

The Decrease in Farnesoid X Receptor, Pregnane X Receptor and Constitutive Androstane Receptor in the Liver after Intestinal Ischemia-Reperfusion

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Abstract - Purpose. Intestinal ischemia-reperfusion (I/R) damages remote organs, including the liver, and promotes multi-organ failure (MOF). However, the molecular mechanisms underlying acute liver injury after intestinal I/R have not been completely elucidated. Farnesoid X receptor (FXR), pregnane X receptor (PXR) and constitutive androstane receptor (CAR) regulate metabolizing enzymes and transporters, and coordinately prevent hepatotoxicity reflecting an inability of appropriate excretion of endogenous toxic compounds. In this study, we assessed FXR, PXR and CAR expression levels and their localization levels in nuclei in the liver after intestinal I/R. We also investigated the effect of IL-6 on FXR, PXR and CAR expression levels and their localization levels in nuclei in *in vitro* experiments. **Methods.** We used intestinal I/R model rats. Moreover, HepG2 cells were used in *in vitro* study. Real-time PCR and Western blotting were used to assess mRNA and protein expression levels. Nuclear receptor localization in nuclei was analyzed by Western blotting using nuclear extracts. **Results.** FXR and PXR expression levels began to be decreased at 3 h, and FXR, PXR and CAR expression levels were decreased at 6 h after intestinal I/R. The localization levels of FXR, PXR and CAR in nuclei began to be decreased at 3 h, and all of them were decreased at 6 h after intestinal I/R. In HepG2 cells, FXR, PXR and CAR expression levels were decreased by 0.5-1 ng/mL, 0.5-100 ng/mL and 100 ng/mL IL-6 treatment for 24 h, respectively. FXR, PXR and CAR localization levels in nuclei were suppressed by 0.5-10 ng/mL, 10-100 ng/mL and 10-100 ng/mL IL-6 treatment for 24 h, respectively. **Conclusions.** FXR, PXR and CAR expression levels are decreased in the liver after intestinal I/R. IL-6 is one of main causes the decreases in expressions of these receptors.

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INTRODUCTION

Intestinal ischemia-reperfusion (I/R) is a common clinical problem in small bowel transplantation, circulatory shock, and strangulation ileus. Intestinal I/R causes gut dysfunction, characterized histologically by mucosal injury, decreased basement membrane integrity, and decreased barrier function (1). Additionally, intestinal mucosa damage induces local production of inflammatory cytokines. Therefore, intestinal I/R damages remote organs,

including the lung (2), kidney (3) and liver (4), as well as the intestine, and promotes multi-organ failure (MOF) (5). In the liver, sinusoidal and venular endothelial cell damage causes loss of hepatic vascular permeability barrier function (4). In addition, the levels of aspartate transaminase (AST) and

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alanine transaminase (ALT) in serum are significantly increased after intestinal I/R (6). Injury of the intestinal mucosa is induced by production of reactive oxygen species (7) and release of inflammatory cytokines (8). However, the molecular mechanisms underlying acute liver injury after intestinal I/R have not been completely elucidated.

The liver is responsible for a great portion of the body's metabolic needs including the synthesis and control of pathway involved in cholesterol, fatty acid, carbohydrate, amino acid, bile acids and metabolism / detoxification of drugs and xenobiotics. Therefore, the liver failure directly contributes the poor prognosis in intestinal I/R patients. Transporters and metabolic enzymes are pivotal roles in the metabolism/detoxification of endogenous toxic compounds. Multi-drug resistance associated protein (MRP) 2 belongs to the ATP-binding cassette (ABC) transporter superfamily, and is abundant expression in the liver. MRP2 mainly effluxes organic anion compounds, such as bile acids, bilirubin, from liver to the outside of the body. In fact, Mrp2-deficient mouse is significantly increased the accumulation of organic anion compound in the liver (9).

Farnesoid X receptor (FXR), pregnane X receptor (PXR) and constitutive androstane receptor (CAR) are members of the nuclear receptor superfamily, and are highly expressed in the liver. These receptors regulate several metabolizing enzymes and transporters, such as MRP2 (10-14), and coordinately prevent hepatotoxicity (15, 16). Disruption of FXR or PXR in mouse developed severe hepatotoxicity reflecting an inability of appropriate excretion of endogenous toxic compounds, such as bile acids, bilirubin, and cholesterol (17, 18). Moreover, FXR, PXR or CAR agonists prevent hepatotoxicity inducing bile acids, cholesterol (19-21). The increase in MRP2 expression much contributes to the effects of PXR or CAR agonists on the hepatotoxicity induced by endogenous toxic compounds (21).

FXR, PXR and CAR expression levels are affected by inflammation. In fact, lipopolysaccharide (LPS) -induced inflammation is decreased the expression and function of nuclear receptors (22). LPS induces severe liver injury by the release of inflammatory cytokines including IL-6. Moreover, IL-6 mainly causes the decrease several metabolizing enzymes and transporters in LPS-induced inflammation (23). These findings suggest that IL-6 can affect the nuclear receptors expression, and

causes liver injury in LPS-induced inflammation.

We previously reported that MRP2 expression in the liver was significantly decreased after intestinal I/R (24). The decrease in MRP2 expression in the liver was associated with the increase in serum concentration of IL-6 (24). Therefore, we hypothesize that alterations of FXR, PXR and CAR expression levels cause liver injury after intestinal I/R, and IL-6 is important for the alteration of these nuclear receptors. In this study, we assessed FXR, PXR and CAR mRNA levels, their protein expression levels and their localization levels in nuclei in the liver after intestinal I/R. We also investigated the effect of IL-6 on FXR, PXR and CAR expression and function in *in vitro* experiments.

MATERIALS AND METHODS

Chemicals

IL-6 was purchased from Wako (Tokyo, Japan). [³H] Estrone-3-sulfate (E-3-S) (specific activity: 57.3 Ci/mmol) was purchased from Perkin Elmer (Boston, MA). All other reagents were the highest grade available and used without further purification.

Animals

Male Wistar rats, aged 6 weeks, were obtained from Jla (Tokyo, Japan). The rats were housed for at least 1 week at 23°C and 60±10% relative humidity, with a 12-h light/dark cycle (until reaching 250-350 g in weight). During the period of acclimatization, the rats were allowed free access to food and water. The experimental protocols were reviewed and approved by the Hokkaido University Animal Care Committee in accordance with the "Guide for the Care and Use of Laboratory Animals".

Intestinal I/R model

Surgical procedures were carried out as described in a previous report (24). The animals were anesthetized with sodium pentobarbital (30 mg/kg body weight, *i.p.* injection). Through a midline laparotomy, each rat was subjected to 30 min of ischemia by ligating small anastomosing vessels and occluding the superior mesenteric artery (SMA). Reperfusion was induced by removing the clamp. The abdomen was sutured during reperfusion. Rats were killed under surgical anesthesia at 1, 3, 6 and 24 h after reperfusion. Liver was harvested just before killing.

Cell culture

HepG2 cells were maintained in plastic culture flasks (Corning Costar Corp., Cambridge, MA). These stock cells were subcultivated before reaching confluence. HepG2 cells were kept in Dulbecco's modified Eagle's medium (Sigma Aldrich, Tokyo, Japan) with 10% fetal bovine serum (ICN Biomedicals, Inc., Aurora, OH) and 1% penicillin–streptomycin at 37°C under 5% CO₂–95% air at 37°C. The cells were given fresh growth medium every 2 days. When the HepG2 cells had reached confluence, they were harvested with 0.25 mM trypsin and 0.2% EDTA (0.5-1 min at 37°C), resuspended, and seeded into a new flask. In the present study, HepG2 cells were used between passages 137-145. IL-6 was dissolved in water and added to cells at various concentrations for periods of 24 or 48 h.

