine is immunogenic in non-human primates. Vaccine, 2015; 33: 4712-4718.
- 100. Hiraishi Y, Nandakumar S, Choi S, Lee J, Kim Y, Posey J, Sable S, Prausnitz M. Bacillus Calmette-Guérin vaccination using a microneedle patch. Vaccine, 2011; 29: 2626-2636.
- 101. Levin Y, Kochba E, Kenney R. Clinical evaluation of a novel microneedle device for intradermal delivery of an influenza vaccine: Are all delivery methods the same? Vaccine, 2014; 32: 4249-4252.
- 102. Damme P, Oosterhuis-Kafeja F, Van der Wielen M, Almagor Y, Sharon o, Levin Y. Safety and efficacy of a novel microneedle device for dose sparing intradermal influenza vaccination in healthy adults. Vaccine, 2009; 27: 454-459.
- 103. Zhu W, Pewin W, Wang C, Luo Y, Gonzalez G.X, Mohan T, Prausnitz M.R, Wang B.Z. A boosting skin vaccination with dissolving microneedle patch encapsulating M2e vaccine broadens the protective

- efficacy of conventional influenza vaccines. J. Control. Release. 2017; 261: 1-9.
- 104. Mikszta J, Alarcon J, Brittingham J, Sutter D, Pettis R, Harvey N. Improved genetic immunization via micromechanical disruption of skin-barrier function and targeted epidermal delivery. Nat Med, 2002; 8: 415-419.
- 105. Huu D, Nak W, Thavasyappan T, Giang V.H, Min L, Yue Y, Ji J, Doo S. Microneedle arrays coated with charge reversal pH-sensitive copolymers improve antigen presenting cells-homing DNA vaccine delivery and immune responses. J. Control. Release, 2018; 269; 225-234.
- 106. Chen M.C, Lai K.Y, Ling M.H, Lin C.W. Enhancing immunogenicity of antigens through sustained intradermal delivery using chitosan microneedles with a patch-dissolvable design. Acta Biomater. 2018; 65: 66-75.
- 107. Zhang S, Qiu Y, Gao Y. Enhanced delivery of hydrophilic peptides in vitro by transdermal microneedle pretreatment. Acta Pharm Sin B, 2014; 4(1): 100-104.
- 108. Ling M, Chen M. Dissolving polymer microneedle patches for rapid and efficient transdermal delivery of insulin to diabetic rats. Acta Biomater, 2013; 9(11): 8952-8961.
- 109. Gupta J, Felner E, Prausnitz M. Minimally Invasive Insulin Delivery in Subjects with Type 1 Diabetes Using Hollow Microneedles. Diabetes Technol Ther, 2009; 11(7): 471.
- 110. Gupta J, Felner E, Prausnitz M. Rapid pharmacokinetics of intradermal insulin administered using microneedles in type 1 diabetes subjects, Diabetes Technol Ther, 2011; 13(4): 451-456
- 111. Yu W, Jiang G, Zhang Y, Liu D, Xu B, Zhou J. Polymer microneedles fabricated from alginate and hyaluronate for transdermal delivery of insulin. Mater Sci Eng C Mater Biol Appl. 2017; 80: 1987-196.
- 112. Serrano Castañeda P. Desarrollo y Caracterizacion de un parche trasdérmico de pravastatina acoplado a microagujas como promotor físico de la penetración transdérmica. [Master of Science Thesis, Asesor: Dr. José Juan Escobar Chávez]. Mexico D.F: UAM Xochimilco; 2014.
- 113. Wermeling D, Banks S, Hudson D, Gill H, Gupta J, Prausnitz M, Stinchcomb A. Microneedles permit transdermal delivery of a skin-impermeant medication to humans. Proc Natl Acad Sci U S A, 2008; 105(6): 2058-2063.
- 114. Ma Y, Boese S, Luo Z, Nitin N, Gill H. Drug coated microneedles for minimally-invasive treatment of oral carcinomas: development and in vitro evaluation. Biomed Microdevices, 2015; 17(2): 44.
- 115. Vemulapalli V, Yang Y, Friden PM, Banga AK. Synergistic effect of iontophoresis and soluble