Semi-quantitative real-time PCR

Total RNA was prepared from liver of rats and HepG2 cells using an ISOGEN (Nippon Gene, Tokyo, Japan) and an RNase-Free DNase Set (QIAGEN, Tokyo, Japan). Single-strand cDNA was made from 2 µg total RNA by reverse transcription (RT) using a ReverTra Ace (TOYOBO, Osaka, Japan). Semi-quantitative real-time PCR was performed using an ABI PRISM 7700 Sequence Detector (Applied Biosystems, Foster City, CA) with Platinum[®] SYBR[®] Green qPCR SuperMix-UDG (Invitrogen, Carlsbad, CA) or an Mx3000[™] Real-time PCR System (STRATAGENE, Stratagene, La Jolla, CA) with GoTaq[®] qPCR Master Mix

(Promega, Madison, WI) as per the manufacturer's protocol, respectively. PCR was performed using rat FXR-specific primers, rat PXR-specific primers or human FXR-specific primers through 40 cycles of 95°C for 15 s, 50°C for 30 s and 72°C for 30 s; using rat CAR-specific primers through 40 cycles of 95°C for 15 s, 51°C for 30 s and 72°C for 30 s; using human CAR-specific primers through 40 cycles of 95°C for 15 s, 59°C for 5 or 10 s and 72°C for 30 s; using human MRP2-specific primers or human PXR-specific primers through 40 cycles of 94°C for 15 s, 60°C for 30 s and 72°C for 30 s; or using rat or human GAPDH-specific primers after pre-incubation at 95°C for 15 min. The primers specific to rFXR, rPXR, rCAR, rGAPDH, hMRP2, hFXR, hPXR, hCAR and hGAPDH were designed on the basis of sequences in the GenBank[™] database (accession no.: U18374, AF151377, AB104736, AF106860, NM_000392, BC130573, BC017304, BC121120 and NM_002046, respectively). The sequences of the specific primers are shown in Table 1. The PCR products were normalized to amplified GAPDH, which was the internal reference gene (housekeeping gene). Standard curves were prepared for each target and housekeeping gene. The standard curve was established between the threshold cycles (Ct) and the log₁₀(copy numbers) by using the Applied Biosystems sequence detection system software, version 1.9.1. The software calculates the relative amount of the target gene and the housekeeping gene based on the Ct.

Table 1 Primer sequences for real-time PCR

Name	Accession number		Sequences	Product size
rFXR	U18374	Forward	5'-GCAACT GCG TGATGGATATG-3'	226
		Reverse	5'-TTC TCC CTG CAT AGC TTG GT-3'	
rPXR	AF151377	Forward	5'-ATG TCT GAT GCC GCT GTG-3'	206
		Reverse	5'-TCACTG TGAAAC ACC GCA-3'	
rCAR	AB104736	Forward	5'-TGG CAT GAG GAAAGACAT GA-3'	190
		Reverse	5'-AGG CCT GAACTG CAC AAACT-3'	
rGAPDH	AF106860	Forward	5'-ATG GGAAGC TGG TCATCAAC-3'	221
		Reverse	5'-GTG GTT CAC ACC CAT CAC AA-3'	
hMRP2	NM_000392	Forward	5'-ACAGAG GCT GGT GGC AAC C-3'	227
		Reverse	5'-ACC ATT ACC TTG TCACTG TCC ATGA-3'	
hFXR	BC130573	Forward	5'-GCAGAT CAG ACC GTG AAT GA-3'	230
		Reverse	5'-GTT GCC ATT TCC GTG AAAAT-3'	
hPXR	BC017304	Forward	5'-GCAGGAGCAATT CGC CAT T-3'	102
		Reverse	5'-TCG GTG AGC ATAGCC ATG ATC-3'	
hCAR	BC121120	Forward	5'-GCAAGAGGAGAT GGC ACT G-3'	100
		Reverse	5'-AGC CAG CAG GCC TAG CAA C-3'	
hGAPDH	NM_002046	Forward	5'-AAG GTC ATC CCT GAG CTG AA-3'	95
		Reverse	5'-TTC TAG ACG GCAGGT CAG GT-3'	

Total protein extraction from rat liver

Rat liver was homogenized in cold lysis buffer containing 1.0% Triton-X100, 0.1% SDS and 4.5 M urea. The suspension was left to stand for 5 min and sonicated for 20 min at 4°C. Then it was centrifuged at 12,100×g for 15 min at 4°C.

Total protein extraction from HepG2 cells

HepG2 cells were suspended in 1 mL of PBS and centrifuged at 700×g for 5 min at 4 °C. The resulting pellet was suspended in lysis buffer. The suspension was left to stand for 5 min and sonicated for 20 min at 4°C. Then it was centrifuged at 12,100×g for 15 min at 4°C.

Nuclear extraction from rat liver

Nuclear extraction was carried out as described in a previous report with some modification (25). Rat liver was homogenized in solution 1 containing 0.6% Nonidet[®] P-40, 150 mM NaCl, 10 mM N-(2-hydroxyethyl) piperazine-*N'*-2-ethanesulfonic acid (HEPES) (pH 7.9), 1 mM EDTA and 0.5 mM phenylmethylsulfonyl fluoride (PMSF). The homogenate was allowed to stand for 5 min on ice and centrifuged at 2,100×g for 5 min at 4°C, and the pellet was suspended in 50 µL solution 2 containing 25% glycerol, 20 mM HEPES (pH 7.9), 420 mM NaCl, 1.2 mM MgCl₂, 0.2 mM EDTA, 0.5 mM dithiothreitol, 0.5 mM PMSF, 2 mM benzamide, 5 µg/mL pepstatin and leupeptin. The suspension was sonicated for 20 min at 4°C and centrifuged at 3,000×g for 15 s at room temperature.

Nuclear extraction from HepG2 cells

HepG2 cells were suspended in 100 µL solution 1. The suspension was allowed to stand for 5 min on ice and centrifuged at 2,100×g for 5 min at 4°C, and the pellet was suspended in 50 µL solution 2. The suspension was sonicated for 20 min at 4°C and centrifuged at 3,000×g for 15 s at room temperature.

Western blot analysis

Total protein extracts and nuclear extracts were prepared from rat liver or HepG2 cells. The protein concentrations of these samples in clear supernatant were determined by the method of Lowry et al. with bovine serum albumin as a standard (26). Each sample was denatured at 100°C for 3 min in a loading buffer containing 50 mM Tris-HCl, 2% SDS, 5% 2-mercaptoethanol, 10% glycerol, 0.002% BPB and

3.6 M urea and separated on 4.5% stacking and 10% SDS polyacrylamide gels. Proteins were transferred electrophoretically onto nitrocellulose membranes at 15 V for 90 min. The membranes were blocked with PBS containing 0.05% Tween 20 (PBS/T) and 10% non-fat dry milk for 1 h at room temperature. After being washed with PBS/T, the membranes were incubated overnight at room temperature with a mouse monoclonal antibody to MRP2 (Abcam, Cambridge, UK) (diluted 1:50 or 100), a rabbit monoclonal antibody to FXR (Santa Cruz Biotechnology, Santa Cruz, CA) (diluted 1:500), a rabbit monoclonal antibody to PXR (Santa Cruz Biotechnology, Santa Cruz, CA) (diluted 1:500) or a rabbit monoclonal antibody to CAR (Santa Cruz Biotechnology, Santa Cruz, CA) (diluted 1:250) and washed three times with PBS/T for 10 min each time. The membranes were subsequently incubated for 1 h at room temperature with horseradish peroxidase-conjugated goat anti-mouse secondary antibody (Santa Cruz Biotechnology, Santa Cruz, CA) or horseradish peroxidase-conjugated goat anti-rabbit secondary antibody (Santa Cruz Biotechnology, Santa Cruz, CA) at a dilution of 1:2,000 and washed three times with PBS/T for 10 min each time. The bands were visualized by enhanced chemiluminescence according to the instructions of the manufacturer (Amersham Biosciences Corp., Piscataway, NJ).

Measurement of IL-6 level in the liver

Measurement of IL-6 level in the liver was carried out as described in a previous report with some modification (27). For the measurement, the liver tissue was homogenized in PBS containing 2 mM PMSF, 5 µg/mL pepstatin and 5 µg/mL leupeptin. The homogenates were centrifuged at 10,000×g for 45 min at 4°C. The supernatants aliquoted and stored at -80°C. IL-6 was measured by ELISA with rat IL-6 kit (BioSource Int. Inc., CA, USA) as recommended.

Accumulation study

After removal of the growth medium, cells were washed with Hank's Buffered Salt Solution (HBSS) -HEPES (pH 7.4) buffer (25 mM D-glucose, 137 mM NaCl, 5.37 mM KCl, 0.3 mM Na₂HPO₄, 0.44 mM KH₂PO₄, 4.17 mM NaHCO₃, 1.26 mM CaCl₂, 0.8 mM MgSO₄ and 10 mM HEPES) and preincubated at 37°C for 10 min with 0.5 mL of HBSS-HEPES (pH7.4) buffer. Uptake was initiated by applying

HBSS-HEPES (pH 7.4) buffer containing 5 nM [^3H] E-3-S. The uptake study was performed at 37°C for 30 min. After a predetermined time period, uptake was terminated by suctioning off the applied solution and immersing the plates in ice-cold HBSS-HEPES (pH 7.4) buffer. To quantify the radioactivity of [^3H] E-3-S taken up by the cells, the cells were solubilized in 1% SDS/0.2 N NaOH. The remainder of the sample was mixed with 5 mL of scintillation cocktail (ASCI, Amersham Biosciences Corp., Piscataway, NJ, U.S.A.) to measure the radioactivity in a liquid scintillation counter (Packard, 1600TR). The cellular protein content was determined by the method of BCA protein assay with bovine serum albumin as a standard.

Data analysis

Statistical significance was evaluated using Dunnett's test (*in vitro* experiments except for accumulation study of the treatment of IL-6 for 24 h) or Duncan's test (accumulation study of the treatment of IL-6 for 24 h and *in vivo* experiments). A value of $p < 0.05$ was considered significant.

RESULTS

FXR, PXR and CAR expression after intestinal I/R

First, we assessed FXR, PXR and CAR expression after intestinal I/R in the liver. FXR mRNA level was decreased at 3-6 h after intestinal I/R (Fig. 1). PXR level mRNA was decreased at 3-24 h after intestinal I/R (Fig. 1). CAR mRNA level was decreased at 1-24 h after intestinal I/R. FXR protein expression level was 0.62-fold and 0.78-fold decreased at 3 and 6 h, respectively, and 1.6-fold increased at 24 h after intestinal I/R (Fig. 2). PXR protein expression level was 0.74-fold and 0.55-fold decreased at 3 and 6 h after intestinal I/R, respectively (Fig. 2). CAR protein expression level was 0.63- and 0.58-fold decreased at 6 and 24 h after intestinal I/R, respectively (Fig. 2). FXR level in nuclei was 0.75-fold decreased at 6 h after intestinal I/R and 1.4-fold increased at 24 h (Fig. 3). PXR level in nuclei was 0.61-fold decreased at 6 h after intestinal I/R (Fig. 3). CAR level in nuclei was 0.67- and 0.40-fold decreased at 6 and 24 h, respectively (Fig. 3). At 6 h after intestinal I/R, FXR, PXR and CAR mRNA levels, protein expression levels and localization levels in nuclei were decreased.

Amount of IL-6 in the liver after intestinal I/R

Serum concentration of IL-6 was increased at 3 (Supplementary figure 1) and 6 h (24) after intestinal I/R. However, it is not clarified whether production of IL-6 in the liver is increased. Thus, we measured the amount of IL-6 in the liver after intestinal I/R. The amount of IL-6 in the liver was significantly increased at 3 h, and that was highest at 6 h after intestinal I/R (Fig. 4). However, the increase of IL-6 in the liver at 6 h was not statistically significant because the amount of IL-6 in the liver was slightly increased at 6 h in sham-operated rats. The amount of IL-6 in the liver was recovered at 24 h after intestinal I/R (Fig. 4).

Effect of IL-6 on FXR, PXR and CAR expression

Next, we investigated whether IL-6 affected FXR, PXR and CAR expression. First, we confirmed that MRP2, which is a target gene of FXR, PXR and CAR, was affected by IL-6. MRP2 mRNA level (Fig. 5A) and protein expression level (Fig. 5B) were decreased by IL-6 treatment for 24 h. Moreover, the treatment of IL-6 for 24 h increased the accumulation of E-3-S, which is a typical substrate of MRP2. In the inhibited MRP2 function by MK 571, which is a typical inhibitor of MRP2, the effect of the treatment of IL-6 for 24 h on the accumulation of E-3-S was diminished. These results suggested that the increase in accumulation of E-3-S by IL-6 treatment for 24 h was caused by the decreased MRP2 expression. MRP2 expression and function were recovered by IL-6 treatment for 48 h (Fig. 5).

Next, we assessed the effect of IL-6 on FXR, PXR and CAR expression. The treatment of IL-6 for 24 h decreased FXR, PXR and CAR mRNA levels (Fig. 6). Although, FXR mRNA level was decreased in a concentration-dependent manner by IL-6 treatment for 24 h, FXR protein expression level was decreased by 0.5 or 1 ng/mL IL-6 treatment for 24 h, but was not altered by 10 or 100 ng/mL IL-6 treatment for 24 h (Fig. 7). PXR and CAR protein expression levels were decreased in a concentration-dependent manner by IL-6 treatment for 24 h (Fig. 7). FXR level in nuclei was decreased by 0.5-10 ng/mL IL-6 treatment for 24 h (Fig. 8). PXR and CAR levels in nuclei were decreased in nuclei was decreased in a concentration-dependent manner by IL-6 treatment for 24 h (Fig. 8).