- microneedles for transdermal delivery of methotrexate. J. Pharm Pharmacol, 2008; 60(1): 27-33.
- 116. Kaur M, Ita K, Popova I, Parikh S, Bair D. Microneedle-assisted delivery of verapamil hydrochloride and amlodipine besylate. Eur J Pharm Biopharm, 2014; 86(2): 284-291.
- 117. Kolli C, Banga A. Characterization of solid maltose microneedles and their use for transdermal delivery. Pharm Res, 2008; 25(1): 104-113.
- 118. Gomaa Y, Garland M, McInnes , El-Khordagui L, Wilson C, Donnelly R. Laser-engineered dissolving microneedles for active transdermal delivery of nadroparin calcium. Eur J Pharm Biopharm, 2012; 82(2): 299-307.
- 119. Murakami A, Maeda T, Sugioka N, Takada K. Evaluation of self-dissolving needles containing low molecular weight heparin (LMWH) in rats. Int J Pharm, 2008; 349(1-2): 124-129.
- 120. Caffarel Salvador E, Tuan Mahmood T, McElnay J, McCarthy H, Mooney K, Woolfson A, Donnelly R. Potential of hydrogel-forming and dissolving microneedles for use in pediatric populations. Int J Pharm, 2015; 489(1-2): 158-169.
- 121. Manita D, Suyong K, Cheng G, Shayan F, Mingyu J, Yonghao M, Inyoung H, Hyungil J. Anti-obesity effect of a novel caffeine-loaded dissolving microneedle patch in high-fat diet-induced obese C57BL/6J mice J. Control. Release. 2017; 265: 41-47.
- 122. Hiraishi Y, Hirobe S, Iioka H, Quan Y, Kamiyama F, Asada H, Okada N, Nakagawa S. Development of a novel therapeutic approach using a retinoic acidloaded microneedle patch for seborrheic keratosis treatment and safety study in humans. J Control Release, 2013; 171: 93-103.
- 123. Imran Majid. Microneedling Therapy in Atrophic Facial Scars: An Objective Assessment. J Cutan Aesthet Surg, 2009; 2(1): 26-30.
- 124. Aust M, Knobloch K, Reimers K, Redeker J, Ipaktchi R, Ali Altintas M, Gohritz A, Schwaiger N, Vogt P. Percutaneous collagen induction therapy: An alternative treatment for burn scars. Burns, 2010; 36: 836-843.
- 125. Kim S, Park J, Jang Y, Son Y. Management of hypertrophic scar after burn wound using microneedling procedure (dermastamp). Burns, 2009; 35s: S1-S47.
- 126. Doddaballapur S. Microneedling with Dermaroller. J Cutan Aesthet Surg, 2009; 2(2): 110-111.
- 127. Harris A, Naidoo C, Murrell D. Skin needling as a treatment for acne scarring: An up-to-date review of the literature. Int J Women's Dermatology, 2015; 1:77-81.
- 128. Seo K, Kim D, Lee S, Yoon M, Lee H. Skin rejuvenation by microneedle fractional radiofrequency and a human stem cell conditioned

- medium in Asian skin: a randomized controlled investigator blinded split-face study. J Cosmet Laser Ther, 2013; 15(1):25-33.
- 129. Aust M, Reimers K, Repenning C, Stahl F, Jahn S, Guggenheim M, Schwaiger N, Gohritz A, Vogt P. Percutaneous collagen induction: minimally invasive skin rejuvenation without risk of hyperpigmentation-fact or fiction? Plast Reconstr Surg, 2008; 122(5): 1553-63.
- 130. Machekposhti S.A, Soltani M, Najafizadeh P, Ebrahimi S.A, Chen P. Biocompatible polymer microneedle for transdermal delivery of tranexamic acid. J. Control. Release, 2017; 261: 87-92.
- 131. Sachiko H, Risa O, Hiroshi I, Ying Q, Fumio K, Hideo A, Naoki O, Shinsaku N. Clinical study of a retinoic acid-loaded microneedle patch for seborrheic keratosis or senile lentigo. Life Sci, 2017; 168: 24-27.
- 132. Alsantali A. Alopecia areata: a new treatment plan. Clin Cosmet Investig Dermatol, 2011; 4: 107-115.
- 133. Chandrashekar B, Yepuri V, Mysore V. Alopecia Areata-Successful Outcome with Microneedling and Triamcinolone Acetonide. J Cutan Aesthet Surg, 2014; 7(1): 63-64.