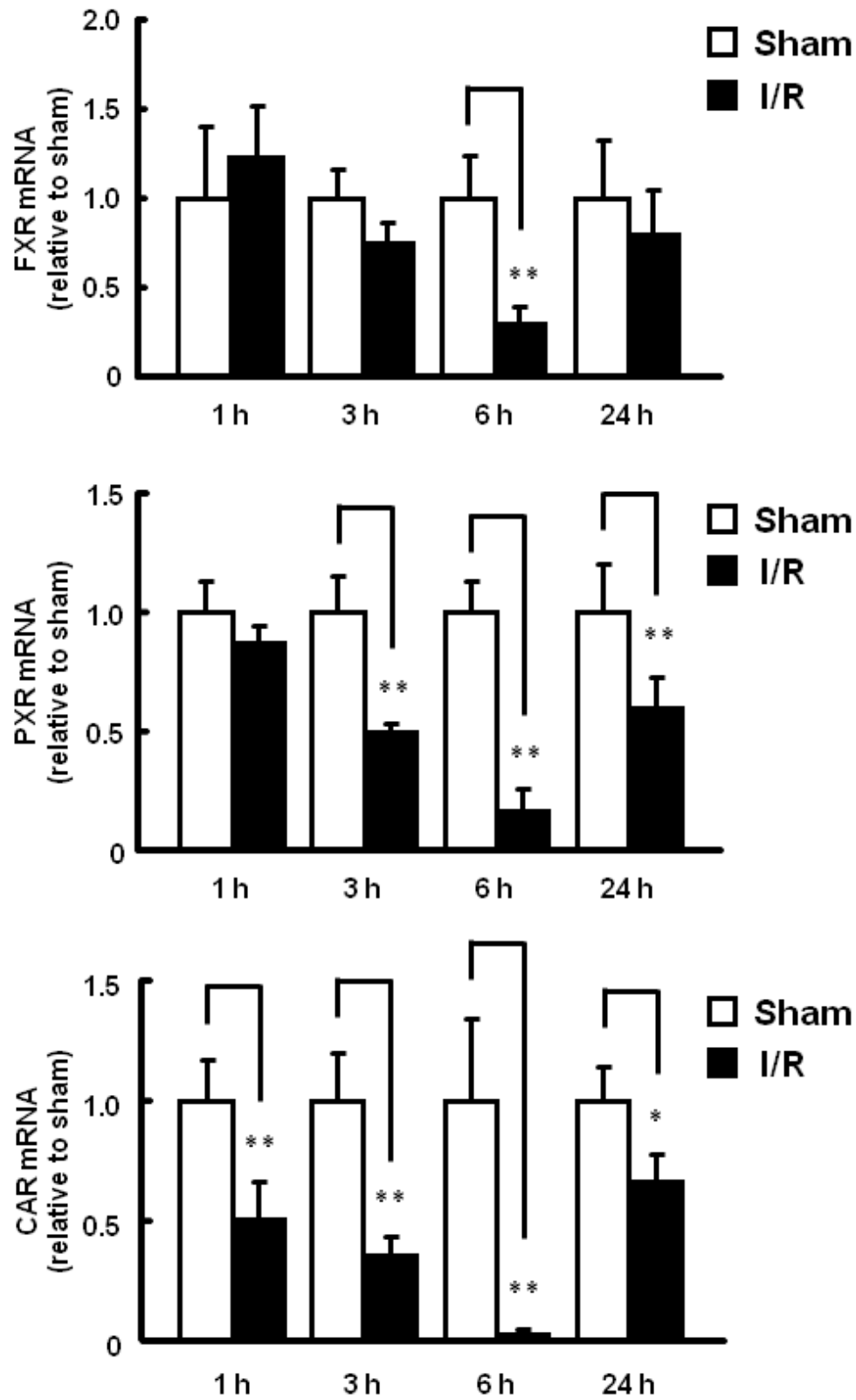


Figure 1. Time courses of FXR, PXR and CAR mRNA levels in the liver after intestinal I/R. Each column represents the mean with S.D. of 3-5 measurements. *, P<0.05 significantly different from sham, **, P<0.01.

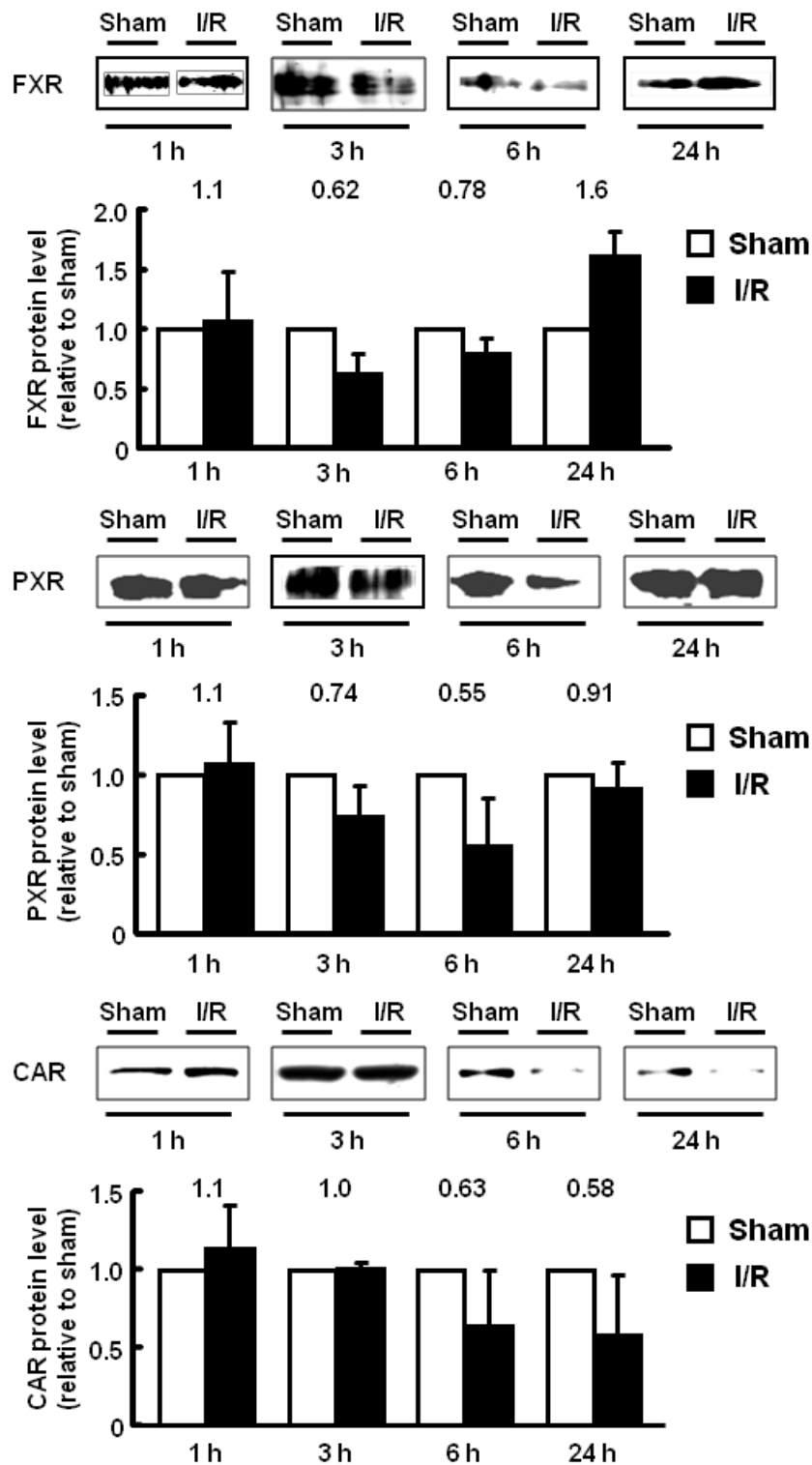


Figure. 2. Time courses of FXR, PXR and CAR protein expression in the liver after intestinal I/R. Whole cell lysates of the liver at 1, 3, 6 and 24 h after intestinal I/R were used for Western blot analysis. Western blot band intensity was determined by densitometry using Scion image program. Each column represents the mean with S.D. of 3-5 measurements.

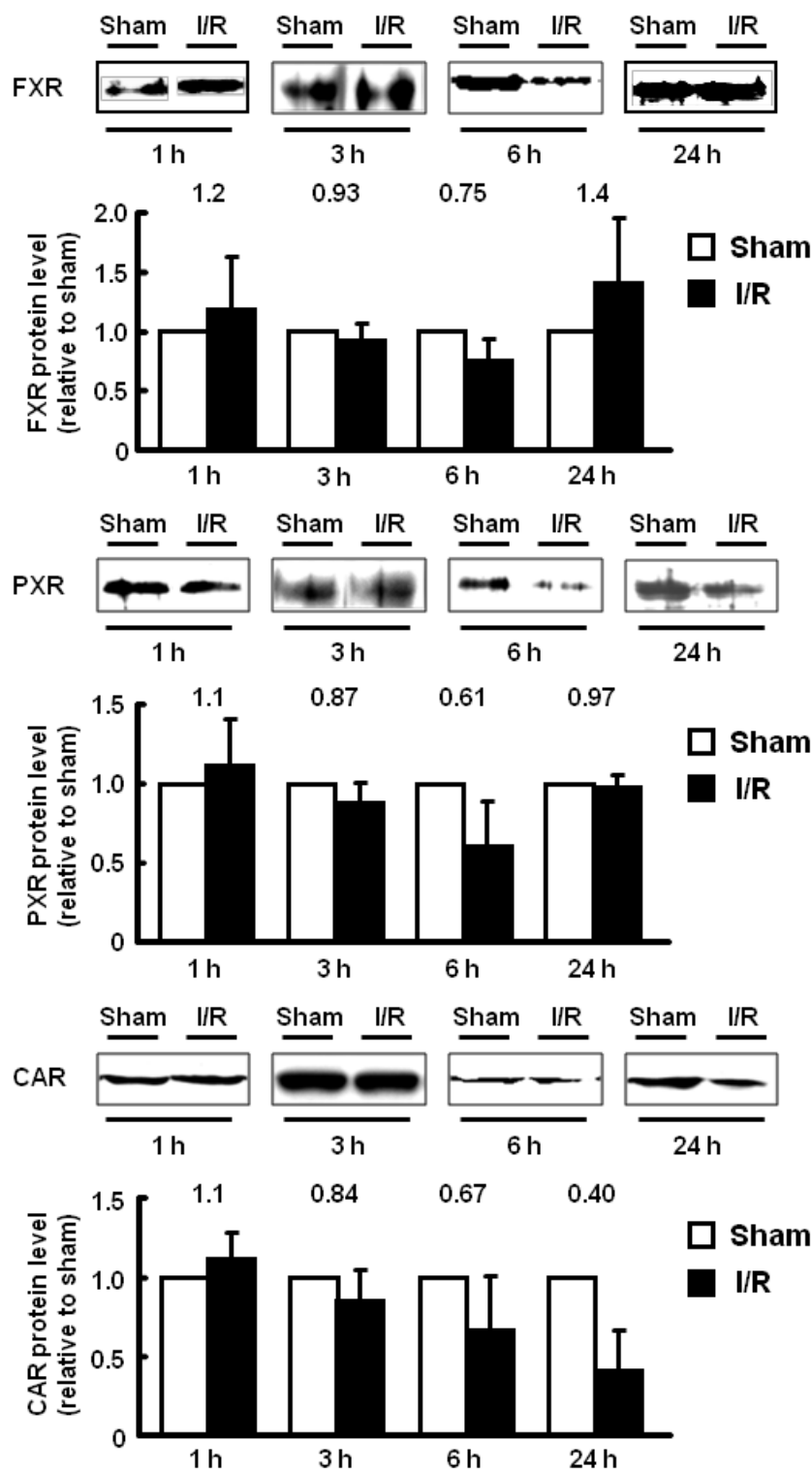


Figure 3. Time courses of FXR, PXR and CAR levels in nuclei in the liver after intestinal I/R. Nuclear extracts of the liver at 1, 3, 6 and 24 h after intestinal I/R were used for Western blot analysis. Western blot band intensity was determined by densitometry using Scion image program. Each column represents the mean with S.D. of 3-5 measurements.

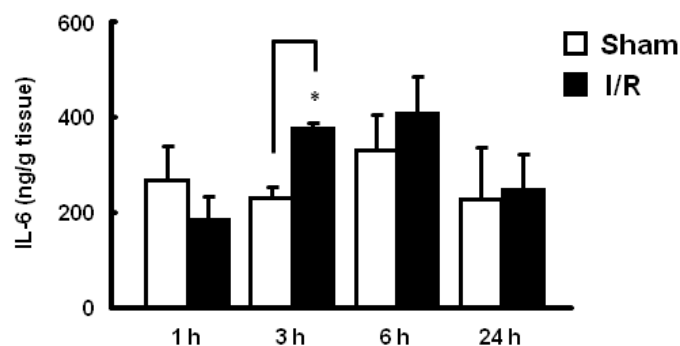


Figure 4. Time courses of amount of IL-6 in the liver homogenate after intestinal I/R. Each column represents the mean with S.D. of 3 measurements. *; $P < 0.05$ significantly different from sham.

Although PXR and CAR mRNA levels were still decreased by IL-6 treatment for 48 h (Fig. 6), FXR, PXR and CAR protein expression levels and the localization levels in nuclei were recovered by IL-6 treatment for 48 h (Figs. 7-8).

DISCUSSION

The purpose of this study was to determine the effect of intestinal I/R on FXR, PXR and CAR function in the liver. FXR, PXR and CAR are known to activate transcription of target genes in nuclei. Therefore, we investigated FXR, PXR and CAR expression levels and localization levels in nuclei.

FXR and PXR expression levels and localization levels in nuclei began to be decreased at 3 h, and all of FXR, PXR and CAR were decreased at 6 h (Figs. 1-3). These results were associated with MRP2 expression level during 1-6 h after intestinal I/R (supplementary figure 2 and previous report (24)), which is a target gene of FXR, PXR and CAR, indicating that intestinal I/R suppressed the activity of FXR, PXR and CAR. IL-6 is biosynthesized in the various cells such as macrophages, and then released from the intracellular to the extracellular space, where it exert its biological actions by binding to IL-6 receptor. At 3 and 6 h after intestinal I/R, the amount of IL-6 in the liver homogenates were increased by 376.32 and 408.01 ng/g tissue, respectively. As well as serum concentration of IL-6, the increase in the amount of IL-6 in the liver homogenate was associated with the decrease in the expression of MRP2, FXR, PXR and CAR during 1-6 h after intestinal I/R. These results suggested that IL-6 is one of main causes of the decrease in MRP2,

FXR, PXR and CAR expression levels. We previously reported the alteration of malondialdehyde (MDA) level, which is a marker of organ injury after I/R, in the liver (28). The increase in MDA level was associated with the increase in IL-6 and with the decrease in MRP2, FXR, PXR and CAR expression levels, suggesting that the decrease in FXR, PXR and CAR expression levels, at least in part, might be caused the liver injury after intestinal I/R. Furthermore, we demonstrated that FXR, PXR and CAR localization levels in nuclei were suppressed by 0.5-10 ng/mL, 10-100 ng/mL and 10-100 ng/mL IL-6 treatment for 24 h, respectively, and MRP2 expression level was decreased by 10-100 ng/mL IL-6 treatment for 24 h. These results also suggest IL-6, at least in part, participated with the suppression of FXR, PXR CAR activities and the decrease in MRP2 expression level. In HepG2 cells which were treated by IL-6 for 24 h, MRP2 expression level was not rigorously associated with FXR, PXR and CAR expression levels. However, one was decreased at least among FXR, PXR and CAR at the IL-6 concentration that decreased MRP2 expression level. Thus, we suggested that FXR, PXR and CAR coordinately regulate the MRP2 transcription. This suggestion is confirmed by the finding that FXR, PXR and CAR regulate several metabolizing enzymes and transporters, and coordinately prevent hepatotoxicity (15, 16).

On the other hand, the recovery of FXR, PXR and CAR expression levels during 6-24 h after intestinal I/R were not associated with the recovery of MRP2 expression level. At 24 h after intestinal I/R, MRP2 expression level was recovered (24).

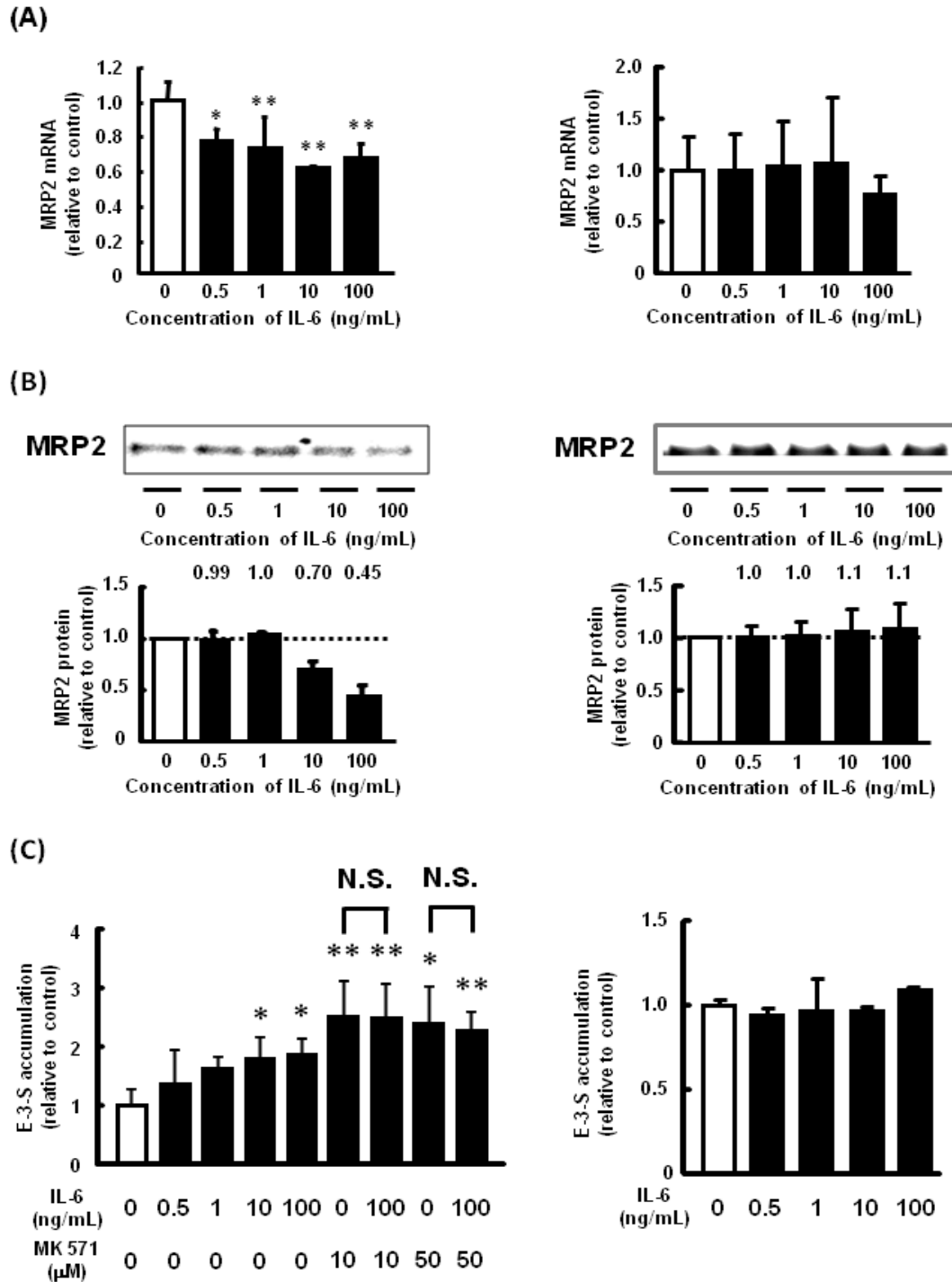


Figure 5. Effects of IL-6 on MRP2 mRNA level (A), protein expression (B) and E-3-S accumulation (C) in HepG2 cells. (A) Measurement of MRP2 mRNA level by real-time PCR. Each column represents the mean with S.D. of 3-7 measurements. (B) Analysis of MRP2 protein expression by Western blotting. Western blot band intensity was determined by densitometry using Scion image program. Each column represents the mean with S.D. of 3-4 measurements. (C) Analysis of E-3S accumulation. Each column represents the mean with S.D. of 3-4 measurements. HepG2 cells were exposed to 0-100 ng/mL IL-6 for 24 h (left side panels) or 48 h (right side panels). Control was incubated in a medium for 24 h (left side panels) or 48 h (right side panels). *, P<0.05 significantly different from the control, **, P<0.01, N.S.; not significant.

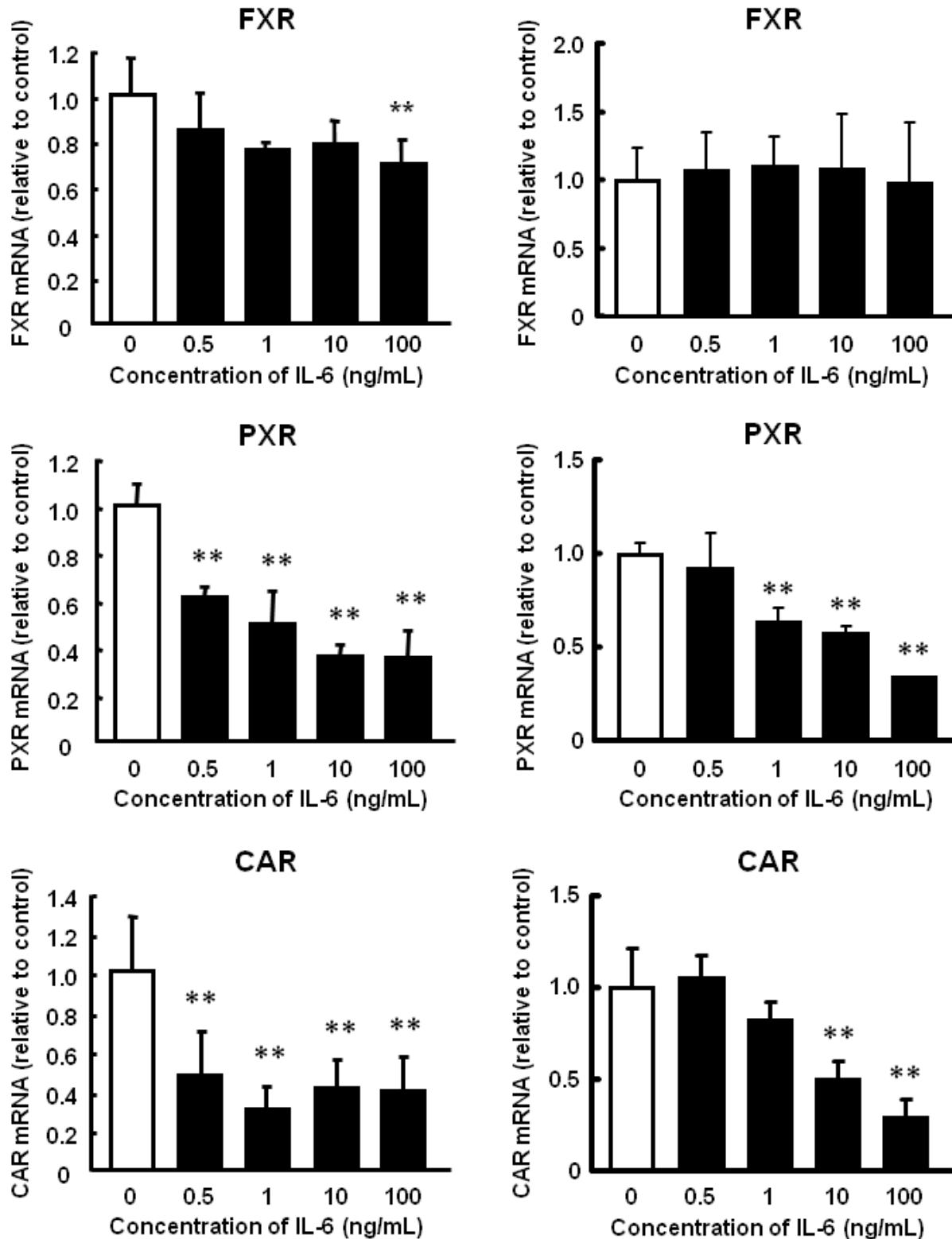


Figure 6. Effects of IL-6 on FXR, PXR and CAR mRNA levels in HepG2 cells. HepG2 cells were exposed to 0-100 ng/mL IL-6 for 24 h (left side panels) or 48 h (right side panels). Control was incubated in a medium for 24 h (left side panels) or 48 h (right side panels). Each column represents the mean with S.D. of 3-7 measurements. **, P<0.01 significantly different from the control.

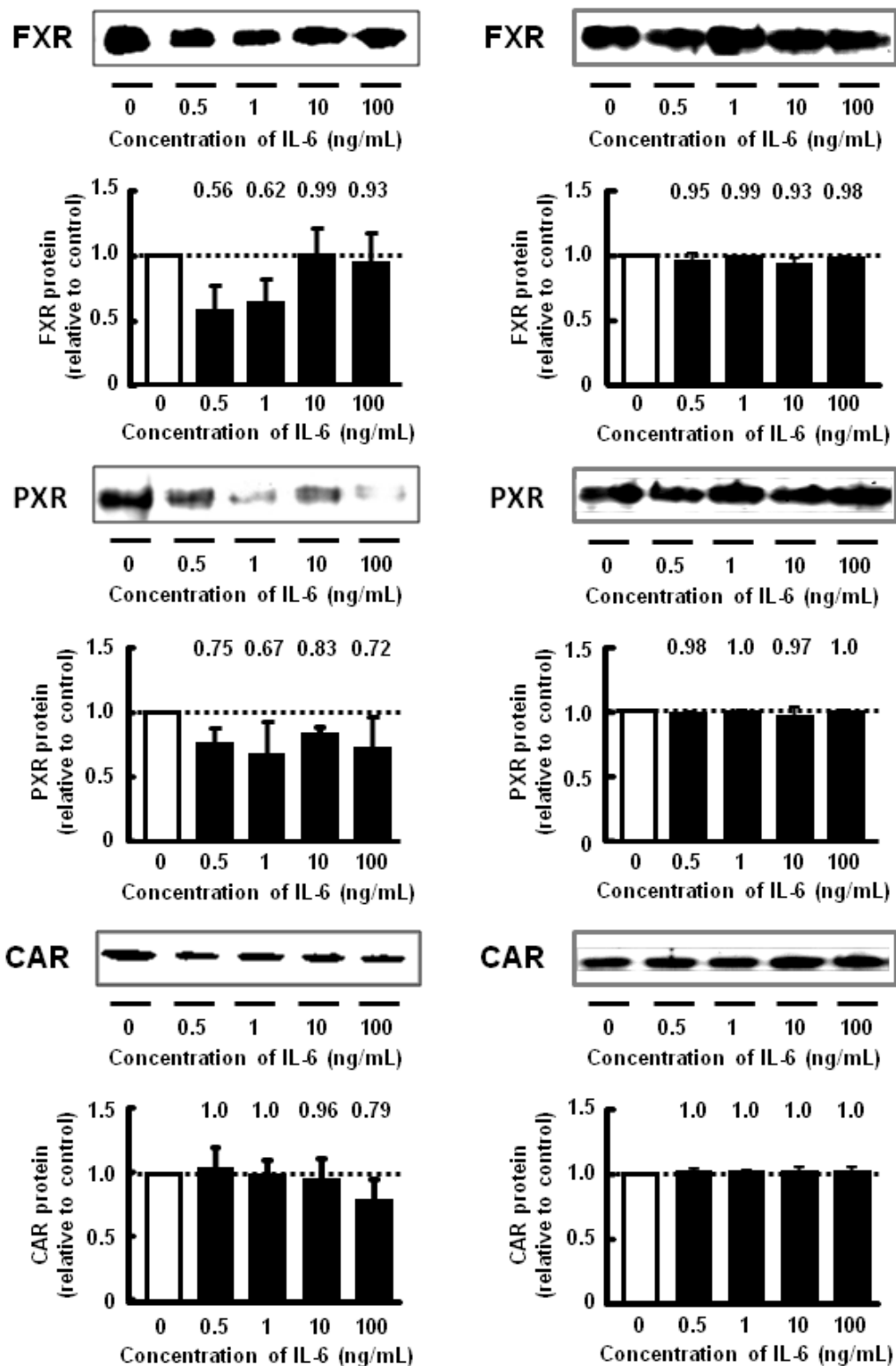


Figure 7. Effects of IL-6 on FXR, PXR and CAR protein expression in HepG2 cells. HepG2 cells were exposed to 0-100 ng/mL IL-6 for 24 h (left side panels) or 48 h (right side panels). Control was incubated in a medium for 24 h (left side panels) or 48 h (right side panels). HepG2 cell lysates used for Western blot analysis. Western blot band intensity was determined by densitometry using Scion image program. Each column represents the mean with S.D. of 3-4 measurements.

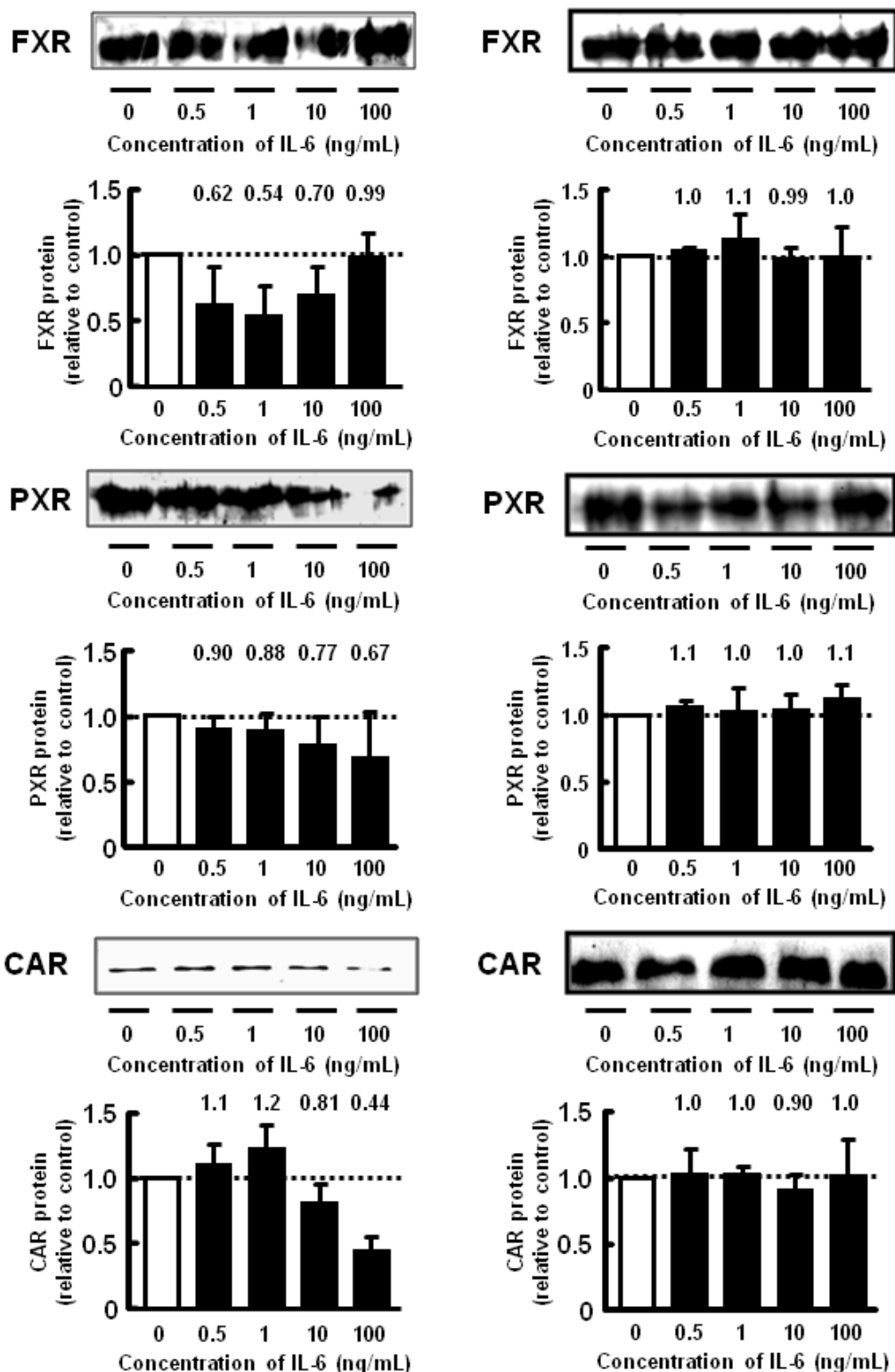
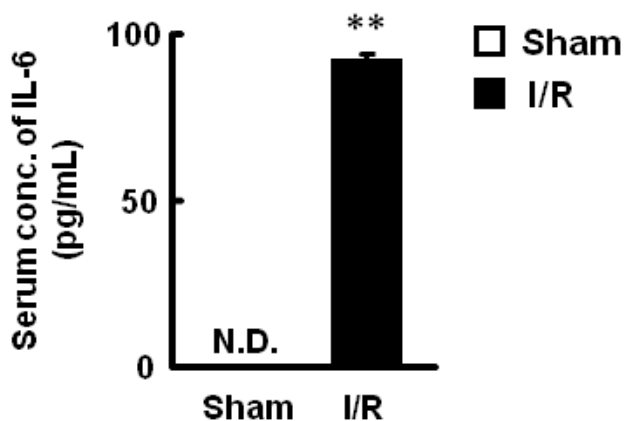
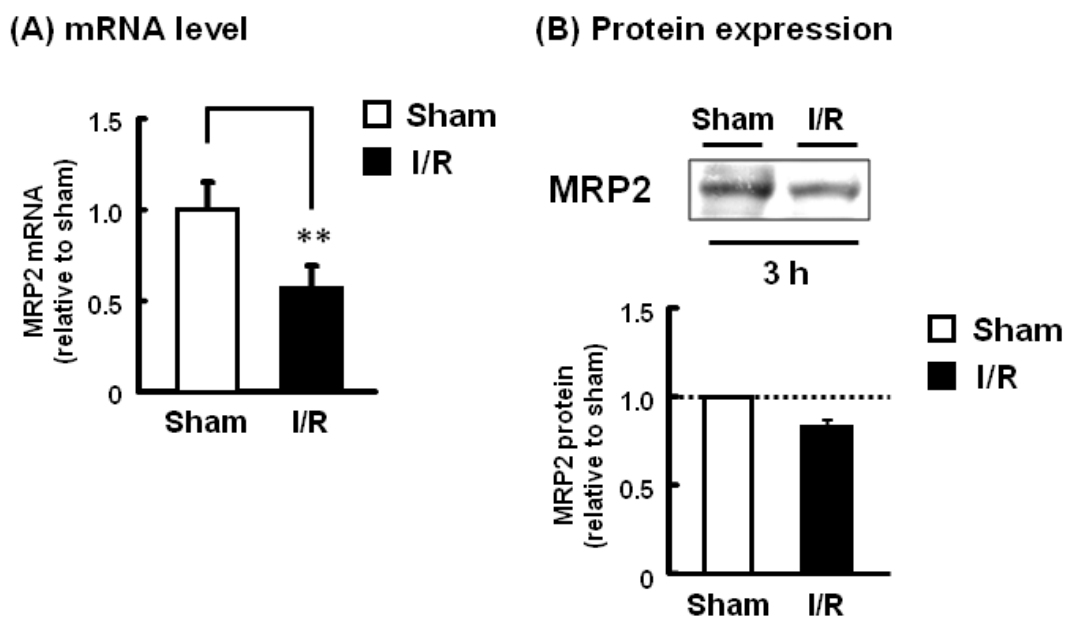


Figure 8. Effects of IL-6 on FXR, PXR and CAR levels in nuclei in HepG2 cells. HepG2 cells were exposed to 0-100 ng/mL IL-6 for 24 h (left side panels) or 48 h (right side panels). Control was incubated in a medium for 24 h (left side panels) or 48 h (right side panels). Nuclear extracts of HepG2 cells used for Western blot analysis. Western blot band intensity was determined by densitometry using Scion image program. Each column represents the mean with S.D. of 3-5 measurements.



Supplementary Figure 1. Serum concentration of IL-6 at 3 h after intestinal I/R. Each column represents the mean and S.D. of 3-4 measurements. Statistical significance was evaluated using Student's t-test. **: P<0.01 significantly different from sham. N.D.; not detect.



Supplementary Figure 2. MRP2 expression at 3 h after intestinal I/R. (A) MRP2 mRNA level at 3 h after intestinal I/R. (B) MRP2 protein expression at 3 h after intestinal I/R. Each column represents the mean with S.D. of 3-5 measurements. Liver crude membranes at 3 h after intestinal I/R were used for Western blot analysis. Western blot band intensity was determined by densitometry using Scion image program. Statistical significance was evaluated using Student's t-test. **: P<0.01 significantly different from sham.

FXR expression level was increased and PXR expression level was recovered, though CAR expression level was still decreased, suggesting that the recovery of MRP2 expression level after intestinal I/R was mainly caused by the increase in

FXR expression level. In the HepG2 cells, FXR expression level was not altered by high concentration IL-6 treatment for 24 h, though that was decreased by low concentration of IL-6 treatment for 24 h. These results also suggested that the

maintenances mechanism of FXR protein expression level and activity worked as cellular protection when sever inflammation occurred. This suggestion is also confirmed by the fact that intestinal I/R injury was recovered at 24 h (24, 27). The PXR protein expression level was recovered at 24 h after intestinal I/R, though PXR mRNA level was still decreased. In the HepG2 cells, PXR protein expression level was recovered with IL-6 treatment for 48 h, though PXR mRNA level was still decreased. These results suggested that PXR protein expression level was recovered by post-transcriptional regulation in IL-6-induced inflammation. The CAR mRNA level and protein expression level were still decreased at 24 h after intestinal I/R. On the other hand, CAR protein expression level was recovered with IL-6 treatment for 48 h in HepG2 cells, though CAR mRNA level was still decreased. Thus, we suggested that the decreased in CAR expression level after intestinal I/R was caused by IL-6 dependent and independent mechanisms.

The amount of IL-6 in the liver homogenate at 6 h in sham-operated rats was slightly increased than that at the other times in sham-operated rats. We did not investigate the factors inducing the IL-6 production by sham-operation, but it is possible that laparotomy may have contributed to this increment. Indeed, several workers reported that laparotomy induced the stress, and produced several stress hormones, particularly catecholamine and adrenocorticotropin (ACTH) (29, 30). These findings suggest that laparotomy may affect the increment of IL-6 by sham operation.

As in the liver, MRP2 expression level in the jejunum has also been shown to be decreased at 6 h after intestinal I/R (24). However, FXR, PXR and CAR expression levels and localization levels in nuclei in the jejunum were not decreased at any time after intestinal I/R (data not shown). These results suggested that decrease in MRP2 expression level after intestinal I/R was cause by different mechanism between the jejunum and liver. This difference between the jejunum and liver is not clear. It has been shown that IL-6 receptor expression level is abundant in the liver compared with that in the jejunum (31). This finding may be one of the reasons for the discrepancy between the jejunum and liver after intestinal I/R.

In conclusion, FXR, PXR and CAR expression levels are decreased in the liver after intestinal I/R.

IL-6 is one of main causes the decreases in expression levels of these receptors. The decrease in the expression levels of these nuclear receptors causes the decrease in target gene expression level, and might be cause the liver injury after intestinal I/R.

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